Screening of Endophytic Fungi of Melaleuca cajuputi Powell Leaves as Antibacterial Sources

by Hary Widjajanti

Submission date: 21-Sep-2022 12:38PM (UTC+0700)

Submission ID: 1905188285

File name: 38_Proceeding_IMC-SciMath_2022_compressed.pdf (346.86K)

Word count: 4852

Character count: 26674

Screening of Endophytic Fungi of *Melaleuca cajuputi* Powell Leaves as Antibacterial Sources

Hary Widjajanti, Salni, Niken Irfa Nastiti and Elisa Nurnawati

Department of Biology, Faculty of Mathematics and Natural Sciences, Sriwijaya University, Palembang, Indonesia

Keywords: Antibacterial, Endophytic fungi, Melaleuca cajuputi Powell.

Abstract:

Melaleuca cajuputi leaves contain high flavonoids, flavonoids are chemical compounds that are antibacterial. Antibacterial bioactive compounds are found in plants, but the use of plants as an antibacterial source requires a large amount of biomass. Endophytic fungi that live in plant tissue can produce secondary metabolites that potentially as an antibacterial compounds. Isolation of the endophytic fungi for antibacterial sources can reduce the large at 1 unt of plant as a source of antibacterial agents. The aims of this research were obtain endophytic fu 63 of M. cajuputi leaves as an antibacterial sources. The research stages were sampling, isolation and purificati 27 of endophytic fungi, cultivation and production of secondary metabolites, antibacterial activity and the minimum inhibitory concentration (MIC) tests, thin layer chromatography, and characterization and identification of endophytic fungi that potentially as an antibacterial sources. The results there were 7 isolates of endophytic fungi from Melaleuca cajuputi leaves. MC1, MC2, MC3, and MC4 fungal isolat 59 ave a strong antibacterial activities. The MIC of MC2 and MC3 extracts on Escherichia coli wei 41 00 µg/mL and 100 µg/mL. The MIC of MC1 and MC4 extracts on Staphylococcus aureus were 200 µg/mL and 700 µg/mL, respectively. The extract of four endophytic fungi contain phenol and flavonoid that potentially as an antibacterial. Endophytic fungi MC1 isolate identified as Scopulariopsis candida, MC2 isolate as Fusarium equiseti, MC3 isolate as Fusarium sporotrichoides, and MC4 isolate as Mucor hiemalis.

1 INTRODUCTION

A number of plants have been passed down from generation to generation used as medicine by almost every community, including in Indonesia. Plants that are used as medicinal plants, have now begun to be explored scientifically by analyzing the content of active compound that cause medicinal properties. The active compound in the plant are too few or minor, therefore if it is to be developed on a large scale there are constraints from the raw material of the plant that must be in large quantities.

Antibacterial compounds in the form of crude extracts produced from plants when purified, the amount will be even less. These obstacles make it very difficult if the development of bioactive compounds from medicinal plants that are rare and endemic will be carried out. This is because if rare plants are explored for bioactive compounds to be taken, then the sustainability of these plants is threatened. These constraints can be anticipated by isolating microbes, especially endophytic fungi that

are symbiotic with these plants. The technique used to take only a few parts of plants commonly used in traditional medicine, then the isolation of endophytic fungi from the plant parts is done, so that rare medicinal plants will remain sustainable and the development of medicinal compounds from these plants through endophytic fungi can be developed.

The development of medicinal plants based on medicinal plants must be considered aspects of preservation of medicinal plants. One technology 191 can be done was isolate endophytic fungi from parts of medicinal plants that are often used 191 traditional medicine and selection of bioactive of secondary metabolites such as antibacterial from the culture of endophytic fungi, thus the development of medicinal plants based on medicinal plants can be done and the preservation of medicinal plants especially that are already scarce 115 be maintained.

Endophytic fungi are fungi that live in plant tissues in a certain period and are able to live by forming colonies in plant tissue without endangering the host. Each higher level plant can contain several 1

endophytic microbes that produce secondary metabolites that are suspected as a result of coevolution or genetic recon 49 ation from host plants to endophytic microbes (Tan and Zou (2001), Stro12 and B.Daisy (2003).

The ability of endophytic fungi to produce secondary metabolites in accordance with their hold plants is a very large and reliable opportunity to produce secondary metabolites from endophytic microbes isolated from these host plants. Approximately 300,000 species of plants scattered on this earth, each plant contains one or more endophytic microbes consisting of bafferia and fungi (Strobel and Daisy, 2003). If 18e endophytes isolated from a medicinal plant can produce the same alkaloids or secondary metabolites as the original plants or even in higher amounts, then we don't need to cut down the original plants to be taken as simplicia, which is light to take decades to be harvested (Radji,2005). According to Stierlie et al. (1995) the use of endophytic microbes in producing active compounds has several advantages, including (1) faster producing of uniform quality, (2) can be produced on a large scale, and (3) the possibility of obtaining new bioactive components by providing conditions that are new different.

M. cajuputi can live on land that has limited nutrients from fertile soil and is rich in nutrients. This plant is also able to live in soil with critical conditions with little nutrient elements. M.cajuputi leaves in Indonesia are widely used as raw materials in the production of eucalyptus oil (Widiana et al, 2014). M.cajuputi leaves has many benefits, one of which can be used as an antibacterial source (Al-Abd et al., 2015). Melaleuca cajuputi has potential as an antibacterial, antioxidant, and even potentially 54 an antifilaria. (Al-Abd² et al., 2016). M.cajuputi has been used traditionally for the treatment of diseases such as coughing, stomach cramps, burns, and influenza. This plant also exhibits antiinflammatory. antidengue, anticancer anticonvulsant activities. The active compound of M.cajuputi leaves were trans caryophyllena, βselinene, germacrena (C15H26O), neopitadiene, cyclohexakarboksal dehide, β-caryophyllena, 3methoxy benzoic acid trimethylsilan; limonene; 1,4 naphthoquinone-5,8-dihydroxy-2-methoxy; carena; α-caryophylena; cineol; patchulin; ethyl benzene; benzene 1-metil-3 (metiletyl), and 1.3% volatile oil [10]. Essential oils are one of the compounds found in plants that have antibacterial activity (Ajizah, 2014).

M. cajuputi leaves also contain high flavonoids, flavonoids are chemical compounds that are

antibacterial (Al-Abd et al., 2015). The aims of the research were 1). Get endophytic ngi isolates from Melaleuca cajuputi Powell that are able to produce secondary metabolites that re antibacterial and antioxidant, 2). Conducting in vitro tests to verify the antibacterial and antioxidant properties of secondary metabolites of the plant's endophytic fungi of Melaleuca cajuputi Powell, 3). Identify the isolates of endophytic fungi of Melaleuca cajuputi Powell which have high potential to produce antibacterial.

2 MATERIALS AND METHOD

2.1 Isolation and Purification of Endophytic Fungi

The sterile leaves are cut 223 cm aseptically, then placed in a commercial Potato Dextrose Agar powder (PDA)(20 g dextrose, 15 agar, and 4 g potato starch) medium. Incubated at room temperature until the fungi appeared to grow. The different fungal colonies on PDA medium are further purified on new PDA medium.

2.2 Cultivation of Endophytic Fungi and Secondary Metabolite Extraction

Twenty agar plug of en 32 hytic fungi isolates inoculated into 500 mL Potato Dextrose Broth (PDB) medium and incubated at room temperature for ± 30 days. After a 62 nge in the color of the medium which indicates the production of secondary metabolites, the medium was 1 tracted using an ethyl acetate solvent (1: 1) and concentrated with a rotary evaporator to obtain a crude extract of secondary metabolites (Jamal et al., 2008).

2.3 Antibacterial Test

A 0.5 McFarland standard is prepared by mixing 0.05 mL of 1.175% barium chloride dihydrate (BaCl₂.2H₂O), with 9.95 mL of 1% sulfuric acid (H₂SO₄). The 0.5 Mc Farland solution measured the absorbance using a UV-Vis spetrophotometer (625 nm). The absorbance value obtained is equivalent to 1.5 28 bacterial cells/mL (Fatisa, 2013).

Escherichia coli, Staphylococcus aureus, Salmonella typhimurium, and Shigella dysentriae were inoculated into a physiological saline solution (0.85% NaCl) then homogenized and measured their

2

absorbance using a UV-Vis spectrophotometer (625 nm). The absorbance of the suspension test bacteria was prepared similar to the McFarland standard solution to reach the density of 1.5×10⁸ CFU/mL (Ku 31 a, 2014).

The antibacterial activity test was performed using disc diffusion method (Kirby-Bauer). The bacterial suspension was inoculated as much as 0.1 ml into a Mueller-Hinton Agar (MHA) medium with a spread plate methods. Crude extracts of secondary metabolites made concentrations of 400 µg/disc (4%) and standard antibiotic tetracycline 30 µg/disc (0.3%) as positive control. The disc paper saturated in a solution of secondary metabolite extract and tetracycline antibiotics then inoculated on the surface agar containing bacterial suspension and incubated atroom temperature for 24 hours (Islam et al., 2013). The criteria for each concentration of antibacterial compounds tested against standard antibiotics were determined by weak: A/B x 100% <50%; medium: 70% <A/B x 100%>50%; strong: A/Bx100% ≥70%, with A was clear zone diameter of extract and B was clear zone diameter standard antibiotic (Chan, 2007).

2.4 Determination of Minimum Inhibitory Concentration (MIC)

Secondary metabolite extracts that had strong antibacterial activity determined the minimum inhibitory concentrations. Determination of KHM using disc diffusion methods. The concentrations to be used were 4%, 3%, 2%, 1% and 0 (as a control). Paper discs that have been saturated with extracts of each of the existing concentrations and tetracycline as a standard antibiotics are placed on the surface of MHA medium that has been in 34 lated with the suspension of the test bacteria. Incubated at room temperature for 24 hours. Inhibition zones are measured and a minimum concentration value that can inhibit bacteria was determined.

2.5 Thin Layer Chromatography Analysis

48 lophytic fungal extracts which have high antibacterial activity then analyzed by TLC. Thin layer 52 matography analysis using silica gel 60 plates with ethyl acetate and n-hexane solvents with the eluent ratio used (ethyl acetate: n-hexane) were 1: 4; 3: 2; 2: 3 and 4: 1. The formed spots were seen by using UV light with a wavelength of 366 nm, and

the Rf value of the compound formed was determined (Jamal et al., 2008).

2.6 Characterization and Identification of Endophytic Fungi

Fungal endophytes were growth on Potato Dextrose Agar (PDA), Czapek Dox Agar (CDA), Czapek Yeast Extract Agar (CYEA), and Malt Extract Agar (MEA) media at room temperature (28°C) for 7 days. Fungal colonies grown in each medium observed about colony color, colony diameter, medium color around the colony, and colony reverse color (Srikandace, 2007). Microscopic morphology observed were hypha (septate or not), hyaline (colorless) or dark pigmented hyphae (greenish or blackish brown, dark black, grayish black), spores (asexual and sexual) with Henrici's Slide Cuture (HSC) methods (Gandjar et al., 1999).

3 RESULTS AND DISCUSSION

3.1 Antibacterial Activity of Secondary Metabolite of Endophytic Fungi

Isolation and purification of endophytic fungi of Melaleuca cajuputi Powell leaves obtained 7 isolates, 5 isolates from the you 27 leaves and 2 isolates from the old leaves. The antibacterial activity test of Melalleuca cajuputi endophytic fungi extract on Escherichia coli showed that there were two endophytic fungi isolates had the highest antibacterial activity, i.e MC3 and MC5 fungi isolates with 11 mm and 10 mm clear zone diameter. Five isolates did not have antibacterial activity on E coli namely MC1, MC2, MC4, MC6, and MC7 isolates. The antibacterial activity of a secondary metabolite extract varies with bacterial depending on the components of the bioactive

Extracts of secondary metabolites of endophytic fungi that have the highest antibacterial activity on *Staphylococcus aureus* were extract of MC1 and MC7 fungal isolates, with 10 mm and 11 mm clear zone diameter.

1

5 ble 1: Antibacterial activity of endophytic fungi of M cajuputi leaves against E coli, S aureus, S typhi, S. dysenteriae using the disc-diffusion methods.

Inclotes -		Zone of i	nhibition in mm (inhibition	%)
Isolates -	E coli	S aureus	S typhi	S dysenteriae
MC1	0	10(90.9)***	0	8,13(42.5)+
MC2	0	0	7(63.4)++	12.04(62.9)++
MC3	11(100)+++	9(81.8)+++	6,12(55.4)++	9.14(47.8)+
MC4	0	0	0	9,02(47.2)+
MC5	10(90.9)+++	9(81.8)+++	0	0
MC6	0	0	0	0
MC7	0	11(100)+++	7,2(65.2)++	8,08(42.3)+
Tetrasicline	11	11	11.04	19.12

Figures in parentheses are inhibition percentages compared to tetracyclin. Antibacterial activity is categorized as strong +++ for inhibition ≥70%, moderate ++ for inhibition 50<70%, and weak + for inhibition 50% (Chan et al., 2007).

The extract of MC3 and MC5 endophytic fungal isolates have antibacterial activity against E. coli with 11 mm and 10 mm clear zone diameter, also have antibacterial activity against S. aureus with both 9 mm clear zone diameter (Table 1). This means that the extract of second 24 metabolites of MC3 and MC5 isolates have a broad spectrum of antibacterial activity, because it has antibacterial activity against Gram-negative and Gram-positive bacteria. T26 shows the differences in the content of bioactive compounds found in endophytic fungi isolates that act as antibacterial. Research conducted by Ayepola and Adeniyi (2008), showed that secondary metabolites from leaf extracts of Eucalyptus cama 26 ensis including the plant family Myrtace 31 have a broad spectrum of antibacterial activity in inhibiting bacterial growth, both Gram positive and Gram negative bacteria.

3.2 Minimum Inhibitory Concentration (MIC) of Secondary Metabolite Extract of Endophytic Fungi

The MIC of secondary metabolite extract of endophytic fungi of *Melaleuca cajuputi* leaves on *Eschrichia coli*. MIC of MC₅ fungal isolates was 400 µg/mL with inhibition zone diameter 6,16 mm. The MIC of secondary metabolite extract of MC₃ endophytic fungal isolates was 100 µg / mL with an inhibition zone diameter 6.02 mm. Based on the MIC value, it showed that MC₃ and MC₃ endophytic fungi isolates had moderate antibacterial activity, when compared with Batos *et al.* (2016) where the *Eugenia calycina* plant extract has a MIC value of *E. coli* was 500 µg / mL, where the range of values of 500 µg / mL - 1000 µg / mL means that it is slightly

active, a value of 100 μg / mL - 500 μg / mL being, and KHM value less than 100 μg / mL means it is very active.

The MIC of the secondary metabolite extract of MC1 isolates on Staphylococcus aureus were 200 μg/mL with 6.1 mm inhibitory zone diameter of 6.1 mm. The M14 of MC7 endophytic fungi isolate was 700 μg/mL with a inhibition zone diameter of 6.02 mm. The concentration 700 µg/mL means that it is large enough to obtain a MIC value, compared 42 ith the results of a study conducted by Banhos et al. (2014) where extracts of the metabolites of the endophytic fungi of the Myrcia guianensis plant produced a MIC of S. aureus was 25 µg/mL. It is suspected that the bioactive compounds in MC7 fungal isolates are not strong enough to inhibit the growth of S. aureus at low concentrations, besides it is suspected that S. aureus bacteria have experienced resistance to bioactive compounds contained in MC7 fungal isolates. According to Vuong et al. (2015), S. aureus is one of the bacteria with a wide spread of antibiotic resistance. This bacterium has been resistant to penicillin, methicillin, even vancomycin, one of the antibiotics that are still commonly used.

3.3 Thin Layer Chromatography (TLC) Analysis

The results show that the color spot formed on the TLC plates for the three extracts of *Melaleuca cajuputi* endophytic fungal isolates which have relatively strong antibacterial activity, the isolates MC1, MC5, and MC7 were yellow. Based on the results of research conducted by Fadel *et al.* (2018), phenolic compounds when tested using thin layer chromatography plates will be yellow, blue or pink.

2

The color spot formed for MC3 isolate extract are yellow which shows phenol and brownish yellow compounds, showing flavonoid compounds, according to research conducted by Nugrahar legtyas et al., (2005) that flavonoid compounds have a brownish-yellow color to a red color. The color spot formed on the plates of each different endophytic fungi isolates showed the components of chemical compounds contained in each different endophytic fungious solates.

This is consistent with the statement of Sharma et al. (2016), different endophytic fungi in a plant can produce different secondary metabolites. Based on the result of Rf value it is known that there are endophytic fungi isolates that have the same Rf value also have the same color spots. This shows that the 16 ossibility of chemical compounds contained in the extract of secondary metabolites of endophytic fungi isolates may be the same. According to Ahamed et al. (2017), the Rf was a general characteristic value that can change depending on toolority of the mobile phase and its fixed phase. The Rf value indicates important information regarding the polarity of the chemical compounds to be identified. In addition, the Rf value can also be used in determining good solvents as a mobile phase in thin layer chromatography tests.

3.4 Characterization and Identification of Endophytic Fungi of *Melaleuca* cajuputi

The macroscopic and microscopic characteristics endophytic fungi isolates of *Melaleuca cajuputi* leaf are presented in Figure 1, 2 and Table 2. Microscopic observations of MC1 endophytic fungi isolates based on Figure 2 shows that hyphal fungi isolates of MC1 endophytes are septate and hyaline-colored, have short conidiophores and there are conidia-forming cells in the cylindrical conidiophore terminal, the conidia are rounded with smooth surfaces.

Based on the microscopic characteristics obtained it is suspected that MC1 endophytic fungi isolates belong to the type of *Scopulariopsis candida*. According to Gandjar *et al.* (1999), *S. candida* has a colony diameter of 3-4 cm within 7 days. Reverse color beige to light brown. Its conidiophores are short, have conidial-forming cells that are cylindrical in shape, conidia in the form of rounded to wide ovals, smooth-walled, and white to beige.

The macroscopic and microscopic characteristics endophytic fungi isolates of *Melaleuca cajuputi* leaf

are presented in Figure 3, 4 and Table 2. Based on microscopic characters of MC3 endophytic fungi isolates obtained that MC3 endophytic fungi isolates was identified as *Fusarium sporotrichiodes*, with characteristic hyphae and insular hyphae, there were two-branched conidia, oval-shaped, three-sided macroconidia, rounded microcondidia, and hyaluronic hyphae. According to Samson *et al.* (1995), *F. sporotrichiodes* has a white colony to yellow and finally brown. Microconidia is commonly found in aerial mycelia in the form of ovoid, piriform or fusoid, 3-5 macroconidias.

Based on macroscopic and microscopic characters of MC5 endophytic fungi isolates on Figure 5, 6, and Table 6. Type of hyphae: septate; hyphae color: hyaline; conidia color: hyaline; clamydospore color: brownish. Based on macroscopic and microscopic characters, MC5 endophytic fungi isolate was identified as Fusarium equiseti. According to Samson et al., (1995), Fusarium equiseti on PDA medium has a whitish color and becomes creamy to brown in color, has a lot of chlamydospores pale brown when old, smooth-walled or rough, in chains or clumps, in hyphae or conidia. The macroscopic and microscopic characteristics endophytic fungi MC7 isolates of Melaleuca cajuputi leaves were presented in Figure 7, 8 and Table 7.

Based on macroscopic and microscopic characters of MC7 endophytic fungi isolates based on Figure 7, 8 and Table 7 obtained results, MC7 endophytic fungi isolates had hyaline-colored hyphae and insulated hyphae, there were rounded and hyaline-colored columns, and had zigospores with rough surfaces. Based on the characteristics obtained, MC7 endophytic fungi isolate is suspected to be a type of *Mucor hiemalis*. The microscopic and macroscopic characteristics are in accordance with Gandjar (1999), *Mucor hiemalis* has a rather creamy yellow colony, a yellowish white reverse color. The columns are round and hyaline, the zigospores are semipurate or round in shape and the surface is rough.

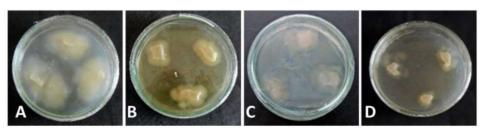


Figure 1: Colony images of MC1 endophytic fungal isolate of *Melaleuca cajuputi* on different medium. (a) PDA, (b) MEA, (c) CDA, (d) CYA.

Table 2: Colony characteristics of MC1 endophytic fungal isolate of Melaleuca cajuputi on different medium.

Characteristics	PDA medium	MEA medium	CDA medium	CYA medium
Colony color	White/ yellowish	Cream	Turbid white	Turbid white
Colony reverse	Turbid white/	Cream	Turbid white	Turbid white
color	yellowish			
Medium color	Turbid white	Turbid white	Turbid white	Turbid white
around colony				
Colony surface	Smooth	Smooth	Smooth	Smooth
Colony	4.75-5.7cm in 7 days	3.35-3.9cm in 7 days	3.05-3.25cm in 7 days	1.85-2.7cm in 7
diameter	•			days



Figure 2: Microscopical characteristics of MC1 isolate.

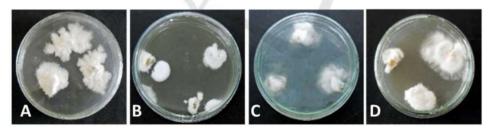


Figure 3: Colony images of MC3 endophytic fungal isolate of *Melaleuca cajuputi* on different medium. (a) PDA, (b) MEA, (c) CDA, (d) CYA.

Table 3: Colony characteristics of MC3 endophytic fungal isolate of Melaleuca cajuputi on different medium.

Characteristics	PDA medium	MEA medium	CDA medium	CYA medium
Colony color	White	White	White	White
Colony reverse color	White	Brownish white	White	White
Medium color around colony	Turbid white	Yellowish	Turbid white	Turbid white
Colony surface	Granular and notched	Powdery	Floccose	Floccose
Colony diameter	3.4-4.05 cm in 3 days	1.8-2.2cm in 7 days	3.4-4.05 cm in 3 days	2.35-5cm in 3 days



Figure 4: Microscopical characteristics of MC3 isolate.

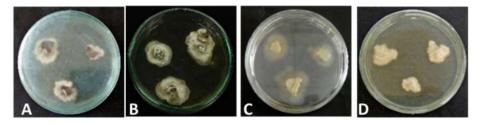


Figure 5: Colony images of MC5 endophytic fungal isolate of *Melaleuca cajuputi* on different medium. (a) PDA, (b) MEA, (c) CDA, (d) CYA.

Table 4: Colony characteristics of MC5 endophytic fungal isolate of Melaleuca cajuputi on different medium.

Characteristics	PDA medium	MEA medium	CDA medium	CYA medium
Colony color	Greys white	White	Yellowish turbid white	Cream
Colony reverse color	Brownish white	Brownish white	Yellowish turbid white	Cream
Medium color around colony	Yellowish	Yellowish	Turbid white	Turbid white
Colony surface	Powdery	Powdery	Smooth	Powdery
Colony diameter	1.8-2.5 cm in 7 days	2.95-3.75 cm in 7 days	3.15-3.95 cm in 7 days	1.75-2.35 cm in 7 days



Figure 6: Microscopical characteristics of MC5 isolate.

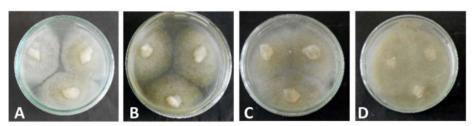


Figure 7: Colony images of MC7 endophytic fungal isolate of *Melaleuca cajuputi* on different medium. (a) PDA, (b) MEA, (c) CDA, (d) CYA.

Table 5: Colony characteristics of MC7 endophytic fungal isolate of Melaleuca cajuputi on different medium.

Characteristics	PDA medium	MEA medium	CDA medium	CYA medium
Colony color	Turbid white/yellowish	Turbid white/yellowish	Turbid white	Turbid white
Colony reverse color	White greyish brown	Turbid white/yellowish	Turbid white	Turbid white
Medium color around colony	Yellowish	Turbid white	Turbid white	Turbid white
Colony surface	Cottony	Cottony	Cottony	Cottony
Colony diameter	4.65-5.25 cm in 3 days	4.85-5.55 cm in 3 days	5-5.75 cm in 3 days	5.5-6.05 cm in 3 days

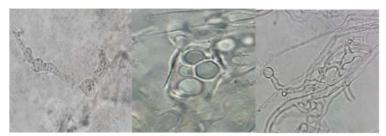


Figure 8: Microscopical characteristics of MC7 isolate.

CONCLUSION

Endophytic fungi obtained from Melaleuca cajuputi Powell obtained 7 isolates, 5 isolates obtained from young leaves, and 2 isolates from old leaves. Ethyl acetate extract of secondary metabolites endophytic fungi of Melaleuca cajuputi Powell which classified as strong and has a high percentage of antibacterial activity, namely isolates of en 35 hytic fungi MC1, MC3, MC5, and MC7 against Escherichia coli and Staphylococcus aureus. Minimum inhibitory concentration (MIC) of secondary metabolite extracts of MC5 and MC3 endophytic fungi isolates against Escherichia coli were 400 µg / mL and 100 μg / mL respectively, while secondary metabolite extracts of MC1 and MC7 endophytic fungi isolates against Staphylococcus aureus bacteria were 200 mg / mL and 700 mg / mL. MC1 fungi isolates were identified as Scopulariopsis candida, MC3 fungi identified as isolates were Fusarium sporotrichiodes, MC5 fungi isolates were identified as Fusarium equiseti, , and MC7 fungi isolates were identified as Mucor hiemalis.

REFERENCES

Ahamed, T., Rahman S.K.M., Shohael A.M. 2017. Thin Layer Chromatographic Profiling and Phytochemical Screening of Six Medicinal Plants in Bangladesh. International Journal of Biosciences. 11(1): 131 - Ajizah, A. 2004. Sensivitas Salmonella typhimmurium terhadap Ekstrak Daun *Psidium guajava* L. *Bioscie* 64 µe. 1(1): 31 – 38.

Al-Abd1, N.M., Zurainee Mohammed Nor, Marzida Mansor, Fadzly Azhar, M.S. Hasan dan Mustafa Kassim. 2015. Antioxidant, Antibacterial Activity, and Phytochemical Characterization of Melaleuca cajuputi extract. BMC Complementary and Alternative Medic 58 15:385 – 397.

Al-Abd2, N.M., Zurainee M.N., Marzida M., MS Hasan dan Mustafa K. 2016. Antifilarial and Antibiotic Activities of Methanolic Extracts of Melaleuca cajuputi flowers. Korean J Parasitol. 54(3): 273 -37 280.

Andries, J R., Paulina, N G., dan Aurelia, S. 2014. Uji Aktivitas Antibakteri Ekstrak Bunga Cengkeh terhadap Bakteri Streptococcus mutans secara In 39 Vitro. Jurnal e-Gigi. 2(2): 1-8.

Ayepola, O.O. dan Adeniyi B.A. 2008. The Antibacterial Activity of Leaf Extracts of Eucalyptus camaldulensis (Myrtaceae). Journal of Apllied Science Research. 22 4(11): 1410 – 1413.

Banhos, E.F., Souza A.Q.L., Andrade J.C., Souza A.D.L., Koolen H.H.F., dan Albuquerque P.M. 2014. Endophytic Fungi from *Myrcia guianensis* at The Brazilian Amazon : Distribution and Bioactivity. Brazilian Joi3 nal of Microbiology. 45(1): 153 – 161.

Bastos, R.G., Rosa C.P., Oliver J.C., Silva N.C., Dias A.L.T., Da Rocha C Q., Vilegas W., Da Silva G.A., dan Da Silva M.A. 2016. Chemical Characterization and Antimicrobial Activity of Hydroethanolic Crude Extract of Eugenia florida DC (Myrtaceae) Leaves. International Jurnal of Pharmacy and Pharmaceutical Sciences, 8(6): 110 - 115.

Chan, E W C., Lim, Y Y., dan Mohammed, O. 2007. Antioxidant and Antibacterial Activity of Leaves of

- 2
- Etlingera Species (Zingiberaceae) in Peninsular Malaysia. Food Chemistry. 104. 1586-1593.
- Fadel, O., Rodrigues D.G., Girard L., Bauduin P., Castera A.R., L'Hermitte A., Gaillard J.C., dan Diat O. 2018. Seperation and Identification of Polar Polyphenols in Oily Formulation Using High Performance Thin Layer Chromatography and Mass Spectroscopy Techniques. 11 Oilseeds & Fats Corps and Lipids. 25(5): 1 – 8.
- Fatisa, Y.2013.Daya antibakteri ekstrak kulit dan biji buah pulsa terhadap *Staphylococcus aureus* dan *Escherichia* coli secara in vitro. Jurnal Peternakan (10): 31-38.
- Islam, M R., Shahnaj, P., Rikta, B., Nusrat, J., Nandita, D., dan Ekramul, I. 2013. Antibacterial and Phytochemical Screening of Ethanol Extracts of Manilkara zapota Leaves and Bark. International Journal of Pharma Sciences. 3(6): 394-397.
- Jamal, Y., Muhammad, I., Atit, K., dan Andria, A. 2008.
 Diversitas dan Profil Metabolit Sekunder Jamur Endofit yang Diisolasi dari Tumbuhan Gambir (*Uncaria gambler*) Serta Aktivitas Biologisnya
 Sebagai Antibakteri. *Berita Biologi*. 9 (1). 149-154.
- Kumala, S. 2014. Mikroba Endofit: Pemanfaatan Mikroba Endofit dalam Bidang Farmasi. Jakarta: PT. ISFI
- 45 Penerbitan. v + 105 hlm.
 Nugrahaningtyas, K.D., Matsjeh S., dan Wahyuni T.D.
 2005. Isolasi dan Identifikasi Senyawa Flavonoid dalam Rimpang Temu Ireng (Curcuma aeruginosa
 Roxb.). Biofarmasi. 3(1): 32 38.
- Radji, M. 2005. Peranan Bioteknologi dan Mikroba Endofit dalam Pengembangan Obat Herbal. Majalah Ilmu Kefarmasian 2 (3):113-126.
- Samson, R A., Ellen, S H., Jens, C F., dan Ole, F. 1995.
 Introduction to Food-Borne Fungi Fourth Edition.
 Netherlands: Centraalbureau voor Schimmelcultures. 1
 46 + 322 hlm.
- Sharma, D., Pramanik A., Agrawal P.K. 2016. Evaluation of Bioactive Secondary Metabolites from Endophytic Fungus Pestalotiopsis neglecta BAB-5510 Isolated from Leaves Cupressus torulosa D.Don. Biotech. 6(1)
- Shibuya, H., Agusta, A., Ohashi, K., Maehara, S., dan Simanjuntak, P. 2005. Biooxidation of (+)-Catechin and (-)-Epicatechin into 3,4-Dihydroxy Flavan Derivatives by The Endophytic Fungus Diaporthe sp. Isolated from A Tea Plant. Chem Pharm Bull. 53(7): 866-867.
- 866-867. 9
 Srikandace, Y., Yatri, H., dan Partomuan, S. 2007. Seleksi Mikroba Endofit Curcuma zedoria dalam Memproduksi Senyawa Kimia Antimikroba. *Jurnal Ilmu Kefarmasian Indonesia*. 5 (2): 77-84
- Stierle, A.,D.Stierle, G.Strobel, G.Bignami, and P.Grothaus. 1995. Bioactive metabolites of the endophytic fungi of pacific yew *Taxus brevifolia*. Elsevier Scientific Publ.,Ireland.
- Strobel, GA, and B.Daisy (2003). Bioprospecting for microbial endophytes and their natural products. *Microbiol. and Mol. Biology Rev* 67(4):491-502.
- Tan, RX and WX.Zou.2001. Endophytes: a rich source of functional metabolites. Nat Prod.Rep. 18:448-459.

- 53
- Widiana, A., Taufikurahman, Limin S.H., Hernaman I. dan Manurung R. 2014. The Potential of Gelam Leaves (Melaleuca cajuputi Powell) as Cattle Feed.
 Pakistan Journal of Nutrition. 13(6): 348 350.
- Visagie, C M., Houbraken J., Jens, C F., Hong, S B., Klaassen, C H W., Perrone, G., Seifert, K A., Varga J., Yaguchi, T., dan Robert, A S. 2014. Identification and Nomenclature of The Genus Penicillium. Studies In 10 Mycology. 78: 343-371.
- Vuong, C., Yeh A.J., Cheng G.Y.C., dan Otto M. 2015. Investigational Drugs to Treat Metichilin-resistant Staphylococcus aureus. Expert Opinion 14 Investigational Drug. 25(1): 73 – 93.
- Zhao, J., Zhou, L., Wang, J., Shan, T., Zhong, L., Liu, X., dan Gao, X. 2010. Endophytic Fungi for Producing Bioactive Compounds Originally from Their Host Plants. Formatex. 567-576

Screening of Endophytic Fungi of Melaleuca cajuputi Powell Leaves as Antibacterial Sources

ORIGINALITY REPORT

23% SIMILARITY INDEX

18%
INTERNET SOURCES

15%

PUBLICATIONS

8%

STUDENT PAPERS

PRIMARY SOURCES

sciencetechindonesia.com

4%

2 icime.usu.ac.id

1 %

Purbowatiningrum Ria Sarjono, Leni Diah Putri, Chlara Eka Budiarti, Nies Suci Mulyani et al. "Antioxidant and antibacterial activities of secondary metabolite endophytic bacteria from papaya leaf ()", IOP Conference Series:

Materials Science and Engineering, 2019

1 %

Submitted to Universitas Sumatera Utara
Student Paper

1 %

Chan, E.W.C.. "Antioxidant and antibacterial activity of leaves of Etlingera species (Zingiberaceae) in Peninsular Malaysia", Food Chemistry, 2007

%

Publication

7	waset.org Internet Source	1 %
8	bennsethicoa.com Internet Source	1 %
9	repository.ub.ac.id Internet Source	1 %
10	Submitted to University of New England Student Paper	<1 %
11	pdfs.semanticscholar.org Internet Source	<1 %
12	irjponline.com Internet Source	<1%
13	coek.info Internet Source	<1%
14	digilib.unimed.ac.id Internet Source	<1%
15	www.ejournal.warmadewa.ac.id Internet Source	<1%
16	"Microbial Interventions in Agriculture and Environment", Springer Science and Business Media LLC, 2019 Publication	<1%
17	etheses.uin-malang.ac.id Internet Source	<1%

18	link.springer.com Internet Source	<1%
19	www.science.gov Internet Source	<1%
20	Sogandi, P Nilasari. "Isolation and molecular identification of Endophytic bacteria from Noni fruits (Morinda citrifolia I.) and their antibacterial activity", IOP Conference Series: Earth and Environmental Science, 2019 Publication	<1%
21	simakip.uhamka.ac.id Internet Source	<1%
22	tede.ufam.edu.br Internet Source	<1%
23	Kapila K. Liyanage, Sehroon Khan, Siraprapa Brooks, Peter E. Mortimer, Samantha C. Karunarathna, Jianchu Xu, Kevin D. Hyde. "Morpho-Molecular Characterization of Two Ampelomyces spp. (Pleosporales) Strains Mycoparasites of Powdery Mildew of Hevea brasiliensis", Frontiers in Microbiology, 2018 Publication	<1%
24	www.mdpi.com Internet Source	<1%
25	doaj.org Internet Source	<1%

26	"Advances in Endophytic Fungal Research", Springer Nature, 2019 Publication	<1%
27	Generoso da Silva GONÇALVES Fernando, Conceição de ARAÚJO Jonatas, da Silva BRASIL Dionízio, Asfury RODRIGUES Rodrigo et al. "Chromatography and bioautography of endophytic fungi extracts of Uncaria tomentosa (Willd.) DC with antibacterial activity", Journal of Medicinal Plants Research, 2019 Publication	<1%
28	ugspace.ug.edu.gh Internet Source	<1%
29	univendspace.univen.ac.za Internet Source	<1%
30	Hairil Fiqri, Adzani Gaisani Arda, Khodijah Adrebi, Wahyu Diah Proborini, Rahma Micho Widyanto. "Evaluation of Total Phenolic, Total Flavonoid, and In Vitro Cytotoxic Activity of Syzygium cumini Extract in Cervical Cancer Cell", Journal of Physics: Conference Series, 2020 Publication	<1%
31	cdn.intechopen.com Internet Source	<1%

_	32	Internet Source	<1%
_	33	pubs.rsc.org Internet Source	<1%
	34	www.hilarispublisher.com Internet Source	<1%
_	35	www.researchgate.net Internet Source	<1%
	36	V Sutanti, D Fuadiyah, LH Hidayat, T Agnizarridlo, K S Anggiarta. "Analysis of the effect of extracted yellow kepok banana peels () on the size and morphology of ", Journal of Physics: Conference Series, 2020 Publication	<1%
	37	ejurnal.stikes-bth.ac.id Internet Source	<1%
			• 70
	38	scitepress.org Internet Source	<1%
_	38		<1 _%

41	Warley de Borges, Keyller Borges, Pierina Bonato, Suraia Said, Monica Pupo. "Endophytic Fungi: Natural Products, Enzymes and Biotransformation Reactions", Current Organic Chemistry, 2009	<1%
42	academicjournals.org Internet Source	<1%
43	biomedpharmajournal.org Internet Source	<1%
44	dergipark.org.tr Internet Source	<1%
45	e-journal.uajy.ac.id Internet Source	<1%
46	ujcontent.uj.ac.za Internet Source	<1%
47	www.journalmrji.com Internet Source	<1%
48	www.smujo.id Internet Source	<1%
49	Advances in Endophytic Research, 2014. Publication	<1%
50	Submitted to B.S. Abdur Rahman University Student Paper	<1%

51	Kangle Niu, Zhengyao Liu, Yuhui Feng, Tianlong Gao, Zhenzhen Wang, Piaopiao Zhang, Zhiqiang Du, Daming Gao, Xu Fang. "Efficient Synthesis of Rare Disaccharides by Engineered β-Glucosidase Based on Hydropathy Index", American Chemical Society (ACS), 2020 Publication	<1%
52	repository.uinjkt.ac.id Internet Source	<1%
53	sith.itb.ac.id Internet Source	<1%
54	www.hindawi.com Internet Source	<1%
55	Akhmad Endang Zainal Hasan, Dimas Andrianto, Husnawati, Faisal Rahman, Nurliani Bermawie, Heddy Julistiono, Eny Ida Riyanti. "ISOLATION AND ANTI-YEAST ACTIVITY OF SECONDARY METABOLITES OF SOURSOP LEAF ENDOPHYTIC FUNGI", Indonesian Journal of Applied Research (IJAR), 2022 Publication	<1%
56	Moraa Obare Ruth, A. Indieka Stephen, Matasyoh Josphat. "Antibacterial activity of endophytic fungi isolated from leaves of medicinal Plant Leucas martinicensis L.	<1%

growing in a Kenyan tropical forest", African Journal of Biochemistry Research, 2020

Publication

57	Ramesha Alurappa, Srinivas Chowdappa. "Antimicrobial Activity and Phytochemical Analysis of Endophytic Fungal Extracts Isolated from Ethno-Pharmaceutical Plant Rauwolfia tetraphylla L.", Journal of Pure and Applied Microbiology, 2018 Publication	<1%
58	eprints.um.edu.my Internet Source	<1%
59	www.ncbi.nlm.nih.gov Internet Source	<1%
60	"Fungi and their Role in Sustainable Development: Current Perspectives", Springer Science and Business Media LLC, 2018 Publication	<1%
61	A. Shah, M.A. Rather, Q.P. Hassan, M.A. Aga, S. Mushtaq, A.M. Shah, A. Hussain, S.A. Baba, Z. Ahmad. "Discovery of anti-microbial and anti-tubercular molecules from: an endophyte of ", Journal of Applied Microbiology, 2017 Publication	<1%

Microbial Diversity and Biotechnology in Food Security, 2014.

<1%



<1_%

www.isca.in 64 Internet Source

Exclude quotes On Exclude bibliography

Exclude matches

Off