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RESEARCH ARTICLE

Immunostimulatory Activity of Ethanol Extract from *Calotropis gigantea* L. Flower in Rats against *Salmonella typhimurium* Infection

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ABSTRACT:

Calotropis gigantea L. flower contains a high value of flavonoid and polyphenol that potentially as an immunostimulatory agent. This research has been conducted to observe and acquire the immunostimulator effect of ethanol extract from *Calotropis gigantea* L. flower (EECGF) in rats, which were induced with *Salmonella typhimurium* (*S. typhi*). Rats were grouped into six, namely normal, negative control, positive control, EECGF dose 40mg/200g, 80mg/200g, and 120mg/200g body weight. Rats were treated for 12 days according to their respective groups, on the sixth day, rats were induced by *S. typhi* except for the normal group. Three days after infection, the Widal test was evaluated, and on the 12th day, an immune system parameter was evaluated. The results obtained, EECGF can increase the value of leukocytes, lymphocytes, monocytes, and neutrophils, followed by an increase in CD₄ T cell values proportionally due to *S. typhi* infection. It was concluded that EECGF had immunostimulatory activity, EECGF dose 80mg/200g had the same character with positive control. However, the highest dose of extract 120mg/200g can cause splenomegaly.

KEYWORDS: *Calotropis gigantea* L., immunostimulator, CD₄ T cell, leukocytes, *Salmonella typhimurium*, chemometrics.

INTRODUCTION:

Humans have the risk of being infected with various types of microorganisms from food with a low level of hygiene. In contrast to diabetes, which is more on metabolic function problems¹. Microorganism infections can be transmitted through food, for example, Salmonella typhimurium (S. typhi), which can cause typhoid fever^{2,3,4}. Infectious agents such as S. typhi that the body can inhibit the function of enter phagolysosomes so that these bacteria are difficult to remove⁵. This bacterial infection can cause splenomegaly one week after entering the blood circulation. S. typhi will activate CD₄ T cells by specific through primary system mechanisms immune histocompatibility complex class 2 (MHC-II) and facilitate macrophages to carry out phagocytosis^{6,7}.

Therefore, a strong immune system is needed to prevent and accelerate the elimination of this infectious agent.

Immunostimulator is a material that can enhance and improve immune function. Immunostimulator agents from natural ingredients have been developed and applied to prevent or as adjunct therapies^{3,8,9}. One of the potentials as an immunostimulator agent is the Calotropis gigantea plant^{10,11}. There are flavonoids, phenolic, alkaloid, sterol, tannin, and anthraquinone compounds in flowers from *Calotropis gigantea*^{12,13}. The ethanol extract of this flower is known to have a high flavonoid and phenolic content¹⁴. This is supported by the presence of free radical scavenger activities¹⁵, hepatoprotective^{16,17}, and has acted in protection against mast cell degranulation¹⁸. Flavonoids can increase lymphocyte proliferation, which affects CD₄ T cells and Th₁ (T-helper) activation so that macrophages are activated and increase phagocyte activity to kill bacteria or pathogenic microorganisms^{5,6,19}.

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Theoretically, flavonoids and phenolics have proven to be useful as immunostimulator agents. However, as far as our scientific knowledge is concerned, there is no publication of interest from *Calotropis gigantea* as an immunostimulator agent, especially in rats infected with *S. typhi*. Therefore, ethanolic flower extract from *Calotropis gigantea* flower (EECGF) is important to investigate its immunostimulatory activity in rats infected with *S. typhi* bacteria. Immunostimulator parameters measured included CD₄ T cells, leukocytes, lymphocytes, monocyte, neutrophils, and macroscopic spleen. It is expected to be developed in nutraceutical and pharmaceutical products as an immunostimulatory agent.

MATERIAL AND METHODS:

Extract Preparation:

The EECGF preparation procedure follows the existing method with some modifications¹². Fresh *Calotropis gigantea* flowers were collected from Panjang, Bengkulu Indonesia, and identified by certificate number 290/K-ID/ANDA/XI/2015 at the Herbarium Laboratory of Universitas Andalas, Padang Indonesia. The dried flower was macerated using 96% ethanol (Bratachem) solvent. Maceration results are filtered and evaporated with a rotary evaporator (Buchi, Germany) at a temperature of 70°C.

Condition and Design Animal:

The use of rats as test animals has received ethical code with certificate number 144/kepkrsmhfkunsri/2016. Rats were divided into six treatment groups, namely normal, infection, positive control, used a branded product containing Phyllanti extract at a dose of 0.54mL/200g, EECGF therapeutic dose of 40, 80, and 120mg/200g. Rats were adapted for seven days by providing standard drinking and eating and the treatment process for twelve days. Bacterial infection was carried out on the 6th day of all groups except normal rats by giving 0.5mL of *S. typhi* suspension of 10^5 CFU/mL intraperitoneal^{2,3}. Widal test is done after three days of infection. The number of leukocytes, CD₄ T cells, lymphocytes, monocyte, neutrophils, and macroscopic spleen were examined on the 12th day.

Widal Assay:

As many as 1mL of blood was taken trough *retro-orbital plexus* and serum inserted into the tube. Blood samples were left to clot and centrifuged at 5000rpm for 10 minutes. Blood plasma is contained in the upper layer was taken using a micropipette and put into a plastic sample tube. A total of 20mL of serum is dripped onto each circle contained on the surface of the glass slide, added one drop reagent, and observed Widal agglutination occurs. Dilution is done until there is no agglutination when mixed Widal serum and reagents (Primaco)^{20,21}.

Determination of CD4 T Cells:

A total of 25μ L of blood was taken using a micropipette and put into a CD₄ Pima test cartridge (Abbott, US). The sample volume is observed until it reaches the maximum limit. Measurements were made by inserting a cartridge into the Alere PimaTM (Abbott, US). Data results were obtained in the form of CD₄ T cell counts in μ L cells/blood units.

Determination of Total Leukocytes:

A 50 μ L blood sample is inserted into the sample withdrawal tube contained in the Sysmex KX-21N hematology analyzer (Sysmex, Kobe, Japan) and press the sample bar. The calculation of total leukocytes is processed when the withdrawal hose enters the sample automatically into the device. The results obtained in the form of total leukocytes 10³ cells/mL.

Determination of Lymphocytes, Monocytes, and Neutrophils:

Smear preparations are made by dripping a sample of fresh blood on a glass object. After being allowed to dry, the preparation was fixed with methanol for 5 minutes. The preparations were stained using Giemsa coloring (1:9 dilution with phosphate buffer pH 6.8 - 7.2) for 30 minutes. Observation and calculation of lymphocytes, monocytes, and neutrophils are done under a microscope Olympus CX21 (Olympus, UK) with a magnification of 40 times.

Evaluation Macroscopic of Spleen:

The macroscopic examination of the spleen includes the size, weight, color, and consistency of the spleen. The identification of each parameter follows the existing protocol²².

Data Analysis:

The data are presented in mean \pm SD, and statistical analysis using software assistance (SPSS Inc., Chicago, IL, USA). A value of *p*<0.05 was expressed where and indicated there were differences between treatment groups. All groups and responses were analyzed using a chemometric approach with principal component analysis (PCA) (Minitab, State College, PA, USA).

RESULT AND DISCUSSION: Widal Test:

Increased body temperature is a clinical represent of typhoid fever that occurs when infected with *S. typhi* bacteria. Increased body temperature occurs to eliminate bacteria triggered by thermoregulators in the hypothalamus. An increase in body temperature indicates the activation of the immune system²³.

Widal test is done on the 3rd day after infection by identifying the antibodies that are formed. All infected

rats had *S. typhi* antibodies at 10mL observation with an antibody titer of 1:160. The agglutination that occurs indicates a positive Widal test and indicates a clinically significant level of antibody response in the serum. Widal tests generally show positive results one week or more after infection with bacteria through the digestive tract. However, in this study, it occurred on the 3rd day after infection because the induction of *S. typhi* was done intraperitoneally. This modified induction procedure causes within 24 hours the bacteria will enter the blood and form antibodies more quickly.

Evaluation CD4 T Cells:

 CD_4 T cells are a type of lymphocyte cells with CD_4 markers on their cell surfaces, which are the most critical part of the immune system. Optimal CD_4 T cell levels characterize an excellent immune system. The choice of CD_4 T cell immunity parameters is based on its primary function as a system that regulates the formation of a specific immune system against certain microbes.

Based on Fig.1, there was an increase in the average number of CD₄ T cells in the EECGF treatment group. CD₄ T cells increase with increasing dose of EECGF given. This pattern of increasing CD4 T cells is similar to the case of aqueous extract of Vernonia amygdalina in rats that are not antigen-induced²⁴. EECGF doses of 120 mg/200 g showed the highest increase in the number of CD₄ T cells compared with doses of 40 and 80 mg/200 g. Based on statistical analysis, CD₄ and leukocyte data were normally distributed (p>0.05), but the variation of the data was not homogeneous (p < 0.05). Therefore, the analysis used the Kruskal-Wallis test and obtained p < 0.05, which means that there were significant differences between treatment groups. The increased dose of extract causes an increase in the effect of immunostimulators in the body. The EECGF group also had an average CD₄ T cell count higher than the normal and infection groups, and this showed the extract was able to boost the immune system.

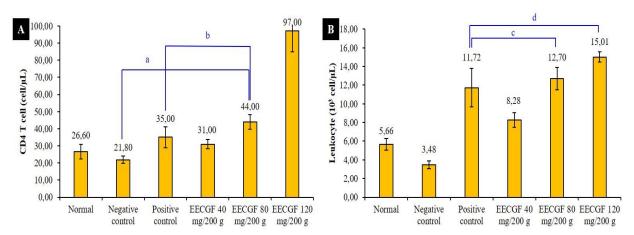


Fig.1: The Results of Measurements of CD₄ T cells (A) and Total Leukocytes (B) in Each Treatment Group, Mann-Whitney Test with significance value; (a) 0.008, (b) 0.043, (c) 0.347, and (d) 0.016

Increased CD₄ T cells can be influenced by active compounds of flavonoids and polyphenols in EECGF. Flavonoids and polyphenols act as immunostimulators by increasing IL-2 activity and lymphocyte proliferation^{9,25}. Lymphocyte proliferation that is affected by CD4 T cells will cause Th1 cells to be activated. Activated Th_1 cells affect IFNy, which can activate macrophages that function to phagocyte antigens. CD₄ T cells also play a role in the formation of antibodies as an identifier in the event of repeated infections^{24,26}. Alkaloid compounds found in Calotropis gigantea flowers can also inhibit bacterial growth so that the activity of S. typhi can be suppressed.

Evaluation Leukocytes Total:

Antigens (*S. typhi*) that enter the blood circulation will stimulate leukocytes to eliminate them. Therefore, leukocytes become a crucial parameter in evaluating

EECGF immunostimulatory activity^{27,28,29}. Based on the results shown in Fig.-1b, there was an increase in total leukocytes in the EECGF group. Total leukocytes tend to increase with increasing doses. This indicates that the specific immune system activity is formed. The flavonoid content in EECGF acts as an immunostimulator, which can accelerate the formation of the immune system^{30,31}.

The EECGF group, with a dose of 120 mg/200 g, had the largest total leukocytes with a high CD₄ T cell count. Products with Phyllanti extract, as a positive control, has high leukocytes, but lower CD₄ T cell counts. In this condition, it is suspected that the non-specific immune system is more dominant, while in EECGF, the formation of specific immune systems occurs more quickly to eliminate antigens. Total leukocyte data alone cannot provide specific information about immune

status, so it is necessary to calculate the number of foreign particles, and dead tissue and then processed to leukocyte cell differentiation, namely lymphocytes, monocytes, and neutrophils³².

Lymphocytes, Monocytes, and Neutrophils:

Lymphocytes are cells that play a significant role in the specific immune system. Each T cell can only interact specifically with antigens that are present on the surface of the antigen-presenting cell (APC) that binds to the major histocompatibility complex (MHC). Observation lymphocytes have large round nuclei that occupy most cells. Sizes vary from 7 to 15 microns. Monocytes can phagocyte and develop into macrophages when they come out of blood vessels and enter tissues. Macrophages also function in processing antigens that are induced at an early stage in the initiation of an response^{33,34}. immune Chemotactic factors pull monocytes into damaged tissue or microbial invasion. Monocytes are classified as mononuclear system cells that play a role in phagocytosis of antigens, destroying foreign particles, and dead tissue and then processed to evoke immune responses. Monocytes have round or long cell nuclei, such as kidneys or horseshoe-like, and have deep indentations. The percentage of normal monocytes is 2-10% of the total components in white blood cells. Neutrophils were chosen as one of the parameters of immunity because of their ability to recognize pathogens directly. Neutrophils are capable of destroying microbes through independent oxygen pathways (lysozyme, lactoferrin, ROI, proteolytic enzymes, cathepsin G, and cationic protein) and oxygen.

Lymphocyte, monocyte, and neutrophil cell profiles are presented in Fig.2. Lymphocyte cells will increase at increasing EECGF doses. A comparison of the number of monocytes and neutrophils is presented in Fig.2b and 2c. When compared with the normal group, the administration of positive control and EECGF provides an increase in all responses determined by either lymphocytes, monocytes, or neutrophils.

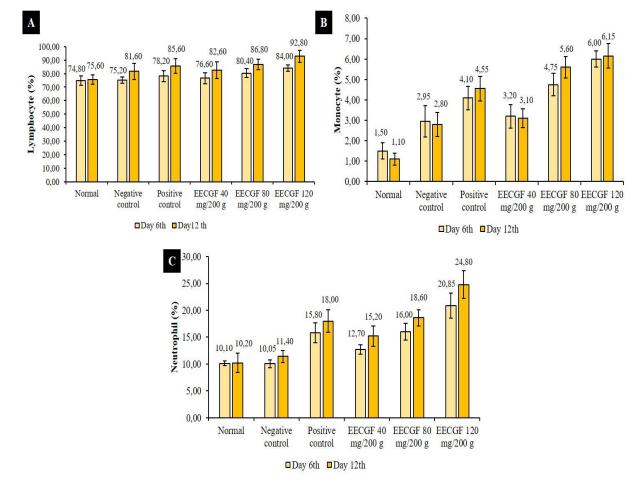
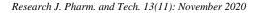


Fig.2: Measurements of Lymphocytes (A), Monocytes (B), and Neutrophils (C)



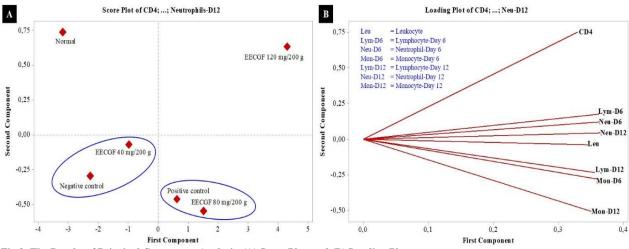


Fig.3: The Results of Principal Component Analysis, (A) Score Plot, and (B) Loading Plot

The chemometric approach with the principal component analysis (PCA) technique obtained results such as Fig.3. This analysis allows us to be able to enter the overall response of all treatment groups and analyze simultaneously. The score plot (Fig.3a) depicts groups with high similarity to be close together³⁵. The positive control with EECGF 80mg/200g had similar properties based on the variables used. Variables or values observed from CD₄ T cells, leukocytes, lymphocytes, monocytes, and neutrophils in the two groups are similar. These results indicate that the administration of extract samples at a dose of 80mg/200g body weight can have pharmacological effects similar to the positive controls used. The infection group and EECGF dose 40 mg/200 are located at adjacent points. EECGF dose of 40mg/200g has not had pharmacological effects because it has properties similar to the infection group.

The loading plot (Fig.3b) is used to reinforce the correlation between the variables being evaluated. The formation of a narrow-angle shows a positive correlation. Some variables that are positively correlated are lymphocytes (day 6) with neutrophils (day 6), neutrophils (day 12) with leukocytes, and lymphocytes with monocytes (day 6). The chemometrics analysis is constructive in grouping and can easily explain the relationship between responses^{36,37,38}.

Evaluation Macroscopic of Spleen:

The spleen is a lymphoid organ that plays a role in the formation of the immune system, which produces lymphocytes B, T, and macrophages. T cell proliferation and the presence of intracellular microorganisms such as *S. typhi* can cause splenomegaly. Evaluation of spleen organs of mice was carried out on a macroscopic change consisting of changes in shape, color, consistency, and weight of the organ.

Macroscopic observation of the spleen (Fig.4f) showed that the EECGF 120 mg/200 g group was abnormal, the spleen enlarged, and the color darkened. The spleen profile in the EECGF 120 mg 200 g group (Fig.4f) was also similar to the group of infectious mice (Fig.4c). The evaluation results in Fig.4, EECGF doses of 40 and 80 mg/200 g show normal spleen.

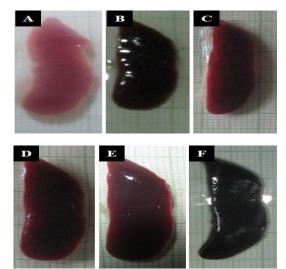


Fig.4: Macroscopic of Spleen (a) Normal, (b) Negative Control, (c) Positive Control, (d) EECGF 40 mg/200 g, (e) EECGF 80 mg/200 g, and (f) EECGF 120 mg/200 g

Fig.4e and f show macroscopic depictions of EECGF 80 and 120 mg/200 g splenic organs. Spleen organs at EECGF 80 mg/200 g showed normal conditions, no lumps, brownish-red color, smooth surface, supple consistency, and a normal size (Table 1). Whereas EECGF 120 mg/200 g did not show any solid mass that was formed, but there was an enlargement of the spleen organ as indicated by the addition of the size and weight of the spleen increased two times the normal state.

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Anatomical	Treatment					
Pathology	Normal	Infection	Positive control	EECGF 40 mg/200 g	EECGF 80 mg/200 g	EECGF 120 mg/200 g
Color	Pink	Black	Brownish-red	Brownish-red	Brownish-red	Black
Shape	Normal	Normal	Normal	Normal	Normal	Normal
Weight (g)	0.50 ± 0.06	0.80±0.10	0.51±0.08	0.51±0.07	0.53±0.08	0.86±0.05
Consistency	Chewy	Hard	Chewy little hard	Chewy little hard	Chewy little hard	Hard

 Table 1: Evaluate the Pathology of Spleen Organ Anatomy (n=5)

There was an enlarged spleen organ in the EECGF group with a dose of 120mg/200 g. The content of polyphenols and flavonoids has a positive correlation with the ability to immunomodulate through stimulation of splenocyte proliferation in the spleen organs³⁹. The administration of the highest dose of EECGF is thought to increase splenocyte proliferation faster than the doses of 40 and 80mg/200g, which can cause splenomegaly.

Free radicals due to bacterial infections and other metabolisms in the body can cause immune cells to be disrupted. The immunomodulatory effects of both flavonoids and phenolics are related to antioxidant activity^{9,30,39}. Sources of flavonoids and phenolics as antioxidant agents from nature are very abundant, for example, white tea and green tea from *Camellia sinensis*^{40,41,42}, herbs *Phyllanti niruri*⁹, and *Calotropis gigantea* flowers¹⁴. High antioxidant agents can increase redox reactions to stabilize free radicals so that they can control immune function^{43,44}.

CONCLUTION:

Ethanol extract of *Calotropis gigantea* L. flower has an immunostimulatory activity. Evaluation of immune system parameters, which include CD_4 T cell counts, total leukocytes, lymphocytes, monocytes, and neutrophils, leads to EECGF at a dose of 80mg/200g having similar activity with positive control. As for the EECGF dose of 120mg/200g, body weight can cause splenomegaly and cause more severe lymph damage. These results are scientific information on the pharmacology of natural materials, phytotherapy, and the basis for further product development.

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CONFLICT OF INTEREST:

The authors declare no conflict of interest.

REFERENCES:

1. Wahyuningrum R, Wahyono D, Mustofa M, Prabandari YS. A Qualitative Study Discovering the Common Medication-Therapy Problems in Patients with Type 2 Diabetes Mellitus (T2DM) in Indonesia. Asian Journal of Pharmaceutical and Clinical Research. 2017; 10(7): 246-250.

- Mathur R, Oh H, Zhang D, Park SG, Seo J, Koblansky A et al. A Mouse Model of Salmonella typhi Infection. Cell. 2012; 151(1): 590-602.
- Fodouop SPC, Gatsing D, Tangue BT, Tagne RS, Tala SD, Tchoumboué J et al. Effect of Salmonella typhimurium Infection on Rat's Cell Oxidation and In Vivo Antioxidant Activity of Vitellaria paradoxa and Ludwigia abyssinica Aqueous Extract. Asian Pacific Journal of Tropical Disease. 2015; 5(1): 38-46.
- 4. Manore D, Pillai S, Joshi A, Punashiya R. Preliminary Phytochemical Screening and Antibacterial Activity of Ethyl Acetate Extract of *Cuscuta reflexa* Roxb. Research Journal of Pharmacy and Technology. 2012; 5(1): 79-82.
- Kurtz JR, Goggins JA, McLachlan JB. Salmonella infection: Interplay Between the Bacteria and Host Immune System. Immunology Letters. 2017; 190: 42-50.
- Lin L, Chen Z, Dai F, Wei JJ, Tang XY, Sun GB. CD4+ T Cells Induce Productions of IL-5 and IL-13 Through MHCII on ILC2s in a Murine Model of Allergic Rhinitis. Auris Nasus Larynx. 2019; 46(4): 533-541.
- Wu J, Cui Y, Zhu W, Bai S, Zhao N, Liu B. Critical Role of OX40/OX40L in ILC2-Mediated Activation of CD4+T Cells During Respiratory Syncytial Virus Infection in Mice. International Immunopharmacology. 2019; 76: 105784.
- 8. Goel A, Kumar S, Bhatia AK. Effect of *Ocimum sanctum* on the Development of Protective Immunity Against *Salmonella typhimurium* Infection Through Cytokines. Asian Pacific Journal of Tropical Medicine. 2010; 3(9): 682-686.
- Pratiwi G, Martien R, Murwanti R. Chitosan Nanoparticle as a Delivery System for Polyphenols from Meniran Extract (*Phyllanthus niruri* L.): Formulation, Optimization, and Immunomodulatory Activity. International Journal of Applied Pharmaceutics. 2019; 11(2): 50-58.
- Pardesi GS, Chhaya G, Vaidya MD. Immunomodulatory Activity of *Calotropis gigantea* by Cyclophosphamide Induced Myelosuppression. Pharmacologyonline. 2008; 2: 164-167.
- 11. Mishra P, Yadav KS, Shrivastava P. An updated Review on Phytochemistry, Pharmacological Activity and Medicinal uses of *Calotropis gigantea* R. Br. Research Journal of Pharmacognosy and Phytochemistry. 2017; 9(2): 135-138.
- 12. Dhivya R, Manimegalai K. Preliminary Phytochemical Screening and GC-MS Profiling of Ethanolic Flower Extract of *Calotropis gigantea* Linn. (Apocynaceae). Journal of Pharmacognosy and Phytochemistry. 2013; 2(3): 28-32.
- Shastry VM, Patil VR, Chaudhari RY. Evaluation of Acute and Chronic Anti-Inflammatory Activity of Whole Plant of *Calotropis* gigantea in Various Experimental Animal Models. Research Journal of Pharmacy and Technology. 2011; 4(5): 815–818.
- David M, BharathK R, Bhavani M. Study of *Calotropis gigantea* R. Br. Extracts on Growth and Survival Dynamics of Selected Pathogenic Microorganisms. International Journal of Biological Engineering. 2012; 1(1): 1-5.
- Usmani S. Screening for Antioxidant and Free Radical Scavenging Potential of Extracts of Leaves and Flowers of *Calotropis* gigantea. Asian Journal of Pharmaceutical and Clinical Research. 2013; 6: 97-100.
- Lodhi G, Singh H, Pant K, Hussain Z. Hepatoprotective Effects of *Calotropis gigantea* Extract Against Carbon Tetrachloride Induced Liver Injury in Rats. Acta Pharmaceutica. 2009; 59(1): 89-96.
- Kshirsagar A, Purnima A. Evaluation of *Calotropis gigantea* R. Br. Flower Extract on Alcohol Induced Hepatotoxicity. Journal of Cell and Tissue Research. 2008; 8(3): 1551-1556.

- Vadnere GP, Gaud RS, Singhai AK, Agrawal AS. Effect of *Calotropis gigantea* Flowers Extracts on Mast Cell Degranulation in Rats. Pharmacologyonline. 2010; 3: 298-303.
- Kanashiro A, Souza JG, Kabeya LM, Azzolini AECS, Lucisano-Valim YM. Elastase Release by Stimulated Neutrophils Inhibited by Flavonoids: Importance of the Catechol Group. Zeitschrift Fur Naturforschung C Journal of Biosciences. 2007; 62(5-6): 357-361.
- Nurjayadi M, Kurnia Dewi F, Kartika IR, Hasan U, Setianingsih I, asiah N *et al.* Development of Antibody Anti-FimC-Salmonella typhi as a Detection Kit Model of Typhoid Diseases by Antigen Capture Approach. Biocatalysis and Agricultural Biotechnology. 2019; 19: 101157.
- Beig FK, Ahmad F, Ekram M, Shukla I. Typhidot M and Diazo test vis-à-vis Blood Culture and Widal Test in the Early Diagnosis of Typhoid Fever in Children in a Resource Poor Setting. The Brazilian Journal of Infectious Diseases. 2010; 14: 589-593.
- 22. Silva SD, Feliciano RP, Boas LV, Bronze MR. Application of FTIR-ATR to Moscatel Dessert Wines for Prediction of Total Phenolic and Flavonoid Contents and Antioxidant Capacity. Food Chemistry. 2014; 150: 489-493.
- 23. Brooks G, Carroll KC, Butel J, Morse S. Medical Microbiology. 26th ed. McGraw-Hill Publishing, New York. 2013.
- Momoh MA, Adikwu MU, Oyi AR. Vernonia amygdalina Extract and CD4 Cell Counts: An Immune Study. Global Journal of Biotechnology and Biochemistry. 2010; 5(2): 92-96.
- Venkatalakshmi P, Vadivel V, Brindha P. Role of Phytochemicals as Immunomodulatory Agents: A Review. International Journal of Green Pharmacy. 2016; 10(1): 1-18.
- 26. Momoh M, Muhamed U, Agboke A, Akpabio E, Uduma EO. Immunological Effect of Aqueous Extract of *Vernonia amygdalina* and a Known Immune Booster Called Immunace® and Their Admixtures on HIV/AIDS Clients: A Comparative Study. Asian Pacific Journal of Tropical Biomedicine. 2012; 2(3): 181-184.
- Akilandeswari K, Anuradha R. Immunostimulatory Activity of Rosa centifolia in Azathioprine Induced Rats. Research Journal Pharmacology and Pharmacodynamics. 2012; 4(4): 210-212.
- Gaba M, Gaba P, Singh S, Dhingra N. Inhibition of LFA-1/ICAM-
 Interaction: A Therapeutic Strategy for Surmounting Inflammation. Asian Journal of Pharmaceutical Research. 2015; 5(1): 37-47.
- Frolova AY, Chernyavskikh SD, Quyet HD, Thanh VV. The Effect of the Incubation Temperature on Morphological and Physical Parameters of *Hypophthalmichthys molitrix* Erythrocytes and Polymorphonuclear Leukocytes. Research Journal of Pharmacy and Technology. 2017; 10(3): 795-798.
- Al-Rekabi MD, Ali SH, Al-Basaisi H, Hashim F, Hussein AH, Abbas HK. Immunomodulatory Effects of Quercetin in Patient with Active Rheumatoid Arthritis. Journal of Advanced Medical Research. 2014; 4(2): 11.
- 31. Hemalatha M, Arirudran B, Thenmozhi A, Rao USM. Antimicrobial Effect of Separate Extract of Acetone, Ethyl Acetate, Methanol and Aqueous from Leaf of Milkweed (*Calotropis gigantea* L.). Asian Journal of Pharmaceutical Research. 2011; 1(4): 102-107.
- Aboderin FI, Oyetayo VO. Haematological Studies of Rats Fed Different Doses of Probiotic, Lactobacillus plantarum, Isolated from Fermenting Corn Slurry. Pakistan Journal of Nutrition. 2006; 5(2): 102-105.
- 33. Yarana C, Thompson H, Chaiswing L, Butterfield DA, Weiss H, Bondada S *et al.* Extracellular Vesicle-Mediated Macrophage Activation: An Insight into the Mechanism of Thioredoxin-Mediated Immune Activation. Redox Biology. 2019; 26: 101237.
- Bellomo A, Gentek R, Bajénoff M, Baratin M. Lymph Node Macrophages: Scavengers, Immune Sentinels and Trophic Effectors. Cellular Immunology. 2018; 330: 168-174.
- 35. Cai T, Wu H, Qin J, Qiao J, Yang Y, Wu Y *et al.* In Vitro Evaluation by PCA and AHP of Potential Antidiabetic Properties of Lactic Acid Bacteria Isolated from Traditional Fermented Food. LWT 2019; 115: 108455.

- Shirsat MK. Chemometric Study of Herbal Extracts of Calotropis gigantea Linn. By FT-IR Spectroscopy. Research Journal of Pharmacy and Technology. 2011; 4(7): 1052-1054.
- Hoang VD. Chemometrics-assisted Spectrophotometric Determination of Ciprofloxacin and Naphazoline in Eye Drops. Asian Journal of Research in Chemistry. 2014; 7(5): 461-465.
- Kathirvel S, Sahiti V, Suneetha A. Spectrophotometric Estimation of Risedronate Sodium in Bulk and Pharmaceutical Formulations using Multivariate Technique. Research Journal of Pharmacy and Technology. 2010; 3(4): 1209-1212.
- Lin JY, Tang CY. Determination of Total Phenolic and Flavonoid Contents in Selected Fruits and Vegetables, as well as their Stimulatory Effects on Mouse Splenocyte Proliferation. Food Chemistry. 2007; 101(1): 140-147.
- 40. Shiyan S, Hertiani T, Martien R, Nugroho AK. Optimization and Validation of RP-HPLC/UV Detection for Several Compounds Simultaneously in Semi-Purified Extract of White Tea. Rasayan Journal of Chemistry. 2019; 12(3): 1098-1109.
- 41. Shiyan S, Hertiani T, Martien R, Nugroho AK. Optimization of a Novel Kinetic-Assisted Infundation for Rich-EGCG and Polyphenols of White Tea (*Camellia sinensis*) Using Central Composite Design. International Journal of Applied Pharmaceutics. 2018; 10(6): 259-267.
- 42. Setyawan EI, Setyawati EP, Rohman A, Nugroho AK. Central Composite Design for Optimizing Extraction of EGCG from Green Tea Leaf (*Camellia sinensis* L.). International Journal of Applied Pharmaceutics. 2018; 10(6): 211-216.
- Krifa M, Bouhlel I, Ghedira-Chekir L, Ghedira K. Immunomodulatory and Cellular Anti-Oxidant Activities of an Aqueous Extract of *Limoniastrum guyonianum* Gall. Journal of Ethnopharmacology. 2013; 146(1): 243-249.
- 44. Cheryomushkina IV, Korneeva OS. Innovative Biotechnology Probiotic Feed Additives C and Immunostimulatory Effects. Research Journal of Pharmacy and Technology. 2017; 10(4): 1165-1167.