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Steroid Compounds from *Gynura pseudochina* (Lour) DC

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ABSTRACT

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Daun dewa (Gynura pseudochina Lour DC) is a one of popular traditional medicine to treat various diseases. This research was conducted to isolate chemical compounds from daun dewa leaves using various chromatographic techniques. A steroid mixture namelye of β -sitosterol (1a) and stigmasterol (1b) were isolated for the first time from methanol extract of daun dewa. The structures were determined base on spectral evidence including IR, NMR 1D and NMR 2D.

Keywords: *Gynura pseudochina*, β -sitosterol, stigmasterol.

ABSTRAK

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*Daun dewa (Gynura pseudochina Lour DC) merupakan salah satu tumbuhan tradisional yang dimanfaatkan untuk mengobati berbagai penyakit. Penelitian ini bertujuan untuk mengisolasi senyawa kimia dari daun tumbuhan daun dewa. Suatu campuran steroid yaitu β -sitosterol (1a) dan stigmasterol (1b) telah diisolasi untuk pertama kalinya dari ekstrak metanol *Gynura pseudochina* (Lour) DC. Struktur kedua senyawa ditetapkan berdasarkan data-data spektroskopi yaitu IR, NMR 1D dan 2D.*

Kata kunci: *Gynura pseudochina*, β -sitosterol, stigmasterol

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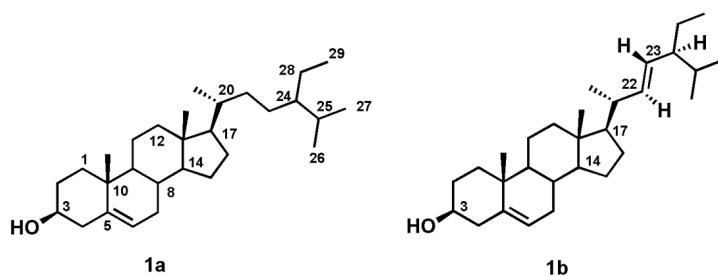
INTRODUCTION

Gynura pseudochina (Lour) DC which locally name is daun dewa belongs to the Asteraceae family. The plant is a well-known traditional herbs in South East Asia and it is widely used to treat eruptive fever, ulcer, detoxification, bleeding, rash, diabetes militus, herves and cancer (Lemmens & Bunyaphratharsa, 2003 ; Hew et.al., 2013). This genus contains some bioactive compounds such as alkaloid, flavonoid and terpenoid. Some

extract of this genus have been reported as antioxidant (Wan, et al., 2011), antihyperglycemia, antihypertension (Wu et al., 2011), antiangiogenic (Seow, et al., 2011) NF- κ B inhibitory (Siriwatanametanon & Heinrich, 2011) and antidiabetic (Hassan et al., 2010).

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Some phenolic compounds such as quercetin 3-rutinoside, 3,5-dicaffeoylquinic acid, 4,5-di-caffeoylquinic acid, and 5-monocaffeoylquinic acid have been isolated from *G. pseudochina* (Siriwatanametanon & Heinrich, 2011). As a part of our research on phytochemistry from *G. pseudochina*, two steroid compound namely β -sitosterol (**1a**) and stigmasterol (**1b**) have been isolated from methanol extract of this plant. The steroid was isolated as a mixture compound. The isolation and the structure elucidation of a mixture steroid will be reported.



EXPERIMENTAL SECTION

Materials

Daun dewa (*G. pseudochina*) were collected from Bandung, West Java, Indonesia. The the plant species was identified at Herbarium Bogoriense, Cibinong, Indonesia. Vacuum liquid chromatography (VLC) was carried out using Merck silica gel 60 GF₂₅₄, while column chromatography was carried out using Merck silica gel 60 (70-230 mesh). Silica gel plates (Merck Kieselgel 60 GF₂₅₄, 0.25 mm) were used for TLC analysis. Visualization of TLC plates was carried out under UV at 254 nm, as well as by spraying the plates with cerium sulfat 1,5 % in sulfuric acid 2 N. The organic solvents were used in this research should be pro analysis (p.a) and distilled, i.e., chloroform, methanol, *n*-hexane, ethyl acetate and acetone.

Instrumentation

Melting points were determined using Fisher John Apparatus. IR spectra were determined with a Perkin Elmer FTIR Spectrum One spectrometer using KBr pellets. ^1H and ^{13}C NMR spectra were recorded with Agilent DD2 spectrometer, operating at 500 (^1H) and 125 (^{13}C) MHz, using residual and deuterated solvent peaks as reference standards.

Procedure

The dried powder of *G. pseudochina* (1 kg) were extracted with methanol at room temperature for 24 hour (3 L), the process were repeated for three times. The methanol extracts were evaporated under reduce pressure to give a dark-green residue (35.71 g). Fractionation of methanol extract using vacuum liquid chromatography (VLC) (silica gel, eluted with *n*-hexane:EtOAc = 10:0 → 0:10 and EtOAc:MeOH = 9:1) afforded ten major fraction A-J (2.14; 0.39; 0.08; 0.46; 0.62; 0.53; 0.78; 1.04; 1.76 and 5.05 g respectively). Fraction B (0.39 g) was further separated by column chromatography (silica gel, eluted with *n*-hexane:EtOAc = 98:2 to 85:15) to give compound **1** (165 mg).

RESULTS AND DISCUSSION

A solid white compound (165 mg) with m.p 123-125 °C was isolated from methanol extract of *G. pseudochina* after separated by several chromatographic techniques. The isolated compound always show one spot on TLC in various of eluen system. The IR spectra showed the functional group of steroid compound such as C-H aliphatic group (2933 - 2869 and 1463 -1384 cm^{-1}), isolated C=C (1642 cm^{-1}) and hydroxyl group (3430 cm^{-1}) that supported by C-O group (1053 cm^{-1}). NMR spectra (^1H -NMR, ^{13}C -NMR and HMQC) analysis of isolated compound was indicated to be a mixture of two steroid compounds. It's based on the presence of twelve methyl groups, four of them are singlet methyl. Typically, a steroid only has six methyl groups including two of them are singlet methyl. Based on the signal intensity of these four methyl singet on ^{13}C -NMR spectra (δ_{C} 12.0, 12.1, 12.2 qnd 12.4 ppm) then the composition of the steroid mixture was 2 : 1. The assumption was supported by comparison signal intensity between methyne vinylic signal at δ_{C} 129.4 & 138.4 (for minor compound) with methyne vinylic signal at δ_{C} 121.8 ppm (for two steroid mixture). By combining data from the ^1H -NMR, ^{13}C -NMR, HMQC and observed the signal intensity of carbon, the signal of each steroid could be determined.

The ^{13}C -NMR data (Table 1) of major compound (**1a**) disclosed the presence of 29 carbon signals that indicated of stigmastane steroid. These signals were including 6 signals for methyl carbon (δ_{C} 12.0; 12.1; 18.9; 19.2; 19.5 and 20.0 ppm), 11 signals for methylene carbon (δ_{C} 21.3; 23.2; 24.4; 26.2; 28.4; 31.8; 32.0; 34.1; 37.4; 39.9 and 42.4 ppm), 7 signals for metin carbon (δ_{C} 29.3; 32.0; 36.3; 46.0; 50.3; 56.2; 56.9 ppm), one signal for oxymetin (δ_{C} 71.9 ppm), one signal for methin sp^2 (δ_{C} 121.8 ppm) and the rest were signals for quartenary carbon (δ_{C} 36.6 and 42.5 ppm) including quartenary carbon sp^2 (δ_{C} 140.9 ppm). Therefore, the major compound was stigmastane steroid containing a hydroxyl group and a double bond. The ^1H -NMR spectra (Table 1) showed an olefinic proton at δ_{H} 5.34 ppm (1H, *br d*, $J = 4.8$ Hz) and an oxymetin proton at δ_{H} 3.52 ppm (1H, *m*) that characteristic for stigmast-5-en-3-ol steroid. Furthermore, the two of six metil signal that characteristic for stigmastane steroid were appeared as singlet at δ_{H} 0.67 ppm (3H, *s*) and 1.00 ppm (3H, *s*) to be located at C-18 and C-19 respectively. Another three metil signals were displayed as doublet at δ_{H} 0.91 ppm (3H, *d*, $J = 6.4$ Hz); 0.83 ppm (3H, *d*, $J = 6.7$ Hz) and 0.80 ppm (3H, *d*, $J = 6.4$ Hz) to be located at C-21, C-26, C-27 respectively. The rest methyl signal is for C-29, displayed as triplet at δ_{H} 0.84 ppm (3H, *t*, $J = 6.3$ Hz). The long range correlation in the HMBC spectrum between a proton signal at δ_{H} 1.00 ppm with the quartenary sp^2 carbon signal at δ_{C} 140.9 ppm, secured the position of this singlet methyl signal at C-19. The correlation in the COSY spectra between two of doublet methyl signal at δ_{H} 0.83 and 0.80 ppm with the methin signal at δ_{H} 1.66 ppm, confirmed the position of these methyl signals at C-26 and C-27 respectively (Figure). Stereochemistry of hydroxyl group at C-3 was determined base on the NOESY spectra. The NOESY spectra showed correlation between methyl signal at δ_{H} 1.00 ppm (H-19) with a methylene signal at δ_{H} 1.49 ppm (H-2), secured the orientation of this methylene signal as an axial, so that another methylene signal at δ_{H} 1.83 ppm (H-2) is an equatorial. Furthermore, the correlation between oxymethin signal at δ_{H} 3.52 ppm (H-3) with methylene signal at δ_{H} 1.83 ppm (H-2), confirmed the oxymethin signal as an axial orientation so that the hydroxyl group is an equatorial (3 β). Thus, the major compound (**1a**) was assigned as stigmasta-5-en-3 β -ol or β -sitosterol. Comparison NMR data with those reported by Greca et al., 1990 showed high similarity. Other COSY, HMBC and NOESY correlations in support for the structure **1a** are shown in Table 1.

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Table 1 NMR data of major compound (**1a**) in CDCl₃

No	δ_c ppm	δ_H (multiplicity, <i>J</i> Hz)	COSY	HMBC	NOESY
1	37.4	a. 1.84 (1H, <i>m</i>) H-eq b. 1.07 (1H, <i>m</i>) H-ax	H2b, H1b H2a, H1a	C5, C3, C10, C2, C19 C9, C2, C19	- H3-ax
2	31.8	a. 1.83 (1H, <i>m</i>) H-eq b. 1.49 (1H, <i>m</i>) H-ax	H3, H1b H2a, H1a	C3, C4, C10 C3, C4	H3-ax H19
3	71.9	3.52 (1H, <i>m</i>) H-ax	H4, H2a, H2b	-	H2a-eq, H1b-ax
4	42.4	2.25 (2H, <i>m</i>)	H3	C5, C6, C3, C10, C2	-
5	140.9	-	-	-	-
6	121.8	a. 5.34 (1H, <i>br d</i> , 4.8)	H4, H7a, H7b	C4, C10, C7, C8	-
7	32.0	a. 1.97 (1H, <i>m</i>) b. 1.51 (1H, <i>m</i>)	H6, H8, H7b H6, H8	C5, C6, C9, C8 C5, C6, C14, C8	- -
8	32.0	1.44 (1H, <i>m</i>)	H9	C9, C7, C14	-
9	50.3	0.92 (1H, <i>m</i>)	H11a	C8, C19, C12	-
10	36.6	-	-	-	-
11	21.3	a. 1.50 (1H, <i>m</i>) b. 1.43 (1H, <i>m</i>)	H9 H12a	C9, C13, C12, C8 C9, C12, C8	- -
12	39.9	a. 2.00 (1H, <i>m</i>)	H11a, H11b, H12b	C14, C9, C11	-
		b. 1.16 (1H, <i>m</i>)	H12a, H11a	C19, C9, C18	-
13	42.5	-	-	-	-
14	56.9	0.99 (1H, <i>m</i>)	H24a	C13, C15, C8	-
15	24.4	a. 1.56 (1H, <i>m</i>) b. 1.05 (1H, <i>m</i>)	H14 H16a	C14, C13, C16 C14, C16	- -
16	28.4	a. 1.84 (1H, <i>m</i>) b. 1.26 (1H, <i>m</i>)	H15a, H15b H17	C17, C13 C14, C20	- -
17	56.2	1.12 (1H, <i>m</i>)	H16a	C13, C12, C20	H-21
18	12.0	0.67 (3H, <i>s</i>)	-	C12, C13, C14, C17	H-8, H20
19	19.5	1.00 (3H, <i>s</i>)	-	C5, C9, C1, C10	H2-ax
20	36.3	1.35 (1H, <i>m</i>)	H21	C17, C16, C21	H-18
21	18.9	0.91 (3H, <i>d</i> , 6.4)	H20	C17, C20, C22	H-17
22	34.1	a. 1.32 (1H, <i>m</i>) b. 1.01 (1H, <i>m</i>)	H23 -	C17, C20, C21 C17, C23	- -
23	26.2	1.16 (2H, <i>m</i>)	H22a	C24, C22, C28	-
24	46.0	0.92 (1H, <i>m</i>)	-	C25, C23, C28, 29	-
25	29.3	1.66 (1H, <i>m</i>)	H26, H27	C24, C23, C28, C26, C27	-
26	20.0	0.83 (3H, <i>d</i> , 6.7)	H25	C24, C26	-
27	19.2	0.80 (3H, <i>d</i> , 6.4)	H25	C24, C26	-
28	23.2	1.25 (2H, <i>m</i>)	H29	C24, C25, C23, C29	-
29	12.1	0.84 (3H, <i>t</i> , 6.3)	H28	C24, C28	-

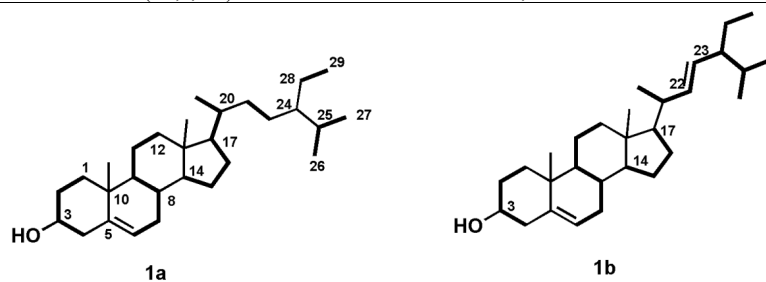


Figure 1. COSY correlation of compound **1a** and **1b**

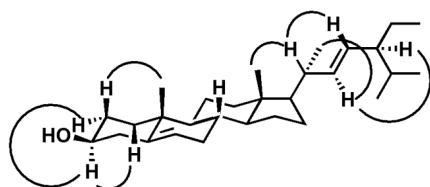


Figure 2. Some NOESY correlation of compound **1b**

The ^{13}C -NMR data (Table 2) of minor compound (**1b**) disclosed the presence of 29 carbon signals that indicated of stigmastane steroid. These signals were including 6 signals for methyl carbon (δ_{C} 12.2; 12.4; 19.5; 19.1; 21.2 and 21.4 ppm), 9 signals for methylene carbon (δ_{C} 21.3; 24.5; 25.6; 29.0; 31.8; 32.0; 37.4; 39.9 and 42.4 ppm), 7 signals for methin carbon (δ_{C} 32.0; 32.1 40.6; 51.4; 50.3; 56.1; 57.0 ppm), one signal for oxymetin (δ_{C} 71.9 ppm), three signals for methin sp^2 (δ_{C} 121.8; 129.4 and 138.4 ppm) and the rest were signals for quaternary carbon (δ_{C} 36.6 and 42.3 ppm) including quaternary carbon sp^2 (δ_{C} 140.9 ppm). These data indicated that the minor and the major steroid compound have the same functional group on tetracyclic skeleton but they are different in the side chain. The ^1H -NMR spectra (Table 2) of compound **1b** showed an olefinic proton signal at δ_{H} 5.34 (1H, *br d*, $J = 4.8$ Hz) and an oxymetin proton at δ_{H} 3.52 ppm (1H, *m*) as well as compound **1a**. The presence of two more olefinic proton signal as double doublet at δ_{H} 5.15 (1H, *dd*, 8.7 & 15.2) and 5.01 ppm (1H, *dd*, 8.7 & 15.2) indicated the side chain have a double bond. The two methyl singlet signal at δ_{H} 0.69 and 1.00 ppm to be located at C-18 and C-19 respectively, while three methyl doublet signal at δ_{H} 1.01; 0.84 and 0.80 ppm for C-21, C-26 and C-27 respectively, and a methyl triplet signal at δ_{H} 0.79 for C-29. Two methin signal at δ_{H} 2.04 and 1.52 ppm as multiplet signal to be located at C-20 and C-24. These signal was shifted to downfield in **1b** compared with **1a**, owing to interaction with double bond. The NOESY spectra showed correlation between methin signal at δ_{H} 2.04 ppm (H-20) with a methyl signal at δ_{H} 0.69 ppm (H-18) and an olefinic proton signal at 5.01 ppm (H-23), indicated they were in the same side (axial orientation). While, correlation between an olefinic proton signal at δ_{H} 5.15 ppm (H-22) with methin signal at δ_{H} 1.12 (H-17) and 1.52 (H-24) ppm also indicated they were in the same side (equatorial orientation) but an opposite side to H-20 (Figure 2). These correlation were indicated that both of olefinic proton signal as trans position as well as the coupling constant analysis (J =

15,2 Hz). Thus, the minor compound (**1b**) was assigned as stigmasta-5,22-dien-3 β -ol or stigmasterol. Other COSY, HMBC and NOESY correlations in support for the structure **1b** are shown in Table 2.

Both of the isolated compound are known compound. It has been reported from *G. bicolor* (Zhuo, et al., 2008), *G. divaricata* [10] and *G. segetum* [5], but it is the first reported from *G. pseudochina*.

Table 2 NMR data of minor compound (**1b**) in CDCl₃

No	δ_c ppm	δ_H (multiplicity, <i>J</i> Hz)	COSY	HMBC	NOESY
1	37.4	a. 1.84 (1H, <i>m</i>) H-eq b. 1.07 (1H, <i>m</i>) H-ax	H2b, H1b H2a, H1a	C5, C3, C10, C2, C19 C9, C2, C19	- H3-ax
2	31.8	a. 1.83 (1H, <i>m</i>) H-eq b. 1.49 (1H, <i>m</i>) H-ax	H3, H1b H2a, H1a	C3, C4, C10 C3, C4	H3-ax H-19
3	71.9	3.52 (1H, <i>m</i>) H-ax	H4, H2a, H2b	-	H2a-eq, H1b-ax
4	42.4	2.25 (2H, <i>m</i>)	H3	C5, C6, C3, C10, C2	-
5	140.9	-	-	-	-
6	121.8	a. 5.34 (1H, <i>br d</i> , 4.8)	H4, H7a, H7b	C4, C10, C7, C8	-
7	32.0	a. 1.97 (1H, <i>m</i>) b. 1.51 (1H, <i>m</i>)	H6, H8, H7b H6, H8	C5, C6, C9, C8 C5, C6, C14, C8	- -
8	32.0	1.44 (1H, <i>m</i>)	H9	C9, C7, C14	-
9	50.3	0.92 (1H, <i>m</i>)	H11a	C8, C19, C12	-
10	36.6	-	-	-	-
11	21.3	a. 1.50 (1H, <i>m</i>) b. 1.43 (1H, <i>m</i>)	H9 H12a	C9, C13, C12, C8 C9, C12, C8	- -
12	39.8	a. 2.00 (1H, <i>m</i>) b. 1.16 (1H, <i>m</i>)	H11a, H11b, H12b H12a, H11a	C14, C9, C11 C19, C9, C18	- -
13	42.3	-	-	-	-
14	57.0	0.99 (1H, <i>m</i>)	H24a	C13, C15, C8	-
15	24.5	a. 1.56 (1H, <i>m</i>) b. 1.05 (1H, <i>m</i>)	H14 H16a	C14, C13, C16 C14, C16	- -
16	29.0	a. 1.70 (1H, <i>m</i>) b. 1.26 (1H, <i>m</i>)	H15b, H17 H17	C17 C14, C20	- -
17	56.1	1.12 (1H, <i>m</i>)	H16a	C13, C12, C20	H21
18	12.2	0.69 (3H, <i>s</i>)	-	C14, C17	H8, H20
19	19.5	1.00 (3H, <i>s</i>)	-	C5, C9, C1, C10	H2-ax
20	40.6	2.04 (1H, <i>m</i>)	H21, H22	C17, C21, C22, C23	H18, H23
21	21.4	1.01 (3H, <i>d</i>)	H20	C17, C20, C22	H-17
22	138.4	5.15 (1H, <i>dd</i> , 8.7 & 15.2)	H20, H23	C17, C20, C21, C23, C24	H-24, H, 17
23	129.4	5.01 (1H, <i>dd</i> , 8.7 & 15.2)	H22, H24	C20, C22, C24, C25, C28	H20
24	51.4	1.52 (1H, <i>m</i>)	H28a, H28b, H25	C25, C23, C28, C29, C26, C27	H22
25	32.1	1.44 (1H, <i>m</i>)	H24, H27	C24, C23, C26	-
26	21.2	0.84 (3H, <i>d</i>)	H25	C24, C25, C27	-
27	19.1	0.80 (3H, <i>d</i>)	H25	C24, C25, C26	-
28	25.6	a. 1.41 (1H, <i>m</i>) b. 1.16 (1H, <i>m</i>)	H29 H28a, H29	C24, C25, C23, C29 C23, C24	- -
29	12.4	0.79 (3H, <i>t</i> , 8.8)	H28a, H28b	C24, C28	-

CONCLUSION

The steroid mixture namely β -sitosterol and stigmasterol have been isolated for the first time from the methanol extract of *G. pseudochina*. The structures of these mixture were determined by spectroscopic data including IR, NMR 1D and 2D.

ACKNOWLEDGEMENTS

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STEROID COMPOUNDS FROM *GYNURA PSEUDOCHINA* (LOUR) DC

SENYAWA STEROID DARI *GYNURA PSEUDOCHINA* (LOUR) DC

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ABSTRACT

Daun dewa (*Gynura pseudochina* Lour DC) is a one of popular traditional medicine to treat various diseases. This research was conducted to isolate chemical compounds from daun dewa leaves using various chromatographic techniques. A steroid mixture namely β -sitosterol (**1a**) and stigmasterol (**1b**) were isolated for the first time from methanol extract of daun dewa. The structures were determined base on spectral evidence including IR, NMR 1D and NMR 2D.

Keywords: *Gynura pseudochina*, β -sitosterol, stigmasterol.

ABSTRAK

Daun dewa (*Gynura pseudochina* Lour DC) merupakan salah satu tumbuhan tradisional yang dimanfaatkan untuk mengobati berbagai penyakit. Penelitian ini bertujuan untuk mengisolasi senyawa kimia dari daun tumbuhan daun dewa. Suatu campuran steroid yaitu β -sitosterol (**1a**) dan stigmasterol (**1b**) telah diisolasi untuk pertama kalinya dari ekstrak metanol *Gynura pseudochina* (Lour) DC. Struktur kedua senyawa ditetapkan berdasarkan data-data spektroskopi yaitu IR, NMR 1D dan 2D.

Kata kunci: *Gynura pseudochina*, β -sitosterol, stigmasterol

INTRODUCTION

Gynura pseudochina (Lour) DC which locally name in Indonesia known as daun dewa is belongs to the Asteraceae family. The plant is a well-known traditional herbs in South East Asia and it is widely used to treat eruptive fever, ulcer, detoxification, bleeding, rash, diabetes militus, herpes and cancer (Lemmens & Bunyaphatsara, 2003 ; Hew, Ko & Gam, 2013). This genus contains some bioactive compounds such as pyrrolizidine and

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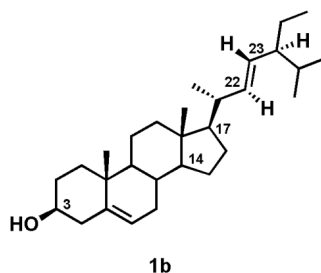
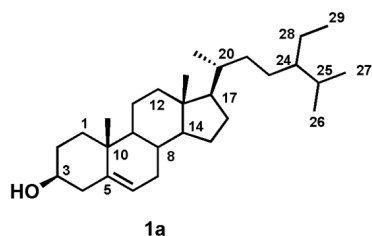
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pyrazine alkaloids (Siriwatanametanon & Heinrich, 2011; Shimizu, et al, 2010), flavonoid, chlorogenic acid (Wan, Yu, Zhou, Tian & Cao, 2011) and terpenoid (Shimizu, et al, 2011). Some extract of this genus have been reported as antioxidant (Wan, et al., 2011), antihyperglycemia, antihypertension (Wu et al., 2011), antiangiogenic (Seow, et al., 2011) NF- κ B inhibitory (Siriwatanametanon & Heinrich, 2011) and antidiabetic (Hassan, Yam, Ahmad, & Yusof, 2010 ; Algarii, et al., 2013).

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Some phenolic compound such as quercetin 3-rutinoside, 3,5-dicaffeoylquinic acid, 4,5-di-caffeoylquinic acid, and 5-monocaffeoylquinic acid have been isolated from *G. pseudochina* (Siriwatanametanon & Heinrich, 2011). These pure compounds showed significant inhibitory activities against α -glucosidase and considerable inhibitory effect against PTP1B which correlated for treatment type-2 diabetic (Chen, 2014). As a part of our research on phytochemistry from *G. pseudochina*, two steroid compound namely β -sitosterol (**1a**) and stigmasterol (**1b**) have been isolated from methanol extract of this plant. The steroid was isolated as a mixture compound. The isolation and the structure elucidation of a mixture steroid will be reported.



EXPERIMENTAL SECTION

Materials

Daun dewa (*G. pseudochina*) were collected from Bandung, West Java, Indonesia. The the plant species was identified at Herbarium Bogoriense, Cibinong, Indonesia. Vacuum liquid chromatography (VLC) was carried out using Merck silica gel 60 GF₂₅₄, while column chromatography was carried out using Merck silica gel 60 (70-230 mesh). Silica gel plates (Merck Kieselgel 60 GF₂₅₄, 0.25 mm) were used for TLC analysis. Visualization of TLC plates was carried out under UV at 254 nm, as well as by spraying the plates with cerium

sulfat 1,5 % in sulfuric acid 2 N. The organic solvents were used in this research should be pro analysis (p.a) and distilled, i.e., chloroform, methanol, *n*-hexane, ethyl acetate and acetone.

Instrumentation

Melting points were determined using Fisher John Apparatus. IR spectra were determined with a Perkin Elmer FTIR Spectrum One spectrometer using KBr pellets. ^1H and ^{13}C NMR spectra were recorded with Agilent DD2 spectrometer, operating at 500 (^1H) and 125 (^{13}C) MHz, using residual and deuterated solvent peaks as reference standards.

Procedure

The dried powder of *G. pseudochina* (1 kg) were extracted with methanol at room temperatur for 24 hour (3 L), the proces were repeated for three times. The methanol extracts were evaporated under reduce pressure to give a dark-green residue (35.71 g). A portion of methanol extract was fractionated on a silica gel vacuum liquid chromatography (VLC) using stepwise gradients of *n*-hexane with increasing amount of ethyl acetate (10:0 to 0:10) and ethyl acetate-methanol, 9:1 to afford ten major fraction A-J (2.14; 0.39; 0.08; 0.46; 0.62; 0.53; 0.78; 1.04; 1.76 and 5.05 g respectively). A greenish solids (0.39 g) contained in the fraction B. This fraction was dissolved in *n*-hexane as little as possible to produce a greenish white solid (260 mg). It was further separated by column chromatography (silica gel, eluted with *n*-hexane-ethyl acetate, 98:2 to 85:15) to give compound **1** (165 mg).

RESULTS AND DISCUSSION

A solid white compound (165 mg) with m.p 123-125 °C was isolated from methanol extract of *G. pseudochina* after separated by several chromatographic techniques. The isolated compound consistently showed one spot on TLC in various eluen system. The IR spectra showed a strong absorption at 2933 and 2869 cm^{-1} were derived from stretching vibration of C-H aliphatic, whereas absorption at 1463 and 1384 cm^{-1} were C-H bending vibration. The absorption at wave number of 3430 cm^{-1} was identified as hydroxyl group (OH) which was supported by C-O vibration at 1053 cm^{-1} . The presence of absorption at 1642 cm^{-1} indicated that compound has an unsaturated bond (C=C). These are a typical absorption of steroid or terpenoid.

NMR spectra ($^1\text{H-NMR}$, $^{13}\text{C-NMR}$ and HMQC) analysis of isolated compound was indicated to be a mixture of two steroid compounds. It's based on the presence of twelve methyl groups, four of them are singlet methyl. Typically, a steroid only has six methyl groups including two of them are singlet methyl. Based on the signal intensity of these four methyl singlet on $^{13}\text{C-NMR}$ spectra (δ_{C} 12.0, 12.1, 12.2 and 12.4 ppm) then the composition of the steroid mixture was 2 : 1. This assumption was supported by comparison signal intensity between methyne vinylic signal at δ_{C} 129.4 & 138.4 (for minor compound) with methyne vinylic signal at δ_{C} 121.8 ppm (for two steroid mixture). By combining data from the $^1\text{H-NMR}$, $^{13}\text{C-NMR}$, HMQC and observed the signal intensity of carbon, the signal of each steroid could be determined.

The $^{13}\text{C-NMR}$ data (**Table 1**) of major compound (**1a**) disclosed the presence of 29 carbon signals that indicated of stigmastane steroid. The $^{13}\text{C-NMR}$ spectra, supported with the information from heteronuclear multiple quantum coherence (HMQC) spectra revealed signal due to 6 signals for methyl carbon (δ_{C} 12.0; 12.1; 18.9; 19.2; 19.5 and 20.0 ppm), 11 signals for methylene carbon (δ_{C} 21.3; 23.2; 24.4; 26.2; 28.4; 31.8; 32.0; 34.1; 37.4; 39.9 and 42.4 ppm), 7 signals for metin carbon (δ_{C} 29.3; 32.0; 36.3; 46.0; 50.3; 56.2; 56.9 ppm), one signal for oxymetin carbon (δ_{C} 71.9 ppm), one signal for methin olefinic carbon (δ_{C} 121.8 ppm) and the rest were signals for quaternary carbon (δ_{C} 36.6 and 42.5 ppm) including quaternary olefinic carbon (δ_{C} 140.9 ppm). Therefore, the major compound was stigmastane steroid containing a hydroxyl group and one double bond. The $^1\text{H-NMR}$ spectra (**Table 1**) showed an olefinic proton at δ_{H} 5.34 ppm (1H, *br d*, $J = 4.8$ Hz) and an oxymetin proton at δ_{H} 3.52 ppm (1H, *m*). These signals are characteristic for stigmast-5-en-3-ol steroid. Furthermore, the two of six metil signal that characteristic for stigmastane steroid were appeared as singlet at δ_{H} 0.67 ppm (3H, *s*) and 1.00 ppm (3H, *s*) to be located at C-18 and C-19 respectively. Another three metil signals were displayed as doublet at δ_{H} 0.91 ppm (3H, *d*, $J = 6.4$ Hz); 0.83 ppm (3H, *d*, $J = 6.7$ Hz) and 0.80 ppm (3H, *d*, $J = 6.4$ Hz) to be located at C-21, C-26, C-27 respectively. The rest methyl signal is for C-29, displayed as triplet at δ_{H} 0.84 ppm (3H, *t*, $J = 6.3$ Hz). The long range correlation in the heteronuclear multiple-bond correlation (HMBC) spectrum between a proton signal at δ_{H} 1.00 ppm with the quaternary sp^2 carbon signal at δ_{C} 140.9 ppm, secured the position of this singlet methyl signal at C-19. The correlation in the correlation spectroscopy (COSY)

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Table 1. NMR data of major compound (**1a**) in CDCl₃

No	δ_C ppm	δ_H (multiplicity, J Hz)	COSY	HMBC	NOESY
1	37.4	a. 1.84 (1H, <i>m</i>) H-eq b. 1.07 (1H, <i>m</i>) H-ax	H2b, H1b H2a, H1a	C5, C3, C10, C2, C19 C9, C2, C19	- H3-ax
2	31.8	a. 1.83 (1H, <i>m</i>) H-eq b. 1.49 (1H, <i>m</i>) H-ax	H3, H1b H2a, H1a	C3, C4, C10 C3, C4	H3-ax H19
3	71.9	3.52 (1H, <i>m</i>) H-ax	H4, H2a, H2b	-	H2a-eq, H1b-ax
4	42.4	2.25 (2H, <i>m</i>)	H3	C5, C6, C3, C10, C2	-
5	140.9	-	-	-	-
6	121.8	a. 5.34 (1H, <i>br d</i> , 4.8)	H4, H7a, H7b	C4, C10, C7, C8	-
7	32.0	a. 1.97 (1H, <i>m</i>) b. 1.51 (1H, <i>m</i>)	H6, H8, H7b H6, H8	C5, C6, C9, C8 C5, C6, C14, C8	- -
8	32.0	1.44 (1H, <i>m</i>)	H9	C9, C7, C14	-
9	50.3	0.92 (1H, <i>m</i>)	H11a	C8, C19, C12	-
10	36.6	-	-	-	-
11	21.3	a. 1.50 (1H, <i>m</i>) b. 1.43 (1H, <i>m</i>)	H9 H12a	C9, C13, C12, C8 C9, C12, C8	- -
12	39.9	a. 2.00 (1H, <i>m</i>) b. 1.16 (1H, <i>m</i>)	H11a, H11b, H12b H12a, H11a	C14, C9, C11 C19, C9, C18	- -
13	42.5	-	-	-	-
14	56.9	0.99 (1H, <i>m</i>)	H24a	C13, C15, C8	-
15	24.4	a. 1.56 (1H, <i>m</i>) b. 1.05 (1H, <i>m</i>)	H14 H16a	C14, C13, C16 C14, C16	- -
16	28.4	a. 1.84 (1H, <i>m</i>) b. 1.26 (1H, <i>m</i>)	H15a, H15b H17	C17, C13 C14, C20	- -
17	56.2	1.12 (1H, <i>m</i>)	H16a	C13, C12, C20	H-21
18	12.0	0.67 (3H, <i>s</i>)	-	C12, C13, C14, C17	H-8, H20
19	19.5	1.00 (3H, <i>s</i>)	-	C5, C9, C1, C10	H2-ax
20	36.3	1.35 (1H, <i>m</i>)	H21	C17, C16, C21	H-18
21	18.9	0.91 (3H, <i>d</i> , 6.4)	H20	C17, C20, C22	H-17
22	34.1	a. 1.32 (1H, <i>m</i>) b. 1.01 (1H, <i>m</i>)	H23 -	C17, C20, C21 C17, C23	- -
23	26.2	1.16 (2H, <i>m</i>)	H22a	C24, C22, C28	-
24	46.0	0.92 (1H, <i>m</i>)	-	C25, C23, C28, 29	-
25	29.3	1.66 (1H, <i>m</i>)	H26, H27	C24, C23, C28, C26, C27	-
26	20.0	0.83 (3H, <i>d</i> , 6.7)	H25	C24, C26	-
27	19.2	0.80 (3H, <i>d</i> , 6.4)	H25	C24, C26	-
28	23.2	1.25 (2H, <i>m</i>)	H29	C24, C25, C23, C29	-
29	12.1	0.84 (3H, <i>t</i> , 6.3)	H28	C24, C28	-

spectra between two of doublet methyl signal at δ_H 0.83 and 0.80 ppm with the methin signal at δ_H 1.66 ppm, confirmed the position of these methyl signals at C-26 and C-27 respectively (**Figure 1a**). Stereochemistry of hydroxyl group at C-3 was determined base on the nuclear overhauser effect spectroscopy (NOESY) spectra. The NOESY spectra showed correlation between methyl signal at δ_H 1.00 ppm (H-19) with a methylene signal at δ_H 1.49 ppm (H-2), secured the orientation of this methylene signal as an axial, so that another methylene signal at δ_H 1.83 ppm (H-2) is an equatorial. Furthermore, the

correlation between oxymethin signal at δ_H 3.52 ppm (H-3) with methylene signal at δ_H 1.83 ppm (H-2), confirmed the oxymethin signal as an axial orientation so that the hydroxyl group is an equatorial (3β). Base on this evidence, the major compound (**1a**) was assigned as stigmasta-5-en-3 β -ol or β -sitosterol. Comparison NMR data with those reported by Greca, Monaco & Previtera, 1990 showed high similarity. Other COSY, HMBC and NOESY correlations in support for the structure **1a** are shown in **Table 1**.

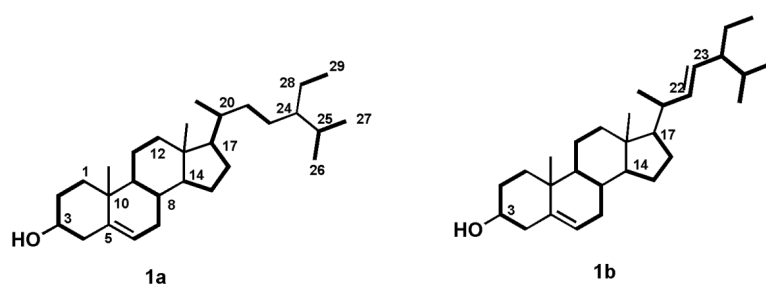


Figure 1. COSY correlation of compound **1a** and **1b**

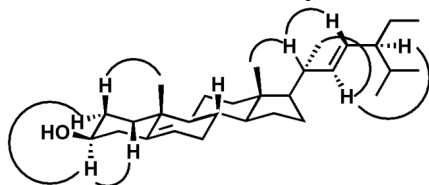


Figure 2. Some NOESY correlation of compound **1b**

The ^{13}C -NMR data (**Table 2**) of minor compound (**1b**) disclosed the presence of 29 carbon signals that indicated of stigmastane steroid. These signals were including 6 signals for methyl carbon (δ_c 12.2; 12.4; 19.5; 19.1; 21.2 and 21.4 ppm), 9 signals for methylene carbon (δ_c 21.3; 24.5; 25.6; 29.0; 31.8; 32.0; 37.4; 39.9 and 42.4 ppm), 7 signals for methin carbon (δ_c 32.0; 32.1 40.6; 51.4; 50.3; 56.1; 57.0 ppm), one signal for oxymetin (δ_c 71.9 ppm), three signals for methin olefinic carbon (δ_c 121.8; 129.4 and 138.4 ppm) and the rest were signals for quartenary carbon (δ_c 36.6 and 42.3 ppm) including quartenary carbon of olefinic (δ_c 140.9 ppm). The presence of these four olefinic were identified for two unsaturated carbon. These data indicated that the minor and the major steroid compound have the same functional group on tetracyclic skeleton but they are different in the side chain. The ^1H -NMR spectra (**Table 2**) of compound **1b** showed an olefinic proton signal at

δ_{H} 5.34 (1H, *br d*, $J = 4.8$ Hz) and an oxymetin proton at δ_{H} 3.52 ppm (1H, *m*) as well as compound **1a**. The presence of two more olefinic proton signal as double doublet at δ_{H} 5.15 (1H, *dd*, 8.7 & 15.2) and 5.01 ppm (1H, *dd*, 8.7 & 15.2) indicated the side chain have a double bond. The two methyl singlet signal at δ_{H} 0.69 and 1.00 ppm to be located at C-18 and C-19 respectively, while three methyl doublet signal at δ_{H} 1.01; 0.84 and 0.80 ppm for C-21, C-26 and C-27 respectively, and a methyl triplet signal at δ_{H} 0.79 for C-29. Two methin signal at δ_{H} 2.04 and 1.52 ppm as multiplet signal to be located at C-20 and C-24 respectively base on the presence of correlation signal at δ_{H} 2.04 with proton olefinic signal at δ_{H} 5.15 and methyl doublet signal at δ_{H} 1.01 ppm, as well as correlation signal δ_{H} 1.52 to signal methylen (δ_{H} 1.41 & 1.16) and methine signal (δ_{H} 1.44) on COSY spectra (**Figure 1b**). These proton signal at C-20 and C-24 were shifted to downfield in **1b** compared with **1a**, owing to interaction with double bond. The NOESY spectra showed correlation between methin signal at δ_{H} 2.04 ppm (H-20) with a methyl signal at δ_{H} 0.69 ppm (H-18) and an olefinic proton signal at 5.01 ppm (H-23), indicated they were in the same side (axial orientation). While, correlation between an olefinic proton signal at δ_{H} 5.15 ppm (H-22) with methin signal at δ_{H} 1.12 (H-17) and 1.52 (H-24) ppm also indicated they were in the same side (equatorial orientation) but an opposite side to H-20 (**Figure 2**). These correlation were indicated that both of olefinic proton signal as *trans* position as well as the coupling constant analysis ($J = 15.2$ Hz). Thus, the minor compound (**1b**) was assigned as stigmasta-5,22-dien-3 β -ol or stigmasterol. Other COSY, HMBC and NOESY correlations in support for the structure **1b** are shown in **Table 2**.

Both of the isolated compound are known compound. It has been reported from *G. bicolor* (Zhuo, et al., 2008), *G. divaricata* (Chen, et al., 2003) and *G. segetum* (Seow, et al., 2011), but according to our knowledge is the first reported from *G. pseudochina*. β -sitosterol and stigmasterol are common phytosterols in plant. They can reduce cholesterol level and as antiinflammation (Huang, Zhong, Chen, Ye & Chen, 2007). Dietary of such phytosterol in food are associated with a cancer reduction with directly inhibit tumor with apoptosis mechanism (Bradford & Award, 2007).

Table 2 NMR data of minor compound (**1b**) in CDCl₃

No	δ_c ppm	δ_H (multiplicity, <i>J</i> Hz)	COSY	HMBC	NOESY
1	37.4	a. 1.84 (1H, <i>m</i>) H-eq b. 1.07 (1H, <i>m</i>) H-ax	H2b, H1b H2a, H1a	C5, C3, C10, C2, C19 C9, C2, C19	- H3-ax
2	31.8	a. 1.83 (1H, <i>m</i>) H-eq b. 1.49 (1H, <i>m</i>) H-ax	H3, H1b H2a, H1a	C3, C4, C10 C3, C4	H3-ax H-19
3	71.9	3.52 (1H, <i>m</i>) H-ax	H4, H2a, H2b	-	H2a-eq, H1b-ax
4	42.4	2.25 (2H, <i>m</i>)	H3	C5, C6, C3, C10, C2	-
5	140.9	-	-	-	-
6	121.8	a. 5.34 (1H, br <i>d</i> , 4.8)	H4, H7a, H7b	C4, C10, C7, C8	-
7	32.0	a. 1.97 (1H, <i>m</i>) b. 1.51 (1H, <i>m</i>)	H6, H8, H7b H6, H8	C5, C6, C9, C8 C5, C6, C14, C8	- -
8	32.0	1.44 (1H, <i>m</i>)	H9	C9, C7, C14	-
9	50.3	0.92 (1H, <i>m</i>)	H11a	C8, C19, C12	-
10	36.6	-	-	-	-
11	21.3	a. 1.50 (1H, <i>m</i>) b. 1.43 (1H, <i>m</i>)	H9 H12a	C9, C13, C12, C8 C9, C12, C8	- -
12	39.8	a. 2.00 (1H, <i>m</i>) b. 1.16 (1H, <i>m</i>)	H11a, H11b, H12b H12a, H11a	C14, C9, C11 C19, C9, C18	- -
13	42.3	-	-	-	-
14	57.0	0.99 (1H, <i>m</i>)	H24a	C13, C15, C8	-
15	24.5	a. 1.56 (1H, <i>m</i>) b. 1.05 (1H, <i>m</i>)	H14 H16a	C14, C13, C16 C14, C16	- -
16	29.0	a. 1.70 (1H, <i>m</i>) b. 1.26 (1H, <i>m</i>)	H15b, H17 H17	C17 C14, C20	- -
17	56.1	1.12 (1H, <i>m</i>)	H16a	C13, C12, C20	H21
18	12.2	0.69 (3H, <i>s</i>)	-	C14, C17	H8, H20
19	19.5	1.00 (3H, <i>s</i>)	-	C5, C9, C1, C10	H2-ax
20	40.6	2.04 (1H, <i>m</i>)	H21, H22	C17, C21, C22, C23	H18, H23
21	21.4	1.01 (3H, <i>d</i>)	H20	C17, C20, C22	H-17
22	138.4	5.15 (1H, <i>dd</i> , 8.7 & 15.2)	H20, H23	C17, C20, C21, C23, C24	H-24, H, 17
23	129.4	5.01 (1H, <i>dd</i> , 8.7 & 15.2)	H22, H24	C20, C22, C24, C25, C28	H20
24	51.4	1.52 (1H, <i>m</i>)	H28a, H28b, H25	C25, C23, C28, C29, C26, C27	H22
25	32.1	1.44 (1H, <i>m</i>)	H24, H27	C24, C23, C26	-
26	21.2	0.84 (3H, <i>d</i>)	H25	C24, C25, C27	-
27	19.1	0.80 (3H, <i>d</i>)	H25	C24, C25, C26	-
28	25.6	a. 1.41 (1H, <i>m</i>) b. 1.16 (1H, <i>m</i>)	H29 H28a, H29	C24, C25, C23, C29 C23, C24	- -
29	12.4	0.79 (3H, <i>t</i> , 8.8)	H28a, H28b	C24, C28	-

CONCLUSION

The steroid mixture namely β -sitosterol and stigmasterol have been isolated for the first time from the methanol extract of *G. pseudochina*. The structures of these mixture were determined by spectroscopic data including IR, NMR 1D and 2D.

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STEROID COMPOUNDS FROM *GYNURA PSEUDOCHINA* (LOUR) DC

SENYAWA STEROID DARI *GYNURA PSEUDOCHINA* (LOUR) DC

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ABSTRACT

Daun dewa (*Gynura pseudochina* Lour DC) is a one of popular traditional medicine to treat various diseases. This research was conducted to isolate chemical compounds from daun dewa leaves using various chromatographic techniques. A steroid mixture namely β -sitosterol (**1a**) and stigmasterol (**1b**) were isolated for the first time from methanol extract of daun dewa. The structures were determined base on spectral evidence including IR, NMR 1D and NMR 2D.

Keywords: *Gynura pseudochina*, β -sitosterol, stigmasterol.

ABSTRAK

Daun dewa (*Gynura pseudochina* Lour DC) merupakan salah satu tumbuhan tradisional yang dimanfaatkan untuk mengobati berbagai penyakit. Penelitian ini bertujuan untuk mengisolasi senyawa kimia dari daun tumbuhan daun dewa. Suatu campuran steroid yaitu β -sitosterol (**1a**) dan stigmasterol (**1b**) telah diisolasi untuk pertama kalinya dari ekstrak metanol *Gynura pseudochina* (Lour) DC. Struktur kedua senyawa ditetapkan berdasarkan data-data spektroskopi yaitu IR, NMR1D dan 2D.

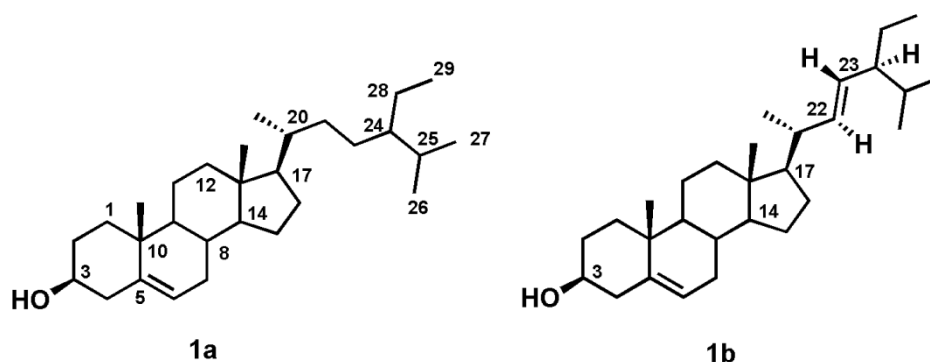
Kata kunci: *Gynura pseudochina*, β -sitosterol, stigmasterol

INTRODUCTION

Gynura pseudochina (Lour) DC which locally name in Indonesia known as daun dewa is belongs to the Asteraceae family. The plant is a well-known traditional herbs in South East Asia and it is widely used to treat eruptive fever, ulcer, detoxification, bleeding, rash, diabetes militus, herves and cancer (Lemmens & Bunyapraphatsara, 2003 ; Hew, Ko & Gam, 2013). This genus contains some bioactive compounds such as pyrrolizidine and

pyrazine alkaloids (Siriwatanametanon & Heinrich, 2011; Shimizu, et al, 2010), flavonoid, chlorogenic acid (Wan, Yu, Zhou, Tian & Cao, 2011) and terpenoid (Shimizu, et al, 2011). Some extract of this genus have been reported as antioxidant (Wan, et al., 2011), antihyperglycemia, antihypertension (Wu et al., 2011), antiangiogenic (Seow, et al., 2011) NF- κ B inhibitory (Siriwatanametanon & Heinrich, 2011) and antidiabetic (Hassan, Yam, Ahmad, & Yusof, 2010 ; Algarii, et al., 2013).

Some phenolic compound such as quercetin 3-rutinoside, 3,5-dicaffeoylquinic acid, 4,5-dicaffeoylquinic acid, and 5-monocaffeoylquinic acid have been isolated from *G. pseudochina* (Siriwatanametanon & Heinrich, 2011). These pure compounds showed significant inhibitory activities against α -glucosidase and considerable inhibitory effect against PTP1B which correlated for treatment type-2 diabetic (Chen, 2014). As a part of our research on phytochemistry from *G. pseudochina*, two steroid compound namely β -sitosterol (**1a**) and stigmasterol (**1b**) have been isolated from methanol extract of this plant. The steroid was isolated as a mixture compound. The isolation and the structure elucidation of a mixture steroid will be reported.



EXPERIMENTAL SECTION

Materials

Daun dewa (*G. pseudochina*) were collected from Bandung, West Java, Indonesia. The plant species was identified at Herbarium Bogoriense, Cibinong, Indonesia. Vacuum liquid chromatography (VLC) was carried out using Merck silica gel 60 GF₂₅₄, while column chromatography was carried out using Merck silica gel 60 (70-230 mesh). Silica gel plates (Merck Kieselgel 60 GF₂₅₄, 0.25 mm) were used for TLC analysis. Visualization of TLC plates was carried out under UV at 254 nm, as well as by spraying the plates with cerium

sulfat 1,5 % in sulfuric acid 2 N. The organic solvents were used in this research should be pro analysis (p.a) and distilled, i.e., chloroform, methanol, *n*-hexane, ethyl acetate and acetone.

Instrumentation

Melting points were determined using Fisher John Apparatus. IR spectra were determined with a Perkin Elmer FTIR Spectrum One spectrometer using KBr pellets. ¹H and ¹³C NMR spectra were recorded with Agilent DD2 spectrometer, operating at 500(¹H) and 125 (¹³C) MHz, using residual and deuterated solvent peaks as reference standards.

Procedure

The dried powder of *G. pseudochina* (1 kg) were extracted with methanol at room temperature for 24 hour (3 L), the process were repeated for three times. The methanol extracts were evaporated under reduce pressure to give a dark-green residue (35.71 g). A portion of methanol extract was fractionated on a silica gel vacuum liquid chromatography (VLC) using stepwise gradients of *n*-hexane with increasing amount of ethyl acetate (10:0 to 0:10) and ethyl acetate-methanol, 9:1 to afford ten major fraction A-J (2.14; 0.39; 0.08; 0.46; 0.62; 0.53; 0.78; 1.04; 1.76 and 5.05 g respectively). A greenish solids (0.39 g) contained in the fraction B. This fraction was dissolved in *n*-hexane as little as possible to produce a greenish white solid (260 mg). It was further separated by column chromatography (silica gel, eluted with *n*-hexane-ethyl acetate, 98:2 to 85:15) to give compound **1** (165 mg).

RESULTS AND DISCUSSION

A solid white compound (165 mg) with m.p 123-125 °C was isolated from methanol extract of *G. pseudochina* after separated by several chromatographic techniques. The isolated compound consistently showed one spot on TLC in various eluent system. The IR spectra showed a strong absorption at 2933 and 2869 cm⁻¹ were derived from stretching vibration of C-H aliphatic, whereas absorption at 1463 and 1384 cm⁻¹ were C-H bending vibration. The absorption at wave number of 3430 cm⁻¹ was identified as hydroxyl group (OH) which was supported by C-O vibration at 1053 cm⁻¹. The presence of absorption at 1642 cm⁻¹ indicated that compound has an unsaturated bond (C=C). These are a typical absorption of steroid or terpenoid.

NMR spectra ($^1\text{H-NMR}$, $^{13}\text{C-NMR}$ and HMQC) analysis of isolated compound was indicated to be a mixture of two steroid compounds. It's based on the presence of twelve methyl groups, four of them are singlet methyl. Typically, a steroid only has six methyl groups including two of them are singlet methyl. Based on the signal intensity of these four methyl singlet on $^{13}\text{C-NMR}$ spectra (δ_{C} 12.0, 12.1, 12.2 and 12.4 ppm) then the composition of the steroid mixture was 2: 1. This assumption was supported by comparison signal intensity between methyne vinylic signal at δ_{C} 129.4 & 138.4 (for minor compound) with methyne vinylic signal at δ_{C} 121.8 ppm (for two steroid mixture). By combining data from the $^1\text{H-NMR}$, $^{13}\text{C-NMR}$, HMQC and observed the signal intensity of carbon, the signal of each steroid could be determined.

The $^{13}\text{C-NMR}$ data (**Table 1**) of major compound (**1a**) disclosed the presence of 29 carbon signals that indicated of stigmastane steroid. The $^{13}\text{C-NMR}$ spectra, supported with the information from heteronuclear multiple quantum coherence (HMQC) spectra revealed signal due to 6 signals for methyl carbon (δ_{C} 12.0; 12.1; 18.9; 19.2; 19.5 and 20.0 ppm), 11 signals for methylene carbon (δ_{C} 21.3; 23.2; 24.4; 26.2; 28.4; 31.8; 32.0; 34.1; 37.4; 39.9 and 42.4 ppm), 7 signals for methin carbon (δ_{C} 29.3; 32.0; 36.3; 46.0; 50.3; 56.2; 56.9 ppm), one signal for oxymetin carbon (δ_{C} 71.9 ppm), one signal for methin olefinic carbon (δ_{C} 121.8 ppm) and the rest were signals for quaternary carbon (δ_{C} 36.6 and 42.5 ppm) including quaternary olefinic carbon (δ_{C} 140.9 ppm). Therefore, the major compound was stigmastane steroid containing a hydroxyl group and one double bond. The $^1\text{H-NMR}$ spectra (**Table 1**) showed an olefinic proton at δ_{H} 5.34 ppm (1H, *br d*, $J=4.8$ Hz) and an oxymetin proton at δ_{H} 3.52 ppm (1H, *m*). These signals are characteristic for stigmast-5-en-3-ol steroid. Furthermore, the two of six methyl signal that characteristic for stigmastane steroid were appeared as singlet at δ_{H} 0.67 ppm (3H, *s*) and 1.00 ppm (3H, *s*) to be located at C-18 and C-19 respectively. Another three methyl signals were displayed as doublet at δ_{H} 0.91 ppm (3H, *d*, $J=6.4$ Hz); 0.83 ppm (3H, *d*, $J=6.7$ Hz) and 0.80 ppm (3H, *d*, $J=6.4$ Hz) to be located at C-21, C-26, C-27 respectively. The rest methyl signal is for C-29, displayed as triplet at δ_{H} 0.84 ppm (3H, *t*, $J=6.3$ Hz). The long range correlation in the heteronuclear multiple-bond correlation (HMBC) spectrum between a proton signal at δ_{H} 1.00 ppm with the quaternary sp^2 carbon signal at δ_{C} 140.9 ppm, secured the position of this singlet methyl signal at C-19. The correlation in the correlation spectroscopy (COSY)

Table 1. NMR data of **major compound** (**nama senyawa**) (**1a**) in CDCl₃

No	δ_{C} ppm	δ_{H} (multiplicity, <i>J</i> Hz)	COSY	HMBC	NOESY
1	37.4	a. 1.84 (1H, <i>m</i>) H-eq b. 1.07 (1H, <i>m</i>) H-ax	H2b, H1b H2a, H1a	C5, C3, C10, C2, C19 C9, C2, C19	- H3-ax
2	31.8	a. 1.83 (1H, <i>m</i>) H-eq b. 1.49 (1H, <i>m</i>) H-ax	H3, H1b H2a, H1a	C3, C4, C10 C3, C4	H3-ax H19
3	71.9	3.52 (1H, <i>m</i>) H-ax	H4, H2a, H2b	-	H2a-eq, H1b-ax
4	42.4	2.25 (2H, <i>m</i>)	H3	C5, C6, C3, C10, C2	-
5	140.9	-	-	-	-
6	121.8	a. 5.34 (1H, <i>brd</i> , 4.8)	H4, H7a, H7b	C4, C10, C7, C8	-
7	32.0	a. 1.97 (1H, <i>m</i>) b. 1.51 (1H, <i>m</i>)	H6, H8, H7b H6, H8	C5, C6, C9, C8 C5, C6, C14, C8	- -
8	32.0	1.44 (1H, <i>m</i>)	H9	C9, C7, C14	-
9	50.3	0.92 (1H, <i>m</i>)	H11a	C8, C19, C12	-
10	36.6	-	-	-	-
11	21.3	a. 1.50 (1H, <i>m</i>) b. 1.43 (1H, <i>m</i>)	H9 H12a	C9, C13, C12, C8 C9, C12, C8	- -
12	39.9	a. 2.00 (1H, <i>m</i>) b. 1.16 (1H, <i>m</i>)	H11a, H11b, H12b H12a, H11a	C14, C9, C11 C19, C9, C18	- -
13	42.5	-	-	-	-
14	56.9	0.99 (1H, <i>m</i>)	H24a	C13, C15, C8	-
15	24.4	a. 1.56 (1H, <i>m</i>) b. 1.05 (1H, <i>m</i>)	H14 H16a	C14, C13, C16 C14, C16	- -
16	28.4	a. 1.84 (1H, <i>m</i>) b. 1.26 (1H, <i>m</i>)	H15a, H15b H17	C17, C13 C14, C20	- -
17	56.2	1.12 (1H, <i>m</i>)	H16a	C13, C12, C20	H-21
18	12.0	0.67 (3H, <i>s</i>)	-	C12, C13, C14, C17	H-8, H20
19	19.5	1.00 (3H, <i>s</i>)	-	C5, C9, C1, C10	H2-ax
20	36.3	1.35 (1H, <i>m</i>)	H21	C17, C16, C21	H-18
21	18.9	0.91 (3H, <i>d</i> , 6.4)	H20	C17, C20, C22	H-17
22	34.1	a. 1.32 (1H, <i>m</i>) b. 1.01 (1H, <i>m</i>)	H23 -	C17, C20, C21 C17, C23	- -
23	26.2	1.16 (2H, <i>m</i>)	H22a	C24, C22, C28	-
24	46.0	0.92 (1H, <i>m</i>)	-	C25, C23, C28, 29	-
25	29.3	1.66 (1H, <i>m</i>)	H26, H27	C24, C23, C28, C26, C27	-
26	20.0	0.83 (3H, <i>d</i> , 6.7)	H25	C24, C26	-
27	19.2	0.80 (3H, <i>d</i> , 6.4)	H25	C24, C26	-
28	23.2	1.25 (2H, <i>m</i>)	H29	C24, C25, C23, C29	-
29	12.1	0.84 (3H, <i>t</i> , 6.3)	H28	C24, C28	-

spectra between two of doublet methyl signal at δ_{H} 0.83 and 0.80 ppm with the methin signal at δ_{H} 1.66 ppm, confirmed the position of these methyl signals at C-26 and C-27 respectively (**Figure 1a**). Stereochemistry of hydroxyl group at C-3 was determined base on the nuclear overhauser effect spectroscopy (NOESY) spectra. The NOESY spectra showed correlation between methyl signal at δ_{H} 1.00 ppm (H-19) with a methylene signal at δ_{H} 1.49 ppm (H-2), secured the orientation of this methylene signal as an axial, so that another methylene signal at δ_{H} 1.83 ppm (H-2) is an equatorial. Furthermore, the

correlation between oxymethin signal at δ_H 3.52 ppm (H-3) with methylene signal at δ_H 1.83 ppm (H-2), confirmed the oxymethin signal as an axial orientation so that the hydroxyl group is an equatorial (3β). Base on this evidence, the major compound (**1a**) was assigned as stigmasta-5-en-3 β -ol or β -sitosterol. Comparison NMR data with those reported by Greca, Monaco & Previtera, 1990 showed high similarity. Other COSY, HMBC and NOESY correlations in support for the structure **1a** are shown in **Table 1**.

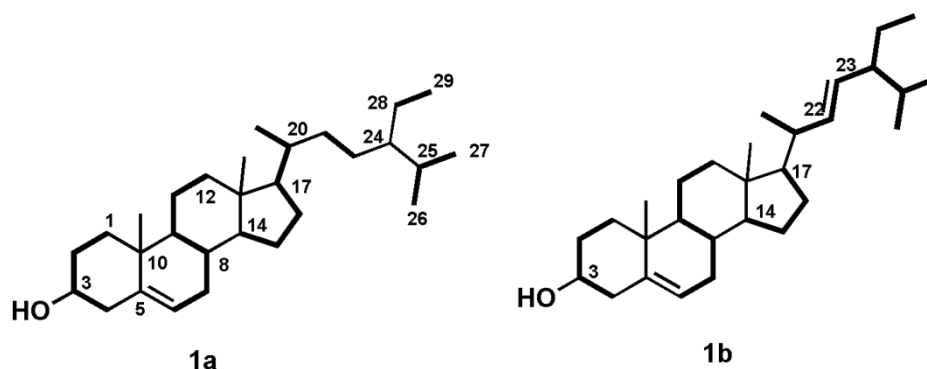


Figure 1. COSY correlation of compound **1a** and **1b**

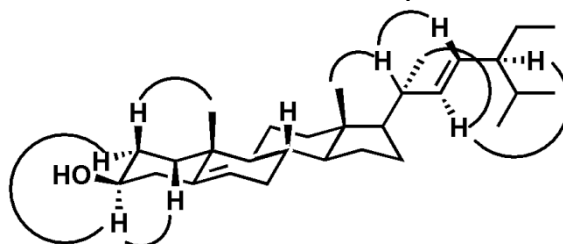


Figure 2. Some NOESY correlation of compound **1b**

The ^{13}C -NMR data (**Table 2**) of minor compound (**1b**) disclosed the presence of 29 carbon signals that indicated of stigmastane steroid. These signals were including 6 signals for methyl carbon (δ_C 12.2; 12.4; 19.5; 19.1; 21.2 and 21.4 ppm), 9 signals for methylene carbon (δ_C 21.3; 24.5; 25.6; 29.0; 31.8; 32.0; 37.4; 39.9 and 42.4 ppm), 7 signals for metin carbon (δ_C 32.0; 32.140.6; 51.4; 50.3; 56.1; 57.0 ppm), one signal for oxymetin (δ_C 71.9 ppm), three signals for methinolefinic carbon (δ_C 121.8; 129.4 and 138.4 ppm) and the rest were signals for quaternary carbon (δ_C 36.6 and 42.3 ppm) including quaternary carbon of olefinic (δ_C 140.9 ppm). The presence of these four olefinic were identified for two unsaturated carbon. These data indicated that the minor and the major steroid compound have the same functional group on tetracyclic skeleton but they are different in the side chain. The ^1H -NMR spectra (**Table 2**) of compound **1b** showed an olefinic proton signal at

δ_{H} 5.34 (1H, *br d*, $J=4.8$ Hz) and an oxymetin proton at δ_{H} 3.52 ppm (1H, *m*) as well as compound **1a**. The presence of two more olefinic proton signal as double doublet at δ_{H} 5.15 (1H, *dd*, 8.7 & 15.2) and 5.01 ppm (1H, *dd*, 8.7 & 15.2) indicated the side chain have a double bond. The two methyl singlet signal at δ_{H} 0.69 and 1.00 ppm to be located at C-18 and C-19 respectively, while three methyl doublet signal at δ_{H} 1.01; 0.84 and 0.80 ppm for C-21, C-26 and C-27 respectively, and a methyl triplet signal at δ_{H} 0.79 for C-29. Two methin signal at δ_{H} 2.04 and 1.52 ppm as multiplet signal to be located at C-20 and C-24 respectively base on the presence of correlation signal at δ_{H} 2.04 with proton olefinic signal at δ_{H} 5.15 and methyl doublet signal at δ_{H} 1.01 ppm, as well as correlation signal δ_{H} 1.52 to signal methylen (δ_{H} 1.41 & 1.16) and methine signal (δ_{H} 1.44) on COSY spectra (**Figure 1b**). These proton signal at C-20 and C-24 were shifted to downfield in **1b** compared with **1a**, owing to interaction with double bond. The NOESY spectra showed correlation between methin signal at δ_{H} 2.04 ppm (H-20) with a methyl signal at δ_{H} 0.69 ppm (H-18) and an olefinic proton signal at 5.01 ppm (H-23), indicated they were in the same side (axial orientation). While, correlation between an olefinic proton signal at δ_{H} 5.15 ppm (H-22) with methin signal at δ_{H} 1.12 (H-17) and 1.52 (H-24) ppm also indicated they were in the same side (equatorial orientation) but an opposite side to H-20 (**Figure 2**). These correlation were indicated that both of olefinic proton signal as *trans* position as well as the coupling constant analysis ($J=15.2$ Hz). Thus, the minor compound (**1b**) was assigned as stigmasta-5,22-dien-3 β -ol or stigmasterol. Other COSY, HMBC and NOESY correlations in support for the structure **1b** are shown in **Table 2**.

Both of the isolated compound are known compound. It has been reported from *G. bicolor* (Zhuo, et al., 2008), *G. divaricata* (Chen, et al., 2003) and *G. segetum* (Seow, et al., 2011), but according to our knowledge is the first reported from *G. pseudochina*. β -sitosterol and stigmasterol are common phytosterols in plant. They can reduce cholesterol level and as antiinflammation (Huang, Zhong, Chen, Ye & Chen, 2007). Dietary of such phytosterol in food are associated with a cancer reduction with directly inhibit tumor with apoptosis mechanism (Bradford & Award, 2007).

Table 2 NMR data of **minor compound** (**nama senyawa**) (**1b**) in CDCl₃

No	$\delta_{\text{C}}(\text{ppm})$	δ_{H} (multiplicity, <i>J</i> Hz)	COSY	HMBC	NOESY
1	37.4	a. 1.84 (1H, <i>m</i>) H-eq b. 1.07 (1H, <i>m</i>) H-ax	H2b, H1b H2a, H1a	C5, C3, C10, C2, C19 C9, C2, C19	- H3-ax
2	31.8	a. 1.83 (1H, <i>m</i>) H-eq b. 1.49 (1H, <i>m</i>) H-ax	H3, H1b H2a, H1a	C3, C4, C10 C3, C4	H3-ax H-19
3	71.9	3.52 (1H, <i>m</i>) H-ax	H4, H2a, H2b	-	H2a-eq, H1b-ax
4	42.4	2.25 (2H, <i>m</i>)	H3	C5, C6, C3, C10, C2	-
5	140.9	-	-	-	-
6	121.8	a. 5.34 (1H, <i>br d</i> , 4.8)	H4, H7a, H7b	C4, C10, C7, C8	-
7	32.0	a. 1.97 (1H, <i>m</i>) b. 1.51 (1H, <i>m</i>)	H6, H8, H7b H6, H8	C5, C6, C9, C8 C5, C6, C14, C8	- -
8	32.0	1.44 (1H, <i>m</i>)	H9	C9, C7, C14	-
9	50.3	0.92 (1H, <i>m</i>)	H11a	C8, C19, C12	-
10	36.6	-	-	-	-
11	21.3	a. 1.50 (1H, <i>m</i>) b. 1.43 (1H, <i>m</i>)	H9 H12a	C9, C13, C12, C8 C9, C12, C8	- -
12	39.8	a. 2.00 (1H, <i>m</i>) b. 1.16 (1H, <i>m</i>)	H11a, H11b, H12b H12a, H11a	C14, C9, C11 C19, C9, C18	- -
13	42.3	-	-	-	-
14	57.0	0.99 (1H, <i>m</i>)	H24a	C13, C15, C8	-
15	24.5	a. 1.56 (1H, <i>m</i>) b. 1.05 (1H, <i>m</i>)	H14 H16a	C14, C13, C16 C14, C16	- -
16	29.0	a. 1.70 (1H, <i>m</i>) b. 1.26 (1H, <i>m</i>)	H15b, H17 H17	C17 C14, C20	- -
17	56.1	1.12 (1H, <i>m</i>)	H16a	C13, C12, C20	H21
18	12.2	0.69 (3H, <i>s</i>)	-	C14, C17	H8, H20
19	19.5	1.00 (3H, <i>s</i>)	-	C5, C9, C1, C10	H2-ax
20	40.6	2.04 (1H, <i>m</i>)	H21, H22	C17, C21, C22, C23	H18, H23
21	21.4	1.01 (3H, <i>d</i>)	H20	C17, C20, C22	H-17
22	138.4	5.15 (1H, <i>dd</i> , 8.7 & 15.2)	H20, H23	C17, C20, C21, C23, C24	H-24, H, 17
23	129.4	5.01 (1H, <i>dd</i> , 8.7 & 15.2)	H22, H24	C20, C22, C24, C25, C28	H20
24	51.4	1.52 (1H, <i>m</i>)	H28a, H28b, H25	C25, C23, C28, C29, C26, C27	H22
25	32.1	1.44 (1H, <i>m</i>)	H24, H27	C24, C23, C26	-
26	21.2	0.84 (3H, <i>d</i>)	H25	C24, C25, C27	-
27	19.1	0.80 (3H, <i>d</i>)	H25	C24, C25, C26	-
28	25.6	a. 1.41 (1H, <i>m</i>) b. 1.16 (1H, <i>m</i>)	H29 H28a, H29	C24, C25, C23, C29 C23, C24	- -
29	12.4	0.79 (3H, <i>t</i> , 8.8)	H28a, H28b	C24, C28	-

CONCLUSION

The steroid mixture namely β -sitosterol and stigmasterol have been isolated for the first time from the methanol extract of *G. pseudochina*. The structures of these mixture were determined by spectroscopic data including IR, NMR 1D and 2D.

ACKNOWLEDGEMENTS

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Reviewer Guide

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Sebaiknya campuran senyawa steroid tersebut **dilengkapi** dengan GCMS sehingga diketahui Mr masing-masing senyawa tersebut. Senyawa tersebut merupakan senyawa yang sering ditemukan pada tumbuhan dan sering merupakan campuran β -sitosterol, stigmasterol dan kolesterol. Dalam pembahasan sebaiknya tabel NMR kedua senyawa dijadikan satu tabel sehingga kelihatan perbedaan pergeseran kimia kedua senyawa steroid. Pembahasan spektrum NMR kedua senyawa steroid tersebut dicari pembeda yang spesifik sehingga jelas yang pergeseran kimia masing-masing senyawa

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STEROID COMPOUNDS FROM *GYNURA PSEUDOCHINA* (LOUR) DC

SENYAWA STEROID DARI *GYNURA PSEUDOCHINA* (LOUR) DC

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ABSTRACT

Daun dewa (*Gynura pseudochina* Lour DC) is a one of popular traditional medicine to treat various diseases. This research was conducted to isolate chemical compounds from daun dewa leaves using various chromatographic techniques. A steroid mixture namely β -sitosterol (**1a**) and stigmasterol (**1b**) were isolated for the first time from methanol extract of daun dewa. The structures were determined base on spectral evidence including IR, NMR 1D and NMR 2D.

Keywords: *Gynura pseudochina*, β -sitosterol, stigmasterol.

ABSTRAK

Daun dewa (*Gynura pseudochina* Lour DC) merupakan salah satu tumbuhan tradisional yang dimanfaatkan untuk mengobati berbagai penyakit. Penelitian ini bertujuan untuk mengisolasi senyawa kimia dari daun tumbuhan daun dewa. Suatu campuran steroid yaitu β -sitosterol (**1a**) dan stigmasterol (**1b**) telah diisolasi untuk pertama kalinya dari ekstrak metanol *Gynura pseudochina* (Lour) DC. Struktur kedua senyawa ditetapkan berdasarkan data-data spektroskopi yaitu IR, NMR 1D dan 2D.

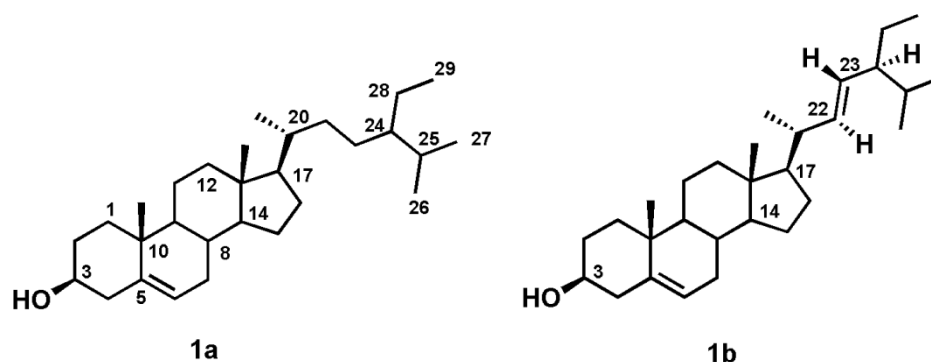
Kata kunci: *Gynura pseudochina*, β -sitosterol, stigmasterol

INTRODUCTION

Gynura pseudochina (Lour) DC which locally name in Indonesia known as daun dewa is belongs to the Asteraceae family. The plant is a well-known traditional herbs in South East Asia and it is widely used to treat eruptive fever, ulcer, detoxification, bleeding, rash, diabetes militus, herves and cancer (Lemmens & Bunyapraphatsara, 2003 ; Hew, Ko & Gam, 2013). This genus contains some bioactive compounds such as pyrrolizidine and

pyrazine alkaloids (Siriwatanametanon & Heinrich, 2011; Shimizu, et al, 2010), flavonoid, chlorogenic acid (Wan, Yu, Zhou, Tian & Cao, 2011) and terpenoid (Shimizu, et al, 2011). Some extract of this genus have been reported as antioxidant (Wan, et al., 2011), antihyperglycemia, antihypertension (Wu et al., 2011), antiangiogenic (Seow, et al., 2011), NF- κ B inhibitory (Siriwatanametanon & Heinrich, 2011) and antidiabetic (Hassan, Yam, Ahmad, & Yusof, 2010 ; Algarii, et al., 2013).

Some phenolic compound such as quercetin 3-rutinoside, 3,5-dicaffeoylquinic acid, 4,5-dicaffeoylquinic acid, and 5-monocaffeoylquinic acid have been isolated from *G. Pseudocina* (Siriwatanametanon & Heinrich, 2011). These pure compounds showed significant inhibitory activities against α -glucosidase and considerable inhibitory effect against PTP1B which correlated for treatment type-2 diabetic (Chen, 2014). As a part of our research on phytochemistry from *G. pseudochina*, two steroid compound namely β -sitosterol (**1a**) and stigmasterol (**1b**) have been isolated from methanol extract of this plant. The steroid was isolated as a mixture compound. The isolation and the structure elucidation of a mixture steroid will be reported.



EXPERIMENTAL SECTION

Materials

Daun dewa (*G. pseudochina*) were collected from Bandung, West Java, Indonesia. The the plant species was identified at Herbarium Bogoriense, Cibinong, Indonesia. Vacuum liquid chromatography (VLC) was carried out using Merck silica gel 60 GF₂₅₄, while column chromatography was carried out using Merck silica gel 60 (70-230 mesh). Silica gel plates (Merck Kieselgel 60 GF₂₅₄, 0.25 mm) were used for TLC analysis. Visualization of TLC plates was carried out under UV at 254 nm, as well as by spraying the plates with cerium

sulfat 1,5 % in sulfuric acid 2 N. The organic solvents were used in this research should be pro analysis (p.a) and distilled, i.e., chloroform, methanol, *n*-hexane, ethyl acetate and acetone.

Instrumentation

Melting points were determined using Fisher John Apparatus. IR spectra were determined with a Perkin Elmer FTIR Spectrum One spectrometer using KBr pellets. ^1H and ^{13}C NMR spectra were recorded with Agilent DD2 spectrometer, operating at 500(^1H) and 125 (^{13}C) MHz, using residual and deuterated solvent peaks as reference standards.

Procedure

The dried powder of *G. pseudochina* (1 kg) were extracted with methanol at room temperatur for 24 hour (3 L), the procces were repeated for three times. The methanol extracts were evaporated under reduce pressure to give a dark-green residue (35.71 g). A portion of methanol extract was fractionated on a silica gel vacuum liquid chromatography (VLC) using stepwise gradients of *n*-hexane with increasing amount of ethyl acetate (10:0 to 0:10) and ethyl acetate-methanol (9:1) to afford ten major fraction A-J (2.14; 0.39; 0.08; 0.46; 0.62; 0.53; 0.78; 1.04; 1.76 and 5.05 g respectively). A greenish solids (0.39 g) contained in the fraction B. This fraction was dissolved in *n*-hexane as little as possible to produce a greenish white solid (260 mg). It was further separated by column chromatography (silica gel, eluted with *n*-hexane-ethyl acetate, 98:2 to 85:15) to give compound **1** (165 mg).

RESULTS AND DISCUSSION

A solid white compound (165 mg) with m.p 123-125 °C was isolated from methanol extract of *G. pseudochina* after separated by several chromatographic techniques. The isolated compound consistently showed one spot on TLC in various eluen system. The IR spectra showed a strong absorption at 2933 and 2869 cm^{-1} were derived from stretching vibration of C-H aliphatic, whereas absorption at 1463 and 1384 cm^{-1} were C-H bending vibration. The absorption at wave number of 3430 cm^{-1} was identified as hydroxyl group (OH) which was supported by C-O vibration at 1053 cm^{-1} . The presence of absorption at 1642 cm^{-1} indicated that compound has an unsaturated bond (C=C). These are a typical absorption of steroid or terpenoid.

NMR spectra ($^1\text{H-NMR}$, $^{13}\text{C-NMR}$ and HMQC) analysis of isolated compound was indicated to be a mixture of two steroid compounds. It's based on the presence of twelve methyl group, four of them are singlet methyl. Typically, a steroid only has six methyl groups including two of them are singlet methyl. Based on the signal intensity of these four methyl singlet on $^{13}\text{C-NMR}$ spectra (δ_{C} 12.0, 12.1, 12.2 and 12.4 ppm) then the composition of the steroid mixture was 2: 1. This assumption was supported by comparison signal intensity between methine vinylic signal at δ_{C} 129.4 & 138.4 (for minor compound) with methine vinylic signal at δ_{C} 121.8 ppm (for two steroid mixture). By combining data from the $^1\text{H-NMR}$, $^{13}\text{C-NMR}$, HMQC and observed the signal intensity of carbon, the signal of each steroid could be determined.

The $^{13}\text{C-NMR}$ data (**Table 1**) of major compound (**1a**) disclosed the presence of 29 carbon signals that indicated of stigmastane steroid. The $^{13}\text{C-NMR}$ spectra, supported with the information from heteronuclear multiple quantum coherence (HMQC) spectra revealed signal due to 6 signals for methyl carbon (δ_{C} 12.0; 12.1; 18.9; 19.2; 19.5 and 20.0 ppm), 11 signals for methylene carbon (δ_{C} 21.3; 23.2; 24.4; 26.2; 28.4; 31.8; 32.0; 34.1; 37.4; 39.9 and 42.4 ppm), 7 signals for methin carbon (δ_{C} 29.3; 32.0; 36.3; 46.0; 50.3; 56.2; 56.9 ppm), one signal for oxymetin carbon (δ_{C} 71.9 ppm), one signal for methin olefinic carbon (δ_{C} 121.8 ppm) and the rest were signals for quaternary carbon (δ_{C} 36.6 and 42.5 ppm) including quaternary olefinic carbon (δ_{C} 140.9 ppm). Therefore, the major compound was stigmastane steroid containing a hydroxyl group and one double bond. The $^1\text{H-NMR}$ spectra (**Table 1**) showed an olefinic proton at δ_{H} 5.34 ppm (1H, *br d*, $J=4.8$ Hz) and an oxymetin proton at δ_{H} 3.52 ppm (1H, *m*). These signals are characteristic for stigmast-5-en-3-ol steroid. Furthermore, the two of six methyl signal that characteristic for stigmastane steroid were appeared as singlet at δ_{H} 0.67 ppm (3H, *s*) and 1.00 ppm (3H, *s*) to be located at C-18 and C-19 respectively. Another three methyl signals were displayed as doublet at δ_{H} 0.91 ppm (3H, *d*, $J = 6.4$ Hz); 0.83 ppm (3H, *d*, $J = 6.7$ Hz) and 0.80 ppm (3H, *d*, $J = 6.4$ Hz) to be located at C-21, C-26, C-27 respectively. The rest methyl signal is for C-29, displayed as triplet at δ_{H} 0.84 ppm (3H, *t*, $J = 6.3$ Hz). The long range correlation in the heteronuclear multiple-bond correlation (HMBC) spectrum between a proton signal at δ_{H} 1.00 ppm with the quaternary sp^2 carbon signal at δ_{C} 140.9 ppm, secured the position of this singlet methyl signal at C-19. The correlation in the correlation

spectroscopy (COSY) spectra between two of doublet methyl signal at δ_H 0.83 and 0.80 ppm with the methin signal at δ_H 1.66 ppm, confirmed the position of these methyl signals at C-26 and C-27 respectively (**Figure 1a**). Stereochemistry of hydroxyl group at C-3 was determined base on the nuclear overhauser effect spectroscopy (NOESY) spectra. The NOESY spectra showed correlation between methyl signal at δ_H 1.00 ppm (H-19) with a methylene signal at δ_H 1.49 ppm (H-2), secured the orientation of this methylene signal as an axial, so that another methylene signal at δ_H 1.83 ppm (H-2) is an equatorial. Furthermore, the correlation between oxymethin signal at δ_H 3.52 ppm (H-3) with methylene signal at δ_H 1.83 ppm (H-2), confirmed the oxymethin signal as an axial orientation so that the hydroxyl group is an equatorial (3β). Correlation between H-18 with H-20 as well as correlation between H-17 with H-21 were secured the side chain on the axial position. These NOESY correlation are equal to compound **1b** (**Figure 2**) Base on this evidence, the major compound (**1a**) was assigned as stigmasta-5-en- 3β -ol or β -sitosterol. Comparison NMR data with those reported by Greca, Monaco & Previtera, 1990 showed high similarity. Other HMBC correlations for supported the structure of **1a** are shown in **Table 1** while COSY correlation are shown in **Figure 1**.

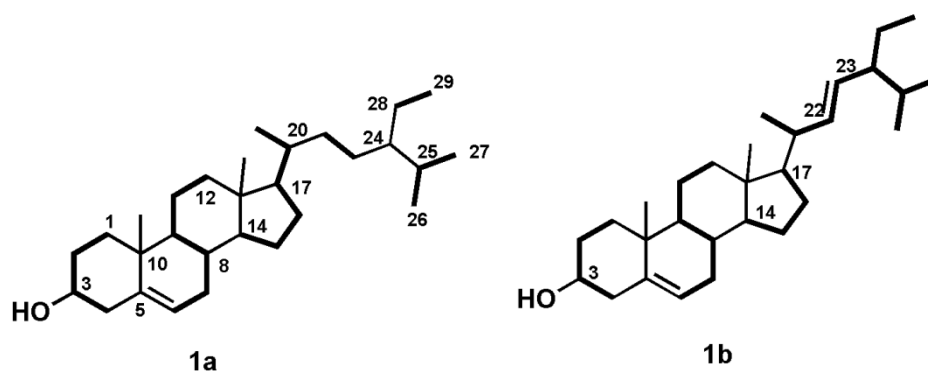


Figure 1. COSY correlation of compound **1a** and **1b**

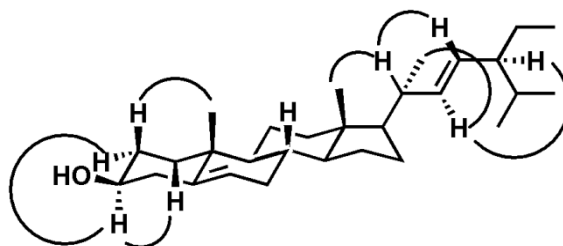


Figure 2. Some NOESY correlation of compound **1b**

Table 1. NMR data of β -sitosterol (**1a**) and stigmasterol (**1b**) in CDCl₃

No	1a		1b		1a & 1b HMBC
	δ_C ppm	δ_H (multiplicity, <i>J</i> Hz)	δ_C ppm	δ_H (multiplicity, <i>J</i> Hz)	
1	37.4	a. 1.84 (1H, <i>m</i>) H-eq b. 1.07 (1H, <i>m</i>) H-ax	37.4	a. 1.84 (1H, <i>m</i>) H-eq b. 1.07 (1H, <i>m</i>) H-ax	C5,C3,C10,C2, C19 C9, C2, C19
2	31.8	a. 1.83 (1H, <i>m</i>) H-eq b. 1.49 (1H, <i>m</i>) H-ax	31.8	a. 1.83 (1H, <i>m</i>) H-eq b. 1.49 (1H, <i>m</i>) H-ax	C3, C4, C10 C3, C4
3	71.9	3.52 (1H, <i>m</i>) H-ax	71.9	3.52 (1H, <i>m</i>) H-ax	-
4	42.4	2.25 (2H, <i>m</i>)	42.4	2.25 (2H, <i>m</i>)	C5, C6, C3, C10, C2
5	140.9	-	140.9	-	-
6	121.8	a. 5.34 (1H, <i>brd</i> , 4.8)	121.8	a. 5.34 (1H, <i>br d</i> , 4.8)	C4, C10, C7, C8
7	32.0	a. 1.97 (1H, <i>m</i>) b. 1.51 (1H, <i>m</i>)	32.0	a. 1.97 (1H, <i>m</i>) b. 1.51 (1H, <i>m</i>)	C5, C6, C9, C8 C5, C6, C14, C8
8	32.0	1.44 (1H, <i>m</i>)	32.0	1.44 (1H, <i>m</i>)	C9, C7, C14
9	50.3	0.92 (1H, <i>m</i>)	50.3	0.92 (1H, <i>m</i>)	C8, C19, C12
10	36.6	-	36.6	-	-
11	21.3	a. 1.50 (1H, <i>m</i>) b. 1.43 (1H, <i>m</i>)	21.3	a. 1.50 (1H, <i>m</i>) b. 1.43 (1H, <i>m</i>)	C9, C13, C12, C8 C9, C12, C8
12	39.9	a. 2.00 (1H, <i>m</i>) b. 1.16 (1H, <i>m</i>)	39.8	a. 2.00 (1H, <i>m</i>) b. 1.16 (1H, <i>m</i>)	C14, C9, C11 C19, C9, C18
13	42.5	-	42.3	-	-
14	56.9	0.99 (1H, <i>m</i>)	57.0	0.99 (1H, <i>m</i>)	C13, C15, C8
15	24.4	a. 1.56 (1H, <i>m</i>) b. 1.05 (1H, <i>m</i>)	24.5	a. 1.56 (1H, <i>m</i>) b. 1.05 (1H, <i>m</i>)	C14, C13, C16 C14, C16
16	28.4	a. 1.84 (1H, <i>m</i>) b. 1.26 (1H, <i>m</i>)	29.0	a. 1.70 (1H, <i>m</i>) b. 1.26 (1H, <i>m</i>)	C17, C13 C14, C20
17	56.2	1.12 (1H, <i>m</i>)	56.1	1.12 (1H, <i>m</i>)	C13, C12, C20
18	12.0	0.67 (3H, <i>s</i>)	12.2	0.69 (3H, <i>s</i>)	C12*, C13*, C14, C17
19	19.5	1.00 (3H, <i>s</i>)	19.5	1.00 (3H, <i>s</i>)	C5, C9, C1, C10
20	36.3	1.35 (1H, <i>m</i>)	40.6	2.04 (1H, <i>m</i>)	C16 *, C17, C21, C22**, C23**
21	18.9	0.91 (3H, <i>d</i> , 6.4)	21.4	1.01 (3H, <i>d</i>)	C17, C20, C22
22	34.1	a. 1.32 (1H, <i>m</i>) b. 1.01 (1H, <i>m</i>)	138.4	5.15 (1H, <i>dd</i> , 8.7 & 15.2)	C17, C20, C21, C23**, C24** C17, C23
23	26.2	1.16 (2H, <i>m</i>)	129.4	5.01 (1H, <i>dd</i> , 8.7 & 15.2)	C22, C24, C28, C20**, C25**, C28**
24	46.0	0.92 (1H, <i>m</i>)	51.4	1.52 (1H, <i>m</i>)	C25, C23, C28, C29, C26**, C27**
25	29.3	1.66 (1H, <i>m</i>)	32.1	1.44 (1H, <i>m</i>)	C24, C23, C28 *, C26, C27
26	20.0	0.83 (3H, <i>d</i> , 6.7)	21.2	0.84 (3H, <i>d</i>)	C24,C25, C27
27	19.2	0.80 (3H, <i>d</i> , 6.4)	19.1	0.80 (3H, <i>d</i>)	C24, C25**, C26
28	23.2	1.25 (2H, <i>m</i>) -	25.6	a. 1.41 (1H, <i>m</i>) b. 1.16 (1H, <i>m</i>)	C24, C25, C23,C29 C23**, C24 **
29	12.1	0.84 (3H, <i>t</i> , 6.3)	12.4	0.79 (3H, <i>t</i> , 8.8)	C24, C28

* : only for compound **1a**** : only for compound **1b**

The ¹³C-NMR data (**Table 1**) of minor compound (**1b**) disclosed the presence of 29 carbon signals that indicated of stigmastane steroid. These signals were including 6 signals

for methyl carbon (δ_C 12.2; 12.4; 19.5; 19.1; 21.2 and 21.4 ppm), 9 signals for methylene carbon (δ_C 21.3; 24.5; 25.6; 29.0; 31.8; 32.0; 37.4; 39.9 and 42.4 ppm), 7 signals for methin carbon (δ_C 32.0; 32.140.6; 51.4; 50.3; 56.1; 57.0 ppm), one signal for oxymethin (δ_C 71.9 ppm), three signals for methin olefinic carbon (δ_C 121.8; 129.4 and 138.4 ppm) and the rest were signals for quaternary carbon (δ_C 36.6 and 42.3 ppm) including quaternary carbon of olefinic (δ_C 140.9 ppm). Some of these carbon signals overlapped with the major compound, especially in the tetracyclic skeleton. The significant difference with the major compound is in the amount of carbon olefinic. The minor compound has four olefinic carbons which were identified as two unsaturated bonds. These data indicated that two steroid compounds have the same functional group on tetracyclic skeleton but they are different in the side chain. The $^1\text{H-NMR}$ spectra of compound **1b** (Table 1) was similar to compound **1a** on tetracyclic skeleton. However, compound **1b** has two additional *trans*-olefinic protons at δ_H 5.15 (1H, *dd*, 8.7 & 15.2 Hz) and 5.01 ppm (1H, *dd*, 8.7 & 15.2 Hz). These signal were indicated the side chain have a double bond at C-22 and C-23 respectively. The HMBC correlation from olefinic proton H-22 (δ_H 5.15) to C-17 (δ_C 56.1), C-20 (δ_C 40.6) and methyl carbon C-21 (δ_C 21.4), as well as HMBC correlation from olefinic proton H-23 (δ_H 5.01) to C-24 (δ_C 51.4) and methylene carbon C-28 (δ_C 25.6) were supported the assignment of double bond at C-22 and C-23. These proton signal at C-20 and C-24 were shifted to downfield in **1b** compared to **1a**, owing to interaction with double bond. The NOESY spectra showed correlation between methin signal at δ_H 2.04 ppm (H-20) with a methyl signal at δ_H 0.69 ppm (H-18) and an olefinic proton signal at 5.01 ppm (H-23), indicated they were in the same side (axial orientation). While, correlation between an olefinic proton signal at δ_H 5.15 ppm (H-22) with methin signal at δ_H 1.12 (H-17) and 1.52 (H-24) ppm also indicated they were in the same side (equatorial orientation) but an opposite side to H-20 (Figure 2). These correlation were indicated that both of olefinic proton signal as *trans* position as well as the coupling constant analysis ($J = 15,2$ Hz). Thus, the minor compound (**1b**) was assigned as stigmasta-5,22-dien-3 β -ol or stigmasterol. Other HMBC correlations are shown in Table 1, while COSY and NOESY correlation are shown in Figure 1 and Figure 2 respectively.

Both of the isolated compound are known compound. It has been reported from *G. Bicolor* (Zhuo, et al., 2008), *G. Divaricata* (Chen, et al., 2003) and *G. Segetum* (Seow, et al., 2011),

but according to our knowledge is the first reported from *G. pseudochina*. β -sitosterol and stigmasterol are common phytosterols in plant. They can reduce cholesterol level and as antiinflammation (Huang, Zhong, Chen, Ye & Chen, 2007). Dietary of such phytosterol in food are associated with a cancer reduction with directly inhibit tumor with apoptosis mechanism (Bradford & Award, 2007).

CONCLUSION

The steroid mixture namely β -sitosterol and stigmasterol have been isolated for the first time from the methanol extract of *G. pseudochina*. The structures of these mixture were determined by spectroscopic data including IR, NMR 1D and 2D.

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No	1a		1b		1a & 1b HMBC
	δ_C ppm	δ_H (multiplicity, <i>J</i> Hz)	δ_C ppm	δ_H (multiplicity, <i>J</i> Hz)	
1	37.4	a. 1.84 (1H, <i>m</i>) H-eq b. 1.07 (1H, <i>m</i>) H-ax	37.4	a. 1.84 (1H, <i>m</i>) H-eq b. 1.07 (1H, <i>m</i>) H-ax	C5,C3,C10,C2, C19 C9, C2, C19
2	31.8	a. 1.83 (1H, <i>m</i>) H-eq b. 1.49 (1H, <i>m</i>) H-ax	31.8	a. 1.83 (1H, <i>m</i>) H-eq b. 1.49 (1H, <i>m</i>) H-ax	C3, C4, C10 C3, C4
3	71.9	3.52 (1H, <i>m</i>) H-ax	71.9	3.52 (1H, <i>m</i>) H-ax	-
4	42.4	2.25 (2H, <i>m</i>)	42.4	2.25 (2H, <i>m</i>)	C5, C6, C3, C10, C2
5	140.9	-	140.9	-	-
6	121.8	a. 5.34 (1H, <i>brd</i> , 4.8)	121.8	a. 5.34 (1H, <i>br d</i> , 4.8)	C4, C10, C7, C8
7	32.0	a. 1.97 (1H, <i>m</i>) b. 1.51 (1H, <i>m</i>)	32.0	a. 1.97 (1H, <i>m</i>) b. 1.51 (1H, <i>m</i>)	C5, C6, C9, C8 C5, C6, C14, C8
8	32.0	1.44 (1H, <i>m</i>)	32.0	1.44 (1H, <i>m</i>)	C9, C7, C14
9	50.3	0.92 (1H, <i>m</i>)	50.3	0.92 (1H, <i>m</i>)	C8, C19, C12
10	36.6	-	36.6	-	-
11	21.3	a. 1.50 (1H, <i>m</i>) b. 1.43 (1H, <i>m</i>)	21.3	a. 1.50 (1H, <i>m</i>) b. 1.43 (1H, <i>m</i>)	C9, C13, C12, C8 C9, C12, C8
12	39.9	a. 2.00 (1H, <i>m</i>) b. 1.16 (1H, <i>m</i>)	39.8	a. 2.00 (1H, <i>m</i>) b. 1.16 (1H, <i>m</i>)	C14, C9, C11 C19, C9, C18
13	42.5	-	42.3	-	-
14	56.9	0.99 (1H, <i>m</i>)	57.0	0.99 (1H, <i>m</i>)	C13, C15, C8
15	24.4	a. 1.56 (1H, <i>m</i>) b. 1.05 (1H, <i>m</i>)	24.5	a. 1.56 (1H, <i>m</i>) b. 1.05 (1H, <i>m</i>)	C14, C13, C16 C14, C16
16	28.4	a. 1.84 (1H, <i>m</i>) b. 1.26 (1H, <i>m</i>)	29.0	a. 1.70 (1H, <i>m</i>) b. 1.26 (1H, <i>m</i>)	C17, C13 C14, C20
17	56.2	1.12 (1H, <i>m</i>)	56.1	1.12 (1H, <i>m</i>)	C13, C12, C20
18	12.0	0.67 (3H, <i>s</i>)	12.2	0.69 (3H, <i>s</i>)	C12*, C13*, C14, C17
19	19.5	1.00 (3H, <i>s</i>)	19.5	1.00 (3H, <i>s</i>)	C5, C9, C1, C10
20	36.3	1.35 (1H, <i>m</i>)	40.6	2.04 (1H, <i>m</i>)	C16 *, C17, C21, C22**, C23**
21	18.9	0.91 (3H, <i>d</i> , 6.4)	21.4	1.01 (3H, <i>d</i>)	C17, C20, C22
22	34.1	a. 1.32 (1H, <i>m</i>) b. 1.01 (1H, <i>m</i>)	138.4	5.15 (1H, <i>dd</i> , 8.7 & 15.2) -	C17, C20, C21, C23**, C24** C17, C23
23	26.2	1.16 (2H, <i>m</i>)	129.4	5.01 (1H, <i>dd</i> , 8.7 & 15.2)	C22, C24, C28, C20**, C25**, C28**
24	46.0	0.92 (1H, <i>m</i>)	51.4	1.52 (1H, <i>m</i>)	C25, C23, C28, C29, C26**, C27**
25	29.3	1.66 (1H, <i>m</i>)	32.1	1.44 (1H, <i>m</i>)	C24, C23, C28 *, C26, C27
26	20.0	0.83 (3H, <i>d</i> , 6.7)	21.2	0.84 (3H, <i>d</i>)	C24,C25, C27
27	19.2	0.80 (3H, <i>d</i> , 6.4)	19.1	0.80 (3H, <i>d</i>)	C24, C25**, C26
28	23.2	1.25 (2H, <i>m</i>) -	25.6	a. 1.41 (1H, <i>m</i>) b. 1.16 (1H, <i>m</i>)	C24, C25, C23,C29 C23**, C24 **
29	12.1	0.84 (3H, <i>t</i> , 6.3)	12.4	0.79 (3H, <i>t</i> , 8.8)	C24, C28

* : only for compound **1a**

** : only for compound **1b**

Table 1. NMR data of β -sitosterol (**1a**) in CDCl₃

No		COSY	HMBC	NOESY			
1		H2b, H1b	C5,C3,C10,C2, C19	-	H2b, H1b	C5,C3, C10,C2,C1 9	-
		H2a, H1a	C9, C2, C19	H3-ax	H2a, H1a	C9, C2, C19	H3-ax
2		H3, H1b	C3, C4, C10	H3-ax	H3, H1b	C3, C4, C10	H3-ax
		H2a, H1a	C3, C4	H19	H2a, H1a	C3, C4	H-19
3		H4, H2a, H2b	-	H2a-eq, H1b-ax	H4, H2a,H2b	-	H2a-eq, H1b-ax
4		H3	C5, C6, C3, C10, C2	-	H3	C5, C6, C3, C10,C2	-
5		-	-	-	-	-	-
6		H4, H7a, H7b	C4, C10, C7, C8	-	H4, H7a,H7b	C4, C10, C7, C8	-
7		H6, H8, H7b	C5, C6, C9, C8	-	H6, H8, H7b	C5, C6, C9, C8	-
		H6, H8	C5, C6, C14, C8	-	H6, H8	C5, C6, C14, C8	-
8		H9	C9, C7, C14	-	H9	C9, C7, C14	-
9		H11a	C8, C19, C12	-	H11a	C8, C19, C12	-
10		-	-	-	-	-	-
11		H9	C9, C13, C12, C8	-	H9	C9, C13, C12, C8	-
		H12a	C9, C12, C8	-	H12a	C9, C12, C8	-
12		H11a, H11b, H12b	C14, C9, C11	-	H11a, H11b, H12b	C14, C9, C11	-
		H12a, H11a	C19, C9, C18	-	H12a, H11a	C19, C9, C18	-
13		-	-	-	-	-	-
14		H24a	C13, C15, C8	-	H24a	C13, C15, C8	-
15		H14	C14, C13, C16	-	H14	C14, C13, C16	-
		H16a	C14, C16	-	H16a	C14, C16	-
16		H15a, H15b	C17, C13	-	H15b, H17	C17	-
		H17	C14, C20	-	H17	C14, C20	-
17		H16a	C13, C12, C20	H-21	H16a	C13, C12, C20	H21
18		-	C12, C13, C14, C17	H-8, H20	-	C14, C17	H8, H20

19		-	C5, C9, C1, C10	H2-ax	-	C5, C9, C1, C10	H2-ax
20		H21	C16, C17, C21	H-18	H21, H22	C17, C21, C22, C23	H18, H23
21		H20	C17, C20, C22	H-17	H20	C17, C20, C22	H-17
22		H23	C17, C20, C21	-	H20, H23	C17, C20, C21, C23, C24	H-24, H, 17
		-	C17, C23	-			
23		H22a	C22, C24, C28	-	H22, H24	C20, C22, C24, C25, C28	H20
24		-	C25, C23, C28, 29	-	H28a, H28b, H25	C25, C23, C28, C29, C26, C27	H22
25		H26, H27	C24, C23, C28, C26, C27	-	H24, H27	C24, C23, C26	-
26		H25	C24, C26	-	H25	C24, C25, C27	-
27		H25	C24, C26	-	H25	C24, C25, C26	-
28		H29	C24, C25, C23, C29	-	H29	C24, C25, C23, C29	-
					H28a, H29	C23, C24	-
29		H28	C24, C28	-	H28a, H28b	C24, C28	-

The ^{13}C -NMR data (**Table 2**) of minor compound (**1b**) disclosed the presence of 29 carbon signals that indicated of stigmastane steroid. These signals were including 6 signals for methyl carbon (δ_{C} 12.2; 12.4; 19.5; 19.1; 21.2 and 21.4 ppm), 9 signals for methylene carbon (δ_{C} 21.3; 24.5; 25.6; 29.0; 31.8; 32.0; 37.4; 39.9 and 42.4 ppm), 7 signals for metin carbon (δ_{C} 32.0; 32.140.6; 51.4; 50.3; 56.1; 57.0 ppm), one signal for oxymetin (δ_{C} 71.9 ppm), three signals for methinolefinic carbon (δ_{C} 121.8; 129.4 and 138.4 ppm) and the rest were signals for quaternary carbon (δ_{C} 36.6 and 42.3 ppm) including quaternary carbon of olefinic (δ_{C} 140.9 ppm). Some of these carbon signal overlapped with the major compound, especially in the tetracyclic skeleton. The significant difference with the major compound is in the amount of carbon olefinic. The minor compound has four olefinic carbons which were identified as two unsaturated bonds. These data indicated that two steroid compounds have the same functional group on tetracyclic skeleton but they are different in the side chain. The ^1H -NMR spectra of compound **1b** was similar to compound **1a** on tetracyclic skeleton. However, compound **1b** has two additional

trans-olefinic protons at δ_{H} 5.15 (1H, *dd*, 8.7 & 15.2 Hz) and 5.01 ppm (1H, *dd*, 8.7 & 15.2 Hz). These signals were indicated the side chain have a double bond at C-22 and C-23 respectively. The HMBC correlation from olefinic proton H-22 (δ_{H} 5.15) to C-17 (δ_{C} 56.1), C-20 (δ_{C} 40.6) and methyl carbon C-21 (δ_{C} 21.4), as well as HMBC correlation from olefinic proton H-23 (δ_{H} 5.01) to C-24 (δ_{C} 51.4) and methylene carbon C-28 (δ_{C} 25.6) were supported the assignment of double bond at C-22 and C-23. These proton signals at C-20 and C-24 were shifted to downfield in **1b** compared to **1a**, owing to interaction with double bond. The NOESY spectra showed correlation between methine signal at δ_{H} 2.04 ppm (H-20) with a methyl signal at δ_{H} 0.69 ppm (H-18) and an olefinic proton signal at 5.01 ppm (H-23), indicated they were in the same side (axial orientation). While, correlation between an olefinic proton signal at δ_{H} 5.15 ppm (H-22) with methine signal at δ_{H} 1.12 (H-17) and 1.52 (H-24) ppm also indicated they were in the same side (equatorial orientation) but an opposite side to H-20 (**Figure 2**). These correlations were indicated that both of olefinic proton signals as *trans* position as well as the coupling constant analysis ($J = 15.2$ Hz). Thus, the minor compound (**1b**) was assigned as stigmasta-5,22-dien-3 β -ol or stigmasterol. Other COSY, HMBC and NOESY correlations in support for the structure **1b** are shown in **Table 2**.

Tanggapan atas saran dari reviewer terhadap manuskrip 293 dengan judul “Steroid Compounds from *Gynura pseudochina*”

- Tabel data NMR telah digabung menjadi satu tabel saja, namun data COSY dan NOESY tidak dapat ditampilkan pada tabel. Tapi korelasi COSY dan NOESY dari senyawa tersebut dapat dilihat pada Gambar 1 dan Gambar 2.
- Pembahasan data NMR juga telah difokuskan pada pembeda yang spesifik yaitu adanya proton dan carbon olefin pada rantai samping.
- Kami tidak dapat melakukan pengukuran GC-MS seperti yang disarankan karena setelah di cek senyawa tersebut telah habis digunakan untuk pengujian bioaktivitas. Berdasarkan penelusuran literatur bahwa campuran phytosterol yang lazim dari tumbuhan adalah β -sitosterol, stigmasterol dan campesterol dengan komposisi sekitar 70%, 20% dan 5% (Christie, 2012). Kalau terdapat campesterol tentunya akan terdapat korelasi COSY antara H-24 dengan gugus metil. Namun pada spektrum COSY tidak ditemukan korelasi COSY antara H-24 ($\sim 0,92$ ppm) dengan proton metil ($\sim 0,85 - 0,70$ ppm). Sehingga disimpulkan tidak terdapat campesterol. Seandainya pada campuran ini terdapat kolesterol (mempunyai 10 metilen), tentunya akan terdapat tambahan satu metilen lainnya, namun hal ini juga tidak ditemukan pada spektrum HSQC.

Referensi:

Christie, W.W. (2012). *Sterols 3. Sterols and Their Conjugates from Plants and Lower Organisms : Structure, Occurrence, Biochemistry and Analysis*. AOCS Lipid Library.
<http://lipidlibrary.aocs.org/Primer/content.cfm?ItemNumber=39358>

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