

DISERTASI

**PENGARUH PEMBERIAN PLATELET RICH FIBRIN
UNTUK MENCEGAH PEMBENTUKAN STRIKTUR
ESOFAGUS AKIBAT CEDERA KAUSTIK BASA
PADA HEWAN COBA TIKUS: TINJAUAN
TERHADAP EKSPRESI Mmp1, Mmp8, Timp1**



PUSPA ZULEIKA

NIM 04013622227001

**PROGRAM STUDI SAINS BIOMEDIS PROGRAM DOKTOR
FAKULTAS KEDOKTERAN
UNIVERSITAS SRIWIJAYA**

2024

HALAMAN PENGESAHAN

**PENGARUH PEMBERIAN PLATELET RICH FIBRIN UNTUK MENCEGAH
PEMBENTUKAN STRIKTUR ESOFAGUS AKIBAT CEDERA KAUSTIK BASA
PADA HEWAN COBA TIKUS: TINJAUAN TERHADAP EKSPRESI Mmp1, Mmp8,
Timp1**

LAPORAN AKHIR DISERTASI

Diajukan Untuk Melengkapi Salah Satu Syarat Memperoleh Gelar
Doktor Program Studi Sains Biomedis Program Doktor

Oleh:

**PUSPA ZULEIKA
04013622227001**

Promotor,

Palembang, Desember 2024
Kopromotor I,

Prof. Dr. Dr. Mgs. M. Irsan Saleh, M.Biomed
NIP 196609291996011001

Prof. Dr. Dr. Krisna Murti, Sp.PA
Subsp.HLE(K), M.Biotech.Stud.Ph.D
NIP 196312101991032002

Kopromotor II,

Dr. Ichel Andriyani Liberty, S.K.M., M.Kes
NIP 199002072015104201

Mengetahui,

Dekan Fakultas Kedokteran Universitas Sriwijaya



dr. Syarif Husin, M.S.
NIP 196112091992031003

SURAT KETERANGAN PENGECEKAN SIMILARITY

Saya yang bertanda tangan di bawah ini

Nama : Puspa Zuleika
NIM : 04013622227001
Prodi : S3 Sains Biomedis

Menyatakan bahwa benar hasil pengecekan similarity Skripsi/Tesis/Disertasi/Lap. Penelitian yang berjudul

PENGARUH PEMBERIAN PLATELET RICH FIBRIN UNTUK MENCEGAH PEMBENTUKAN STRIKTUR ESOFAGUS AKIBAT CEDERA KAUSTIK BASA PADA HEWAN COBA TIKUS: TINJAUAN TERHADAP EKSPRESI Mmp1, Mmp8, Timp1

Adalah 8 %.

Dicek oleh operator *:1. Dosen Pembimbing

2. UPT Perpustakaan

Demikianlah surat keterangan ini saya buat dengan sebenarnya dan dapat saya pertanggung jawabkan.

Indralaya, 8 Januari 2025

Menyetujui
Dosen pembimbing,



Prof. Dr. dr. Mgs. Irsan Saleh, M.Biomed.
NIP: 196609291996011001.



Yang menyatakan,


Puspa Zuleika
NIM: 0401 3622227001

*Lingkari salah satu jawaban, tempat anda melakukan pengecekan Similarity

KATA PENGANTAR

Puji Syukur kepada Tuhan yang Maha Kuasa sehingga disertasi berjudul “**PENGARUH PEMBERIAN PLATELET RICH FIBRIN UNTUK MENCEGAH PEMBENTUKAN STRIKTUR ESOFAGUS AKIBAT CEDERA KAUSTIK BASA PADA HEWAN COBA TIKUS: TINJAUAN TERHADAP EKSPRESI Mmp1, Mmp8, Timp1**” dapat diselesaikan. Disertasi ini disusun untuk melengkapi salah satu syarat guna memperoleh gelar doktor pada Program Studi Sains Biomedis Program Doktor di Fakultas Kedokteran Universitas Sriwijaya.

Terlepas dari berbagai kendala yang dialami Penulis selama penulisan, disertasi ini dapat diselesaikan secara tepat waktu dengan hasil yang memuaskan. Penulis ingin menyampaikan ucapan terima kasih kepada berbagai pihak yang telah mendukung tercapainya penelitian dengan hasil yang memuaskan.

Peneliti secara khusus ingin menyampaikan terima kasih kepada **Prof. Dr. dr. Mgs. Irsan Saleh, M.Biomed., Prof. dr. Krisna Murti, Sp.PA, Subsp.HLE(K), M.BioTech.Stud., Ph.D.,** dan **Dr. Iche Andriyani Liberty, S.K.M., M.Kes.** atas bimbingan dan masukan yang diberikan selama penulisan disertasi ini ini. Penulis juga ingin menyampaikan terima kasih kepada **Prof. Dr. dr. Irfannuddin, Sp.KO, Subsp.APK(K), M.Pd.Ked., Dr. dr. Legiran, M.Kes.,** dan **dr. Agus Surono, Sp.T.H.T.B.K.L., Subsp.BE(K), Ph.D.** atas masukan yang diberikan selama penulisan disertasi ini. Peneliti tak lupa ingin menghaturkan ucapan terima kasih kepada berbagai pihak yang telah mendukung penulisan disertasi ini yang tidak dapat disebutkan satu per satu.

Peneliti berharap bahwa disertasi ini dapat memberikan sekelumit sumbangsih untuk pengembangan ilmu pengetahuan. Akhirnya, Peneliti sangat mengharapkan saran dan kritik yang mendukung perbaikan di masa mendatang.

Palembang, Desember 2024

Puspa Zuleika

ABSTRAK

PENGARUH PEMBERIAN *PLATELET RICH FIBRIN* UNTUK MENCEGAH PEMBENTUKAN STRIKTUR ESOFAGUS AKIBAT CEDERA KAUSTIK BASA PADA HEWAN COBA TIKUS: TINJAUAN TERHADAP EKSPRESI Mmp1, Mmp8, **Timp1**

Cedera kaustik pada esofagus dapat menyebabkan striktur esofagus. Dalam penelitian lain, Mmp1, Mmp8, dan Timp1 berperan dalam patofisiologi striktur esofagus. Sejauh ini berbagai terapi telah diteliti untuk mencegah striktur esofagus dengan hasil dan biaya yang bervariasi. *Platelet-rich fibrin* telah digunakan secara aman dan efektif untuk meningkatkan regenerasi jaringan. Penelitian ini bertujuan untuk membuktikan peran PRF dalam mencegah pembentukan striktur esofagus pada model tikus. Penelitian ini dilakukan dalam dua tahap: validasi model striktur pada tikus secara non-invasif yang dipaparkan dengan NaOH 10% sebanyak 0,1 mL menggunakan sonde dan melihat dampak pemberian PRF pada volume 0,1 mL hingga 0,3 mL terhadap pembentukan striktur pada model tikus yang telah divalidasi sebelumnya. Pemberian PRF untuk tikus berasal dari tikus yang secara khusus disiapkan sebagai donor. Perbedaan ekspresi Mmp1, Mmp8, dan Timp1 diukur antara satu hingga empat belas hari melalui imunohistokimia sementara kerusakan jaringan dan *Stenosis Index* dinilai secara histologis. Penelitian ini berhasil memvalidasi model tikus yang dibuat. Penelitian ini telah menemukan bahwa PRF berpengaruh terhadap ekspresi Mmp1 dan Timp1 tetapi tidak berpengaruh terhadap Mmp8. Penelitian ini juga menemukan bahwa PRF menurunkan kerusakan jaringan dan *Stenosis Index* yang dinilai secara histologis. Striktur esofagus dalam penelitian ini dipengaruhi oleh hari perlakuan, volume PRF yang diberikan, ekspresi Mmp1 dan Timp1, serta kerusakan jaringan yang terjadi. Volume PRF terbaik untuk mencegah striktur ada pada volume 0,3 mL atau volume maksimal.

Kata kunci: *platelet-rich fibrin*, striktur esofagus, Mmp1, Mmp8, Timp1, model tikus

ABSTRACT

ELUCIDATING PLATELET-RICH FIBRIN'S ROLE IN PREVENTION OF CAUSTIC INJURY-RELATED ESOPHAGEAL STRICTURE OF RAT MODEL: EVALUATING CHANGES OF Mmp1, Mmp8, Timp1 EXPRESSION

Caustic esophageal injury may result in esophageal stricture. In other studies, Mmp1, Mmp8, and Timp1 are related to the pathophysiological changes observed in esophageal stricture. Several prospective treatments to prevent esophageal stricture have been studied with varying costs and results. Platelet-rich fibrin has been proven to be safe and efficacious to improve tissue regeneration. This study aims to prove PRF's role in esophageal stricture prevention in the rat model. This study was divided into two phases: validation of a non-invasive rat model exposed to 0.1 mL of 10% NaOH via feeding tube and elucidation of the effect of varying volumes (0.1-0.3 mL) of administered PRF to prevent esophageal stricture on the previously-validated rat model. Platelet-rich fibrin originated from allograft rat donors. Differential expressions of Mmp1, Mmp8, and Timp1 were measured between 1-14 days through standardized immunohistochemical protocols while tissue damage and Stenosis Index were evaluated histologically. This study was able to validate the rat model. Further, this study discovered differential Mmp1 and Timp1 expression after administration while simultaneously discovering a non-significant impact of PRF administration on Mmp8 expression. This study also discovered reduction of histological tissue damage and Stenosis Index after PRF administration. This study discovered the differential role of time, PRF volume administered, Mmp1 and Timp1 expression, and tissue damage in stricture formation. This study discovered that stricture prevention is most significant when PRF was administered at maximum volume (0.3 mL).

Keywords: platelet-rich fibrin, esophageal stricture, Mmp1, Mmp8, Timp1, rat model

RINGKASAN

PENGARUH PEMBERIAN *PLATELET RICH FIBRIN* UNTUK MENCEGAH PEMBENTUKAN STRIKTUR ESOFAGUS AKIBAT CEDERA KAUSTIK BASA PADA HEWAN COBA TIKUS: TINJAUAN TERHADAP EKSPRESI Mmp1, Mmp8, Timp1

Karya tulis ilmiah berupa Disertasi, Desember 2024

Puspa Zuleika, dibimbing oleh Prof. Dr. dr. Mgs. Irsan Saleh, M.Biomed.; Prof. dr. Krisna Murti, Sp.PA, Subsp.HLE(K), M.Biotech.Stud., Ph.D.; dan Dr. Iche Andriyani Liberty, S.K.M., M.Kes.

Program Studi Sains Biomedis Program Doktor, Fakultas Kedokteran Universitas Sriwijaya

Xxi + 205 halaman, 25 tabel, 76 gambar, 10 lampiran

Struktur esofagus merupakan dampak jangka panjang yang mungkin terjadi dari cedera kaustik akibat basa kuat pasca cedera esofagus. Dalam berbagai penelitian yang telah dilaksanakan sebelumnya, diketahui bahwa Mmp1, Mmp8, dan Timp1 berperan dalam patofisiologi striktur esofagus. Sejauh ini berbagai terapi telah diteliti untuk mencegah striktur esofagus dengan hasil dan biaya yang bervariasi. Sebaliknya, *platelet-rich fibrin* telah digunakan secara aman dan efektif untuk meningkatkan regenerasi jaringan. Oleh karena itu, penelitian ini bertujuan untuk membuktikan peran PRF dalam mencegah pembentukan striktur esofagus pada model tikus.

Penelitian ini merupakan penelitian eksperimental *in vivo* dengan desain *post test-only controup group* yang dilakukan dalam dua tahap: validasi model striktur pada tikus secara non-invasif yang dipaparkan dengan NaOH 10% sebanyak 0,1 mL menggunakan sonde dan melihat dampak pemberian PRF pada volume 0,1 mL hingga 0,3 mL terhadap pembentukan striktur pada model tikus yang telah divalidasi sebelumnya. Pemberian PRF untuk tikus berasal dari tikus yang secara khusus disiapkan sebagai donor berdasarkan protokol yang telah disiapkan sebelumnya. Perbedaan ekspresi Mmp1, Mmp8, dan Timp1 diukur antara satu

hingga empat belas hari melalui imunohistokimia sementara kerusakan jaringan dan *Stenosis Index* dinilai secara histologis. Penelitian ini dilaksanakan di Laboratorium *Animal House* dan Laboratorium Bioteknologi Fakultas Kedokteran Universitas Sriwijaya serta Laboratorium Patologi Anatomi RSUP Dr. Mohammad Hoesin Palembang/Fakultas Kedokteran Universitas Sriwijaya antara bulan Januari hingga Juli 2024. Penelitian ini telah disetujui oleh Komite Etik Penelitian Kesehatan Fakultas Kedokteran Universitas Sriwijaya.

Penelitian ini berhasil memvalidasi model tikus non-invasif yang dibuat berdasarkan temuan kerusakan jaringan dan keberadaan *Stenosis Index*. Lebih lanjut, hasil analisis dari penelitian ini telah menemukan bahwa PRF berpengaruh terhadap ekspresi Mmp1 dan Timp1 tetapi tidak berpengaruh terhadap Mmp8. Penelitian ini juga menemukan bahwa PRF menurunkan kerusakan jaringan dan *Stenosis Index* yang dinilai secara histologis. Secara ringkas, dapat disimpulkan bahwa striktur esofagus dalam penelitian ini dipengaruhi oleh hari perlakuan, volume PRF yang diberikan, ekspresi Mmp1 dan Timp1, serta kerusakan jaringan yang terjadi. Volume PRF terbaik untuk mencegah striktur ada pada volume 0,3 mL atau volume maksimal.

Kata kunci: *platelet-rich fibrin*, striktur esofagus, Mmp1, Mmp8, Timp1, model tikus

SUMMARY

ELUCIDATING PLATELET-RICH FIBRIN'S ROLE IN PREVENTION OF CAUSTIC INJURY-RELATED ESOPHAGEAL STRICTURE OF RAT MODEL: EVALUATING CHANGES OF Mmp1, Mmp8, Timp1 EXPRESSION

Scientific paper in the form of Dissertation, December 2024

Puspa Zuleika, supervised by oleh Prof. Dr. dr. Mgs. Irsan Saleh, M.Biomed.; Prof. dr. Krisna Murti, Sp.PA, Subsp.HLE(K), M.Biotech.Stud., Ph.D.; dan Dr. Iche Andriyani Liberty, S.K.M., M.Kes.

Doctorate in Biomedical Science, Faculty of Medicine, Sriwijaya University

Xxi + 205 pages, 25 tables, 76 figures, 10 attachments

Esophageal stricture is one of the most dangerous late complication of caustic injury in esophagus. Several studies have mentioned the roles of Mmp1, Mmp8, and Timp1 in the pathophysiological changes observed in esophageal stricture. Several prospective treatments to prevent esophageal stricture have been studied with varying costs and results. On the other hand, platelet-rich fibrin has been proven to be safe and efficacious to improve tissue regeneration. This study is thus aimed to prove PRF's role in esophageal stricture prevention in the rat model.

This study was conducted as an in vivo experimental study designed with post test-only control group in two phases: validation of a non-invasive rat model exposed to 0.1 mL of 10% NaOH via feeding tube and elucidation of the effect of varying volumes (0.1-0.3 mL) of administered PRF to prevent esophageal stricture on the previously-validated rat model. Platelet-rich fibrin originated from allograft rat donors. Differential expressions of Mmp1, Mmp8, and Timp1 were measured between 1-14 days through standardized immunohistochemical protocols while tissue damage and Stenosis Index were evaluated histologically. This study was conducted in the Animal House and Biotechnology Laboratory, Faculty of Medicine, Sriwijaya University. In addition, this study also involved Pathological

Anatomy Laboratory, Mohammad Hoesin General Hospital/Faculty of Medicine Sriwijaya University. This study was conducted between Januari and June, 2024. This study has been approved by the Ethical Research Committee, Faculty of Medicine, Sriwijaya University.

This study was able to validate the rat model. Further, this study discovered differential Mmp1 and Timp1 expression after administration while simultaneously discovering a non-significant impact of PRF administration on Mmp8 expression. This study also discovered reduction of histological tissue damage and Stenosis Index after PRF administration. Briefly, this study was able to discover the differential role of time, PRF volume administered, Mmp1 and Timp1 expression, and tissue damage in stricture formation. This study discovered that stricture prevention is most significant when PRF was administered at maximum volume (0.3 mL).

Keywords: platelet-rich fibrin, esophageal stricture, Mmp1, Mmp8, Timp1, rat model

HALAMAN PERSETUJUAN PUBLIKASI

Yang bertanda tangan di bawah ini:

Nama : Puspa Zuleika
NIM : 04013622227001
Judul : PENGARUH PEMBERIAN *PLATELET RICH FIBRIN* UNTUK MENCEGAH PEMBENTUKAN STRIKTUR ESOFAGUS AKIBAT CEDERA KAUSTIK BASA PADA HEWAN COBA TIKUS: TINJAUAN TERHADAP EKSPRESI Mmp1, Mmp8, Timp1

Memberikan izin kepada pihak Promotor/Ko-promotor dan Universitas Sriwijaya untuk mempublikasikan hasil penelitian saya untuk kepentingan akademik apabila dalam kurun waktu satu tahun tidak mempublikasikan karya penelitian saya. Dalam kondisi tersebut, saya setuju untuk menempatkan Promotor sebagai penulis korespondensi (*corresponding author*).

Demikian pernyataan ini saya buat dengan sadar dan tanpa ada paksaan dari pihak manapun.

Palembang, Desember 2024

Puspa Zuleika

DAFTAR ISI

HALAMAN JUDUL	i
HALAMAN PENGESAHAN.....	ii
HALAMAN PERSETUJUAN	iii
HALAMAN PERNYATAAN INTEGRITAS	iv
KATA PENGANTAR.....	v
ABSTRAK	vi
ABSTRACT	vii
RINGKASAN	viii
SUMMARY.....	x
HALAMAN PERSETUJUAN PUBLIKASI	xii
DAFTAR ISI.....	xiii
DAFTAR TABEL	xvi
DAFTAR GAMBAR.....	xvii
DAFTAR SINGKATAN	xix
BAB I PENDAHULUAN.....	1
1.1. Latar Belakang	1
1.2. Rumusan Masalah	3
1.3. Hipotesis Penelitian.....	3
1.4. Tujuan Penelitian	3
1.4.1. Tujuan Penelitian Umum.....	3
1.4.2. Tujuan Penelitian Khusus	4
1.5. Manfaat Penelitian	4
1.5.1. Manfaat Akademik	4
1.5.2. Manfaat di Bidang Penelitian dan pengembangan	4
1.5.3. Manfaat Klinis	5
1.6. Keterbaruan Penelitian.....	5
BAB II TINJAUAN PUSTAKA.....	6
2.1. Anatomi Esofagus	6
2.1.1. Anatomi Esofagus Manusia.....	6
2.1.1.1. Lapisan Histologis Esofagus	7
2.1.1.2. Konstriksi pada Esofagus	10
2.1.1.3. Pembagian Segmen Esofagus.....	12
2.1.1.4. Vaskularisasi Esofagus.....	13
2.1.2. Anatomi Esofagus Tikus	15
2.2. Cedera Kaustik Esofagus dan Struktur	19
2.2.1. Definisi	19
2.2.2. Penyebab Cedera Kaustik Esofagus	19
2.2.3. Derajat Kerusakan Esofagus.....	20
2.2.4. Model Hewan Coba	24
2.2.5. Gambaran Klinis Esofagitis Korosif pada Manusia	31
2.2.6. Dampak Esofagitis Korosif Secara Klinis	35
2.2.7. Respons Inflamasi pada Cedera Kaustik Esofagus.....	37
2.2.8. <i>Matrix Metalloproteinase</i> dan Struktur	40
2.2.9. Kaitan Regulasi Ekspresi Timp1, Mmp1, dan Mmp8	53

2.3. Proses Penyembuhan Luka	56
2.3.1. <i>Platelet-rich Fibrin</i>	58
2.4. <i>Platelet-rich Fibrin</i> dalam Perbaikan Jaringan	62
2.5. Efek Biologis <i>Platelet-rich Fibrin</i>	69
2.6. Peran Inflamasi dalam Struktur dan kerja <i>Platelet-rich Fibrin</i>	71
2.7. Kerangka Teori.....	78
2.8.Kerangka Konsep	79
BAB III METODE PENELITIAN.....	80
3.1. Desain Penelitian.....	80
3.2. Waktu dan Tempat Penelitian	80
3.3. Sampel Penelitian.....	80
3.4. Besar Sampel Penelitian.....	80
3.5. Kriteria Inklusi	81
3.6. Variabel Penelitian.....	81
3.6.1. Variabel Bebas.....	81
3.6.2. Variabel terikat	81
3.6.2.1. Fase Pertama Penelitian	81
3.6.2.2. Fase Kedua Penelitian	82
3.7. Definisi Operasional.....	83
3.8. Prosedur Penelitian.....	86
3.8.1. Etik Penelitian.....	86
3.8.2. Prosedur Pembuatan PRF	87
3.8.3. Prosedur Pembuatan Model Hewan untuk Cedera Esofagus	88
3.8.3.1. Prosedur Fase Pertama Menggunakan Sonde	89
3.8.3.2. Prosedur Fase Kedua	90
3.8.4. Pengendalian Nyeri.....	92
3.8.5. Alur Penelitian Fase Pertama.....	93
3.8.5. Alur Penelitian Fase Kedua	94
3.9. Pengumpulan Data	95
3.9.1. Histopatologi.....	95
3.9.1.1. Persiapan Preparat.....	95
3.9.1.2. Pengecatan Hematoksilin-Eosin.....	96
3.9.1.3. Pengecatan <i>Masson's Trichrome</i>	96
3.9.1.4. <i>Stenosis Index</i>	98
3.9.2. Imunohistokimia	100
3.9.2.1. Kuantifikasi dengan ImageJ	101
3.10. Analisis Data	103
BAB IV HASIL DAN PEMBAHASAN	104
4.1. Pembuatan Model Hewan Coba Esofagitik Korosif.....	105
4.2. Berat Badan Tikus.....	107
4.3. Dampak Pemberian PRF Terhadap Perubahan Ekspresi Mmp1.....	107
4.4. Dampak Pemberian PRF Terhadap Perubahan Ekspresi Mmp8.....	110
4.5. Dampak Pemberian PRF Terhadap Perubahan Ekspresi Timp1.....	114
4.6. Dampak Pemberian PRF Terhadap Derajat Kerusakan Esofagus Tikus Berdasarkan Histopatologi.....	117
4.6.1. Skor ISMC (<i>Increase in Submucosal Collagen</i>)	117

4.6.2. Skor DMM (<i>Damage to Muscularis Mucosae</i>)	120
4.6.3. Skor DCDTM (<i>Damage and Collagen Deposition in Tunica Muscularis</i>).....	122
4.6.4. Skor I (<i>Inflammation</i>)	124
4.6.5. Skor F (<i>Fibrosis</i>)	126
4.6.6. Skor N (<i>Necrosis</i>)	128
4.6.7. Skor Histologi Total	131
4.7. Stenosis Index.....	133
4.7.1. Regresi Non Linear Mixed Effects Terhadap <i>Stenosis Index</i>	136
4.8. Pembahasan.....	138
4.8.1. Pembuatan Model Hewan	138
4.8.2. Dampak Cedera Korosif Basa Kuat.....	140
4.8.3. Efektivitas PRF Dalam Mencegah Struktur Esofagus.....	145
4.8.4. Peran PRF pada Fase Hemostasis dan Inflamasi.....	146
4.8.5. Peran PRF pada Fase Hemostasis dan <i>Remodeling</i>	148
4.8.6. Hubungan Volume PRF Terhadap <i>Stenosis Index</i>	152
4.8.7. Keterbatasan Penelitian	156
BAB V SIMPULAN DAN SARAN.....	158
5.1. Simpulan	158
4.2. Saran.....	159
DAFTAR PUSTAKA	160

DAFTAR TABEL

Nomor	Judul	Halaman
	Tabel 2.1. Perbedaan Esofagus Tikus dan Manusia	18
	Tabel 2.2. Senyawa Kaustik yang Umum di Rumah.....	20
	Tabel 2.3. Derajat Kerusakan Esofagus Menurut Zargar	21
	Tabel 2.4. Cedera Esofagus Setelah Paparan Zat Kaustik.....	38
	Tabel 2.5. Sitokin dan Faktor Pertumbuhan yang Memengaruhi STAT	50
	Tabel 2.6. <i>Growth Factors</i> yang Terkandung di PRF	59
	Tabel 3.1. Definisi Operasional.....	83
	Tabel 3.2. Tabel Perlakuan	86
	Tabel 3.3. Pengendalian Nyeri Hewan Coba.....	92
	Tabel 3.4. Penilaian Histopatologi	99
	Tabel 4.1. Perbedaan Berat Badan Tikus Sebelum dan Sesudah Intervensi Penelitian Fase Pertama.....	105
	Tabel 4.2. Nilai <i>Stenosis Index</i> Lumen Esofagus Tikus Setelah Intervensi Penelitian Fase Pertama.....	106
	Tabel 4.3. Berat Badan Awal Tikus Sebelum Intervensi Penelitian Fase Kedua	107
	Tabel 4.4. Perbedaan Ekspresi Protein Mmp1 Antar Kelompok	108
	Tabel 4.5. Perbedaan Ekspresi Protein Mmp8 Antar Kelompok	111
	Tabel 4.6. Perbedaan Ekspresi Protein Timp1 Antar Kelompok	114
	Tabel 4.7. Perbedaan Skor ISMC Antar Kelompok	118
	Tabel 4.8. Perbedaan Skor DMM Antar Kelompok.....	121
	Tabel 4.9. Perbedaan Skor DCDTM Antar Kelompok	123
	Tabel 4.10. Perbedaan Skor I Antar Kelompok	125
	Tabel 4.11. Perbedaan Skor F Antar Kelompok.....	127
	Tabel 4.12. Perbedaan Skor N Antar Kelompok	129
	Tabel 4.13. Perbedaan Skor Histologi Total Antar Kelompok.....	132
	Tabel 4.14. Perbedaan <i>Stenosis Index</i> Antar Kelompok	134
	Tabel 4.15. Regresi Non Linear <i>Mixed Effects</i> Terhadap <i>Stenosis Index</i>	138

DAFTAR GAMBAR

Nomor	Judul	Halaman
	Gambar 2.1. Anatomi dan Histologi Esofagus	7
	Gambar 2.2. Dinding Esofagus	8
	Gambar 2.3. Kelenjar Submukosa Esofagus	9
	Gambar 2.4. Lapisan Otot Esofagus.....	10
	Gambar 2.5. Konstriksi Esofagus	11
	Gambar 2.6. Pembagian Esofagus.....	12
	Gambar 2.7. Arteri yang Memperdarahi Esofagus.....	13
	Gambar 2.8. Vena yang Menjadi Drainase Esofagus.....	14
	Gambar 2.9. Drainase Limfatik Esofagus	15
	Gambar 2.10. Esofagus Bawah Tikus dan Manusia.....	16
	Gambar 2.11. Anatomi <i>Gastroesophageal Junction</i> Tikus dan Manusia.....	17
	Gambar 2.12. Esofagus Setelah Perlakuan.....	22
	Gambar 2.13. Gambaran Endoskopi pada Hari ke-28.....	23
	Gambar 2.14. Fibrosis pada Struktur Esofagus	23
	Gambar 2. 15. Perubahan Histopatologi pada Struktur	24
	Gambar 2.16. Gambaran Esofagus Setelah Cedera Kaustik	25
	Gambar 2.17. Temuan Histopatologis pada BAPN dan <i>Prednisolone</i>	27
	Gambar 2.18. Temuan Histopatologis pada Model Tikus dengan Palifermin	28
	Gambar 2.19. Histopatologi Esofagus Setelah Pemberian GLP-2	29
	Gambar 2.20. Perbedaan Sitokin Proinflamatoris Plasma pada Cedera Kaustik ..	39
	Gambar 2.21. Skema Struktur MMP	41
	Gambar 2.22. Pengaruh Fibroblas dalam <i>Scar</i>	42
	Gambar 2.23. Jaras Persinyalan TGF- β	44
	Gambar 2.24. Patogenesis <i>Remodeling Pasca Eosinophilic Esophagitis</i>	45
	Gambar 2.25. Pembentukan Struktur pada <i>Eosinophilic Esophagitis</i>	46
	Gambar 2.26. Jaras MMPs dan TIMPs dalam Inflamasi.....	47
	Gambar 2.27. Jaras Persinyalan JAK/STAT	48
	Gambar 2.28. Interaksi Jaras JAK/STAT	49
	Gambar 2.29. Aktivasi Jaras MAPK	51
	Gambar 2.30. Berbagai Stimuli pada Jaras JNK/MAPK	52
	Gambar 2.31. Faktor yang Mengaktivasi MAPKKK.....	52
	Gambar 2.32. Peran MMPs dalam Inflamasi	54
	Gambar 2.33. Proses Penyembuhan Luka.....	56
	Gambar 2.34. Skema Pembuatan PRF.....	61
	Gambar 2.35. PRFM yang Dihasilkan dari PRP	62
	Gambar 2.36. Gambaran Histologis Hari ke-20	63
	Gambar 2.37. Temuan Histologis LLLT dan L-PRF pada Kalvaria	64
	Gambar 2.38. Gambaran Histologis Regenerasi <i>N. Ischiadicus</i>	65
	Gambar 2.39. Model Perforasi Membran Timpani	66

Gambar 2.40. Karakterisasi L-PRF dan F-PRF	70
Gambar 3.1. <i>Buffy Coat</i> Hasil Sentrifugasi Pertama dan Kedua	87
Gambar 3.2. <i>Liquid PRF</i> Siap Diberikan ke Tikus.....	88
Gambar 3.3. Pembuatan Cedera Kaustik dengan Sonde	89
Gambar 3.4. Diseksi dan Identifikasi Struktur Esofagus Tikus	90
Gambar 3.5. Pemberian PRF ke Tikus Melalui Sonde.....	91
Gambar 3.6. Alur Penelitian Fase Pertama.....	93
Gambar 3.7. Alur Penelitian Fase Kedua	94
Gambar 3.8. Gambaran Histologi Potongan Melintang Esofagus	98
Gambar 4.1. Perbandingan Lumen Esofagus Tikus yang Normal dan yang Mengalami Struktur Akibat Pemberian NaOH 10% 0,1 mL	106
Gambar 4.2. Perubahan Ekspresi Protein Mmp1 Berdasar Waktu.....	109
Gambar 4.3. Respons Ekspresi Mmp1 Terhadap Volume PRF	109
Gambar 4.4. Ekspresi Mmp1 pada Esofagus Tikus.....	110
Gambar 4.5. Perubahan Ekspresi Protein Mmp8 Seiring Waktu	112
Gambar 4.6. Respons Ekspresi Mmp8 Terhadap Volume PRF	112
Gambar 4.7. Ekspresi Mmp8 pada Esofagus Tikus.....	113
Gambar 4.8. Perubahan Ekspresi Protein Timp1 Seiring Waktu	115
Gambar 4.9. Respons Ekspresi Timp1 Terhadap Volume PRF	115
Gambar 4.10. Ekspresi Timp1 pada Esofagus Tikus	116
Gambar 4.11. Perubahan Skor ISMC Seiring Waktu.....	118
Gambar 4.12. Pengaruh Volume PRF Terhadap Skor ISMC.....	119
Gambar 4.13. Pengecatan <i>Gomori's Trichrome</i> pada Esofagus Tikus.....	121
Gambar 4.14. Perubahan Skor DMM Seiring Waktu.....	121
Gambar 4.15. Pengaruh Volume PRF Terhadap Skor DMM.....	121
Gambar 4.16. Perubahan Skor DCDTM Seiring Waktu	123
Gambar 4.17. Pengaruh Volume PRF Terhadap Skor DCDTM	124
Gambar 4.18. Perubahan Skor I Seiring Waktu	125
Gambar 4.19. Pengaruh Volume PRF Terhadap Skor I	126
Gambar 4.20. Perubahan Skor F Seiring Waktu	127
Gambar 4.21. Pengaruh Volume PRF Terhadap Skor F	128
Gambar 4.22. Perubahan Skor N Seiring Waktu.....	130
Gambar 4.23. Pengaruh Volume PRF Terhadap Skor N.....	130
Gambar 4.24. Perubahan Skor Histologi Total Seiring Waktu	132
Gambar 4.25. Pengaruh Volume PRF Terhadap Skor Histologi Total	133
Gambar 4.26. Perubahan <i>Stenosis Index</i> Seiring Waktu	135
Gambar 4.27. Pengaruh Volume PRF Terhadap <i>Stenosis Index</i>	135
Gambar 4.28. Hubungan Hari, Volume PRF, dan <i>Stenosis Index</i>	136

DAFTAR SINGKATAN

95%CI	: <i>95% confidence interval</i>
ABB	: <i>anorganic bovine bone</i>
ABC	: <i>alveolar blood clots</i>
ABH	: <i>alveolar bone height</i>
ABW	: <i>alveolar bone width</i>
ADMSC	: <i>adipose-derived mesenchymal stem cell</i>
AgNP	: nanopartikel perak
ALG	: alginat
AMI	: <i>acute myocardial infarction</i>
APC	: <i>argon plasma coagulation</i>
A-PRF	: <i>advanced-PRF</i>
ASC	: <i>adipose-derived stem cell</i>
BAPN	: <i>beta-aminopropionitrile</i>
BCP	: <i>biphasic calcium phosphate</i>
BD	: <i>biodegradable</i>
CaCl ₂	: kalsium klorida
cDNA	: <i>circular DNA</i>
CGF	: <i>concentrated growth factors</i>
CI	: <i>confidence interval</i>
CM	: <i>collagen membrane</i>
C-PRF	: <i>concentrated-PRF</i>
CRP	: <i>C-reactive protein</i>
CSBC	: <i>calcium-silicate based-cement</i>
CT	: <i>computed tomography</i>
CT-scan	: <i>computed tomography-scan</i>
CVC	: <i>central venous catheter</i>
DAMPs	: <i>damage-associated molecular patterns</i>
dB	: desibel
dkk.	: dan kawan-kawan
DMEM	: <i>Dulbecco's Modified Eagle Medium</i>
DROOL	: <i>drooling reluctance oropharynx other leukocytosis</i>
ECM	: <i>extracellular matrix</i>
EGF	: <i>epidermal growth factor</i>
ELISA	: <i>enzyme-linked immunosorbent assay</i>
EuropePMC	: <i>European PubMed Central</i>
FDG	: <i>fluoro-d-glucose</i>
FGF	: <i>fibroblast growth factor</i>
GADPH	: <i>glyceraldehyde 3-phosphate dehydrogenase</i>
GDNF	: <i>glial cell line-derived neurotrophic factor</i>
GLP-2	: <i>glucagon-like peptide-2</i>
H&E	: hematoksilin dan eosin
HA	: <i>hyaluronic acid</i>
HCl	: asam klorida
HDS	: <i>histopathological damage score</i>

HES	: hesperidin
HGF	: <i>human gingival fibroblast</i>
HL	: <i>hearing loss</i>
HR	: <i>hazard rasio</i>
IFN	: interferon
IGF	: <i>insulin-like growth factor</i>
IHK	: imunohistokimia
IL	: interleukin
iNOS	: <i>inducible nitric oxide synthase</i>
i-PRF	: <i>injectable-PRF</i>
kgBB	: kilogram berat badan
KOH	: kalium hidroksida
LA	: <i>lumen area</i>
LLLT	: <i>low-level diode laser radiation therapy</i>
L-PRF	: <i>leukocyte- and platelet-rich fibrin</i>
LPS	: lipopolisakarida
LV	: <i>left ventricle</i>
Ly-PRF	: <i>lysate-PRF</i>
MAPK	: <i>mitogen-activated protein kinase</i>
MAPKK	: MAPK kinase
MAPKKK	: MAPKK kinase
MCP	: <i>monocyte chemoattractant protein</i>
MD	: <i>mean difference</i>
MDVP	: <i>multidimensional voice program</i>
MMC	: mitomycin C
MMP	: <i>matrix metalloproteinase</i>
MMPs	: <i>matrix metalloproteinases</i>
MRI	: <i>magnetic resonance imaging</i>
mRNA	: <i>messenger ribonucleic acid</i>
MSC	: <i>mesenchymal stem cell</i>
N.IX	: nervus kranialis IX (nervus glosofaringeus)
N.X	: nervus kranialis X (nervus vagus)
NaCl	: <i>natrium klorida</i>
NADPH	: <i>nicotinamide adenine dinucleotide phosphate</i>
NaOH	: <i>natrium hidroksida</i>
NF-kB	: <i>Nuclear factor kappa-light-chain-enhancer of activated B cells</i>
NGF	: <i>nerve growth factor</i>
NPV	: <i>negative predictive value</i>
PAMPs	: <i>pathogen-associated molecular patterns</i>
PDGF	: <i>platelet-derived growth factor</i>
PET	: <i>positron emission tomography</i>
PEWS	: <i>pediatric early warning system</i>
PGE2	: <i>prostaglandin E2</i>
pH	: <i>power of hydrogen, potential of hydrogen</i>
PLG	: <i>polylactic glicolic acid</i>
PPP	: <i>platelet-poor plasma</i>

PPRF	: <i>pure PRF</i>
PPV	: <i>positive predictive value</i>
PRF	: <i>platelet-rich fibrin</i>
PRFM	: <i>PRF matrix</i>
PRFr	: <i>PRF releasate</i>
PRGF	: <i>platelet-rich growth factors</i>
PRISMA	: <i>Preferred Reporting Items for Systematic reviews and Meta-Analyses</i>
PRP	: <i>platelet-rich plasma</i>
RDW	: <i>red cell distribution width</i>
RNA	: <i>ribonucleic acid</i>
ROS	: <i>reactive oxygen species</i>
RT-PCR	: <i>reverse transcriptase-polymerase chain reaction</i>
RT-qPCR	: <i>reverse transcriptase-quantitative polymerase chain reaction</i>
SD	: <i>Sprague-Dawley</i>
SEMS	: <i>self-expanding metal stent</i>
SEPS	: <i>self-expanding plastic stent</i>
SI	: <i>stenosis index</i>
TA	: <i>total area</i>
TGF	: <i>tumor growth factor</i>
TNF	: <i>tumor necrosis factor</i>
TQ	: <i>thymoquinone</i>
Treg	: <i>sel T regulator</i>
TTS	: <i>through-the-scope balloon</i>
TUNEL	: <i>terminal deoxynucleotidyl transferase-mediated dUTP nick end labeling</i>
VEGF	: <i>vascular endothelial growth factor</i>
WBC	: <i>white blood cell</i>

BAB I

PENDAHULUAN

1.1. Latar Belakang

Cedera kaustik pada esofagus adalah cedera esofagus yang diakibatkan oleh tertelannya zat kimia kaustik yang bersifat asam atau basa.¹ Zat kaustik yang paling sering menyebabkan cedera esofagus karena tertelan adalah basa kuat ($\text{pH} \leq 12$) atau asam kuat ($\text{pH} \geq 2$). Kerusakan jaringan akibat zat basa lebih berat karena proses saponifikasi yang terjadi bersifat tidak kedap air sehingga zat alkali dapat memenetrasi lebih dalam ke jaringan di bawahnya.²

American Association of Poison Control Centers (AAPCC) pada tahun 2013 menyatakan bahwa terdapat hampir 60 ribu kasus paparan zat kaustik, yang terdiri dari 48 ribu kasus akibat paparan zat pemutih dalam rumah ($\text{pH } 9-11$), 7.500 kasus akibat paparan zat asam, dan 4.000 kasus akibat paparan basa.³ Sebanyak 48% kasus di antaranya terjadi pada anak berusia di bawah lima tahun dan 25% dari kasus anak disebabkan oleh zat kimia kaustik yang terdapat di dalam kosmetik, produk perawatan tubuh, atau cairan pembersih rumah tangga. Sebanyak 75% dari 1.173 kematian yang terjadi akibat cedera kaustik pada remaja dan dewasa terjadi akibat kesengajaan dan melibatkan ingesti zat kaustik yang terdapat dalam cairan pembersih rumah tangga dalam jumlah dan konsentrasi yang tinggi.^{4,5}

Benign esophageal stricture adalah komplikasi dari fibrosis esofagus yang disebabkan oleh berbagai cedera esofagus, seperti *gastroesophageal reflux*, radioterapi, konsumsi zat korosif, esofagitis eosinofilik, dan reseksi esofagus parsial.^{6,7} Fibrosis esofagus terjadi akibat inflamasi merangsang aktivasi miofibroblas dan produksi protein matriks ekstraseluler, termasuk kolagen, secara berlebih.⁷ Miofibroblas, selain mengekspresikan protein otot polos juga bersifat kontraktile. Kontraksi sel-sel ini berkontribusi pada patogenesis penyakit dan kegagalan jaringan. Proses perbaikan menghasilkan deposisi substansial komponen matriks ekstraseluler saat jaringan normal tergantikan dengan jaringan parut

permanen. Oleh karena itu, mediator seluler utama fibrosis adalah transformasi fibroblas-miofibroblas dan disposisi komponen matriks ekstraseluler.^{7,8}

Penelitian Koval dkk. (2018) menemukan bahwa proses penyembuhan luka bakar akibat zat kaustik asam secara signifikan dipengaruhi oleh Mmps dan Timps. Penelitian ini menyatakan bahwa Mmp1, Mmp2, Mmp3, Mmp8, Mmp9, dan Timp1 berpengaruh terhadap proses penyembuhan luka baik normal maupun patologis pada mukosa esofagus akibat zat kaustik asam.⁹ Senyawa Mmp1 disintesis oleh fibroblas, kondrosit, makrofag, keratinosit, sel endotel dan osteoblas dan memiliki aktivitas yang signifikan pada lingkungan sekitar luka.^{9,10} Protein Mmp8 berperan dalam fase awal degradasi matriks ekstraseluler karena Mmp8 menjadi enzim kunci dari tahap awal penghancuran matriks ekstraseluler.¹¹ Melihat proses yang terjadi selama proses penyembuhan jaringan, kesetimbangan antara Timp1, Mmp1, dan Mmp8 sangat dipengaruhi oleh inflamasi yang terjadi pada jaringan dan menjadi topik pembahasan yang masih belum banyak diteliti.

Pembentukan striktur adalah komplikasi lambat yang paling sering terjadi pada cedera kaustik esofagus dan dinilai dengan menggunakan skor klasifikasi Zargar $\geq 2A$, konsentrasi senyawa basa yang tertelan, meningkatnya hitung jenis sel darah putih, serta menurunnya rasio protrombin.¹² Sebanyak 71% pasien dengan derajat cedera 2B akan mengalami striktur esofagus; pada pasien dengan derajat Zargar 3, hampir seluruhnya mengalami striktur esofagus.¹³ Striktur esofagus akibat cedera kaustik dapat memberikan gejala disfagia dan rasa tertekan pada substernum yang diikuti dengan rasa mual dan muntah setelah makan, serta penurunan berat badan akibat asupan yang kurang. Kondisi ini dapat terjadi sejak minggu kelima sampai minggu keenam setelah tertelan zat kaustik.¹⁴

Berbagai penelitian telah dilakukan dalam bidang pencegahan striktur esofagus, melibatkan eritropoietin, asam retinoat, ibuprofen, vitamin E, aseetilsistein, *dimethyl sulfoxide*, dan lain-lain. Akan tetapi hingga saat ini protokol tetap untuk penatalaksanaan pencegahan striktur akibat cedera kaustik di esofagus masih belum ditetapkan. Beberapa hambatan yang terjadi dikarenakan sulitnya ketersediaan zat tersebut, toksisitas, biaya yang tinggi, dan keamanan yang masih diragukan. Choukroun dkk. (2001) memperkenalkan konsentrat trombosit yang

disebut *Platelet-Rich Fibrin* (PRF), dapat dibuat tanpa antikoagulan apapun, dan mengandung sejumlah besar sel imun.¹⁵ Beberapa penelitian telah menyatakan PRF saat ini relevan dalam perbaikan jaringan dan beberapa komponen PRF berperan dalam penyembuhan luka dan memiliki kemungkinan untuk penerapan dalam pengobatan regeneratif.¹⁶ Maka dari itu PRF yang kaya dengan *growth factor*, leukosit, sitokin/kemokin, serta memiliki sifat *autologous* dapat dipertimbangkan untuk digunakan sebagai zat atau agen untuk mencegah terjadinya striktur esofagus akibat cedera kaustik pada esofagus.

1.2. Rumusan Masalah

1. Bagaimana pembuatan model striktur basa kuat pada tikus tanpa melalui laparotomi?
2. Apakah pemberian PRF secara topikal pada lumen esofagus dapat mencegah pembentukan striktur esofagus akibat cedera kaustik basa pada hewan coba tikus melalui ekspresi protein Mmp1, Mmp8, dan Timp1?

1.3. Hipotesis Penelitian

Pemberian PRF dapat mencegah pembentukan striktur esofagus akibat cedera kaustik basa pada hewan coba tikus melalui ekspresi protein Mmp1, Mmp8, dan Timp1.

1.4. Tujuan Penelitian

1.4.1. Tujuan Penelitian Umum

1. Menentukan metode pembuatan model tikus yang paling sesuai untuk pembuatan striktur esofagus akibat cedera kaustik basa kuat.
2. Membuktikan pengaruh pemberian PRF untuk mencegah pembentukan striktur esofagus akibat cedera kaustik basa kuat pada hewan coba tikus melalui ekspresi Mmp1, Mmp8, dan Timp1.

1.4.2. Tujuan Penelitian Khusus

1. Membuat model esofagitis korosif akibat trauma senyawa kaustik basa kuat pada tikus menggunakan sonde.
2. Menganalisis derajat kerusakan esofagus tikus yang mendapat perlakuan cedera kaustik basa kuat secara histopatologi.
3. Menganalisis perubahan ekspresi Mmp1, Mmp8, dan Timp1 pada jaringan esofagus akibat cedera kaustik basa kuat pada jaringan esofagus tikus.
4. Menganalisis dampak pemberian PRF terhadap perubahan histologis jaringan esofagus tikus model cedera kaustik basa kuat selama proses penyembuhan jaringan esofagus.
5. Menganalisis dampak pemberian PRF terhadap perubahan ekspresi Mmp1, Mmp8, dan Timp1 pada jaringan esofagus tikus model cedera kaustik basa kuat selama proses penyembuhan jaringan esofagus.
6. Menganalisis dampak variasi volume PRF terhadap proses penyembuhan esofagus tikus model cedera kaustik basa kuat.
7. Menganalisis dampak variasi volume PRF terhadap pencegahan striktur esofagus pada tikus model cedera kaustik basa kuat.

1.5. Manfaat Penelitian

1.5.1. Manfaat Akademik

Secara akademis, penelitian ini diharapkan mampu menjelaskan mekanisme kerja PRF dalam mencegah striktur esofagus akibat cedera kaustik basa kuat.

1.5.2. Manfaat di Bidang Penelitian dan Pengembangan

Untuk pengembangan penelitian, model hewan yang dikembangkan dalam penelitian ini diharapkan dapat menjadi dasar untuk penelitian lebih lanjut mengenai mekanisme kerja PRF dalam pencegahan striktur esofagus akibat cedera kaustik basa kuat.

1.5.3. Manfaat Klinis

Secara klinis, penelitian ini diharapkan dapat menjadi dasar untuk pemanfaatan PRF dalam mencegah striktur esofagus akibat cedera kaustik basa kuat di RSUP Dr. Mohammad Hoesin secara khusus dan *center* lain secara umum.

1.6. Keterbaruan Penelitian

Potensi kebaruan penelitian adalah dalam metode pembuatan model tikus yang bersifat non-invasif untuk striktur esofagus akibat cedera kaustik basa kuat serta mekanisme kerja PRF dalam pencegahan striktur esofagus melalui efeknya terhadap ekspresi Mmp1, Mmp8, dan Timp1. Potensi ini muncul karena:

1. Model tikus untuk striktur esofagus akibat cedera kaustik basa kuat yang paling umum digunakan saat ini bersifat invasif melalui laparotomi.
2. Peneliti masih belum menemukan publikasi penelitian mengenai pencegahan striktur esofagus akibat cedera kaustik basa kuat dengan tatalaksana PRF dan volume PRF yang paling efektif untuk pencegahan striktur esofagus akibat cedera kaustik basa kuat
3. Peneliti masih belum menemukan publikasi penelitian yang menilai ekspresi protein Mmp1, Mmp8, dan Timp1 dengan menggunakan imunohistokimia, derajat kerusakan histopatologi dan *Stenosis Index* setelah pemberian PRF untuk pencegahan striktur esofagus akibat cedera kuat

DAFTAR PUSTAKA

1. Kalipatnapu S, Reddipogu J, George S, Abraham V, Samarasam I. Corrosive injuries of the upper gastrointestinal tract: A review of management practices. *Current Medical Issues*. 2018;16(3):92.
2. Seref K, Sonmez K, Gulburun MA, Ekinci O, Oge CB, Gulbahar O, et al. Protective Effects of Contractubex® on Stricture Formation after Experimental Corrosive Esophageal Burns in Rats. *Arch Med Res*. 2020 Oct;51(7):664–9.
3. Chirica M, Bonavina L, Kelly MD, Sarfati E, Cattan P. Caustic ingestion. *The Lancet*. 2017 May;389(10083):2041–52.
4. Mowry JB, Spyker DA, Brooks DE, Mcmillan N, Schauben JL. 2014 annual report of the American association of poison control centers National Poison Data System (NPDS): 32nd Annual Report. *Clin Toxicol*. 2015;53(10):962–1147.
5. Kurowski JA, Kay M. Caustic Ingestions and Foreign Bodies Ingestions in Pediatric Patients. *Pediatr Clin North Am*. 2017;64(3):507–24.
6. Mizusawa T, Kobayashi M, Terai S. Radial incision and cutting for refractory benign esophageal stricture. *Digestive Endoscopy*. 2019;31(2):e46–7.
7. Zhang Y, Wang Q, Xu Y, Sun J, Ding Y, Wang L, et al. Mitomycin C Inhibits Esophageal Fibrosis by Regulating Cell Apoptosis and Autophagy via lncRNA-ATB and miR-200b. *Front Mol Biosci*. 2021;8(May):1–10.
8. Bülow RD, Boor P. Extracellular Matrix in Kidney Fibrosis: More Than Just a Scaffold. *Journal of Histochemistry and Cytochemistry*. 2019;67(9):643–61.
9. Koval T, Ishchuk T, Grebinyk D, Raetska YB, Sokur O, Savchuk O, et al. Matrix metalloproteinase functioning in case of esophagus acid burn. *Biomedical Research*. 2018;29(16):3169–73.
10. Gioia M, Monaco S, Van Den Steen PE, Sbardella D, Grasso G, Marini S, et al. The Collagen Binding Domain of Gelatinase A Modulates Degradation of Collagen IV by Gelatinase B. *J Mol Biol*. 2009;386(2):419–34.
11. Tsousi A, Witte E, Witte K, Röwert-Huber HJ, Volk HD, Sterry W, et al. MMP8 Is Increased in Lesions and Blood of Acne Inversa Patients: A Potential Link to Skin Destruction and Metabolic Alterations. *Mediators Inflamm*. 2016;2016.
12. Le Naoures P, Hamy A, Lerolle N, Métivier E, Lermite E, Venara A. Risk factors for symptomatic esophageal stricture after caustic ingestion-A retrospective cohort study. *Diseases of the Esophagus*. 2017;30(6):1–6.
13. Hall AH, Jacquemin D, Henny D, Mathieu L, Josset P, Meyer B. Corrosive substances ingestion: a review. *Crit Rev Toxicol*. 2019;49(8):637–69.
14. Lusong MAA De, Timbol ABG, Tuazon DJS. Management of esophageal caustic injury. *World J Gastrointest Pharmacol Ther*. 2017;8(2):90.

15. Choukroun J, Miron RJ. Platelet Rich Fibrin: A Second-Generation Platelet Concentrate. *Platelet Rich Fibrin in Regenerative Dentistry: Biological Background and Clinical Indications.* 2017;1–14.
16. Pavlovic V, Ceric M, Jovanovic V, Trandafilovic M, Stojanovic P. Platelet-rich fibrin: Basics of biological actions and protocol modifications. *Open Medicine.* 2021 Mar 22;16(1):446–54.
17. Ovalle WK, Nahirney PC. *Netter's Essential Histology: With Correlated Histopathology.* Third Edit. Philadelphia: Elsevier Inc.; 2021.
18. Young B, O'Dowd G, Woodford P. *Wheater's Functional Histology: A Text and Colour Atlas.* Sixth. Philadelphia: Elsevier Inc.; 2014.
19. Agur AMR, Dalley AF. *Moore's Essential Clinical Anatomy.* Sixth. Philadelphia: Wolters kluwer; 2019.
20. Standring S, editor. *Gray's Anatomy: The Anatomical Basis of Clinical Practice.* 42nd ed. London: Elsevier Ltd.; 2021.
21. Snell RS. *Clinical Anatomy by Systems.* Philadelphia: Lippincott Williams & Wilkins; 2006.
22. Yeo CJ. *Shackelford's Surgery of the Alimentary Tract, 2 Volume Set.* Eighth edi. Yeo CJ, DeMeester SR, Fleshman JW, Matthews JB, McFadden DW, editors. *Shackelford's Surgery of the Alimentary Tract: 2 Volume Set.* Philadelphia: Elsevier Inc.; 2019.
23. Netter FH. *Netter Atlas of Human Anatomy: A Systems Approach.* Eight. Philadelphia: Elsevier Inc.; 2023.
24. Maynard RL, Downes N. Front-matter. *Anatomy and Histology of the Laboratory Rat in Toxicology and Biomedical Research.* London, UK: Academic Press; 2019. i–iii.
25. Treuting PM, Dintzis SM. Comparative Anatomy and Histology: A Mouse and Human Atlas. In: *Comparative Anatomy and Histology: A Mouse and Human Atlas.* Academic Press; 2012. p. 46–51.
26. Flint PW, Haughey BH, Lund VJ, Niparko JK, Robbins KT, Thomas JR, et al., editors. *Cummins Otolaryngology: Head and Neck Surgery.* Sixth Edit. Philadelphia: Elsevier Inc.; 2015.
27. Elkaramany M. An overview of corrosive injury of the upper gastrointestinal tract: Discussion of types, clinical evaluation, and management procedures. *Advances in Digestive Medicine.* 2018;5(4):115–20.
28. Hoffman RS, Burns MM, Gosselin S. Ingestion of Caustic Substances. *New England Journal of Medicine.* 2020;382(18):1739–48.
29. Bakan V, Çiralik H, Kartal S. A corrosive oesophageal burn model in rats: Double-lumen central venous catheter usage. *African Journal of Paediatric Surgery.* 2015;12(4):247–50.
30. Li L, Itani MI, Salimian KJ, Li Y, Gutierrez OB, Hu H, et al. A patient-like swine model of gastrointestinal fibrotic strictures for advancing therapeutics. *Sci Rep.* 2021;11(1):13344.
31. Yang K, Li X, Zhou B, Zhu Y, Cheng Y. Sodium hydroxide-induced esophageal stricture via an endoscopic injection needle : a novel rabbit model of corrosive injury. *Journal of Interventional Medicine.* 2018;1(1):5–8.

32. Osman M, Russell J, Shukla D, Moghadamfalahi M, Granger DN. Responses of the murine esophageal microcirculation to acute exposure to alkali, acid, or hypochlorite. *J Pediatr Surg.* 2008;43(9):1672–8.
33. Aciksari K, Yanar HT, Hepgul G, Ertekin C, Gunay K, Ozcelik DN, et al. 183 Effect of Beta-aminopropionitrile on the Prevention of Fibrosis in Alkali Burns of the Esophagus: An Experimental Study. *Ann Emerg Med.* 2012;60(4):S66.
34. Karaca G, Aydin O, Pehlivani F, Altunkaya C, Uzun H, Güler O. Effectiveness of thymoquinone, zeolite, and platelet-rich plasma in model of corrosive oesophagitis induced in rats. *Ann Surg Treat Res.* 2017;92(6):396–401.
35. Numanoğlu KV, Tatlı D, Bektaş S, Er E. Efficacy of keratinocyte growth factor (palifermin) for the treatment of caustic esophageal burns. *Exp Ther Med.* 2014;8(4):1087–91.
36. Kantarcioğlu M, Caliskan B, Demirci H, Karacalioglu O, Kekilli M, Polat Z, et al. The efficacy of mesenchymal stem cell transplantation in caustic esophagus injury: An experimental study. *Stem Cells Int.* 2014;2014:4–6.
37. Orozco-Perez J, Aguirre-Jauregui O, Salazar-Montes AM, Sobrevilla-Navarro AA, Lucano-Landeros MS, Armendáriz-Borunda J. Pirfenidone prevents rat esophageal stricture formation. *Journal of Surgical Research.* 2015 Apr;194(2):558–64.
38. Tekin M, Topaloğlu N, Küçük A, Deniz M, Yıldırım S, Erdem H. Protective effect of glucagon-like peptide-2 in experimental corrosive esophagitis. *Diseases of the Esophagus.* 2015;28(3):258–61.
39. Anayurt M, Karaman A, Balcı Ö, Özgürer İF, Karaman İ. The effects of hesperidin on stricture formation in corrosive esophageal burns: an experimental study. *Esophagus.* 2022;19(1):189–96.
40. Türkyılmaz Z, Sönmez K, Karabulut R, Gülbahar Ö, Poyraz A, Sancak B, et al. Mitomycin C decreases the rate of stricture formation in caustic esophageal burns in rats. *Surgery.* 2009 Feb;145(2):219–25.
41. Okugbo S, Anyanhun G, Efobi C, Okugbo O. Presentation and management outcome of childhood corrosive oesophageal injury in Benin City. *African Journal of Paediatric Surgery.* 2020;17(3–4):74–8.
42. Kayamba V, Sinkala E, Mwanamakondo S, Soko R, Kawimbe B, Amadi B, et al. Trends in upper gastrointestinal diagnosis over four decades in Lusaka, Zambia: A retrospective analysis of endoscopic findings. *BMC Gastroenterol.* 2015;15(1):1.
43. Gelu-Simeon M, Chuong AP, Saliba F, Thiery G, Laurent M, Vilain C, et al. Submucosal hematoma: A new distinctive sign during emergency upper digestive endoscopy for ammonia ingestion. *BMC Gastroenterol.* 2018;18(1):1–7.
44. Bahrami-Motlagh H, Hadizadeh-Neisanghalb M, Peyvandi H. Diagnostic accuracy of computed tomography scan in detection of upper gastrointestinal tract injuries following caustic ingestion. *Arch Acad Emerg Med.* 2019;7(1):1–5.

45. Hollenbach M, Tünnemann J, Struck MF, Feisthammel J, Schlosser T, Schaumburg T, et al. Endoscopic findings and outcome in caustic ingestion of acidic and alkaline agents in adults. *Medicine.* 2019 Aug 27;98(35):e16729.
46. Mohammadi AB, Zaare Nahandi M, Ostadi A, Ghorbani A, Hallaj S. Endoscopic, laboratory, and clinical findings and outcomes of caustic ingestion in adults; a retrospective study. *Gastroenterol Hepatol Bed Bench.* 2022;15(1):59–65.
47. Ekpe EE, Ette V. Morbidity and Mortality of Caustic Ingestion in Rural Children: Experience in a New Cardiothoracic Surgery Unit in Nigeria. *ISRN Pediatr.* 2012;2012:1–4.
48. Grey NEO, Malone LDJ, Miller AL, Carroll HF, Khalaf RT, Kramer RE, et al. Magnetic resonance imaging findings following button battery ingestion. *Pediatr Radiol.* 2021;51(10):1856–66.
49. Cheng HT, Seak CJ, Cheng CC, Chen TH, Sung CM, Kang SC, et al. Profiling of inflammatory cytokines in patients with caustic gastrointestinal tract injury. *PLoS One.* 2021;16(11 November):1–12.
50. Aydin E, Beser O, Sazak S, Duras E. Role of RDW in Prediction of Burn after Caustic Substance Ingestion. *Children.* 2017 Dec 29;5(1):5.
51. Sharif AF, Gameel DEG El, Abdo SAEF, Elgebally EI, Fayed MM. Evaluation of Pediatric Early Warning System and Drooling Reluctance Oropharynx Others Leukocytosis scores as prognostic tools for pediatric caustic ingestion: a two-center, cross-sectional study. *Environmental Science and Pollution Research.* 2022;29(4):5378–95.
52. Youn BJ, Kim WS, Cheon JE, Kim WY, Shin SM, Kim IO, et al. Balloon dilatation for corrosive esophageal strictures in children: Radiologic and clinical outcomes. *Korean J Radiol.* 2010;11(2):203–10.
53. Gvalani AK, Deolekar S, Gandhi J, Dalvi A. Antesternal Colonic Interposition for Corrosive Esophageal Stricture. *Indian Journal of Surgery.* 2014;76(1):56–60.
54. Canena JMT, Liberato MJA, Rio-Tinto RAN, Pinto-Marques PM, Romão CMM, Coutinho AVMP, et al. A comparison of the temporary placement of 3 different self-expanding stents for the treatment of refractory benign esophageal strictures: a prospective multicentre study. *BMC Gastroenterol.* 2012;12.
55. Falk GW, Buttar NS, Foster NR, Ziegler KLA, Demars CJ, Romero Y, et al. A combination of esomeprazole and aspirin reduces tissue concentrations of prostaglandin E2 in patients with Barrett's esophagus. *Gastroenterology.* 2012;143(4):917-926.e1.
56. Isa HMA, Hasan KA, Ahmed HY, Mohamed AM. Efficacy and Safety of Endoscopic Esophageal Dilatation in Pediatric Patients with Esophageal Strictures. *International Journal of Pediatrics (United Kingdom).* 2021;2021.
57. Daniel P, Samanta J, Gulati A, Gupta P, Muktesh G, Sinha SK, et al. Can high-frequency mini-probe endoscopic ultrasonography predict outcome of endoscopic dilation in patients with benign esophageal strictures? *Endosc Int Open.* 2020;08(10):E1371–8.

58. Mu HW, Chen CH, Yang KW, Pan CS, Lin CL, Hung DZ. The prevalence of esophageal cancer after caustic and pesticide ingestion: A nationwide cohort study. *PLoS One.* 2020;15(12 December):1–11.
59. Yussof SJM, Omar E, Pai DR, Sood S. Cellular events and biomarkers of wound healing. *Indian Journal of Plastic Surgery.* 2012 May 27;45(02):220–8.
60. Page-McCaw A, Ewald AJ, Werb Z. Matrix metalloproteinases and the regulation of tissue remodelling. *Nat Rev Mol Cell Biol.* 2007;8(3):221–33.
61. Rohani MG, Parks WC. Matrix remodeling by MMPs during wound repair. *Matrix Biology.* 2015;44–46:113–21.
62. Loffek S, Schilling O, Franzke CW. Biological role of matrix metalloproteinases: a critical balance. *European Respiratory Journal.* 2011 Jul 1;38(1):191–208.
63. Hadler-Olsen E, Fadnes B, Sylte I, Uhlin-Hansen L, Winberg JO. Regulation of matrix metalloproteinase activity in health and disease. *FEBS Journal.* 2011;278(1):28–45.
64. Gauglitz GG. Textbook on Scar Management. Textbook on Scar Management. 2020.
65. Rieder F, Biancani P, Harnett K, Yerian L, Falk GW. Inflammatory mediators in gastroesophageal reflux disease: impact on esophageal motility, fibrosis, and carcinogenesis. *Am J Physiol Gastrointest Liver Physiol.* 2010 May;298(5):G571–81.
66. Barret M, Beye B, Leblanc S, Beuvon F, Chaussade S, Batteux F, et al. Systematic review: the prevention of oesophageal stricture after endoscopic resection. *Aliment Pharmacol Ther.* 2015 Jul;42(1):20–39.
67. Cheng E, Souza RF, Spechler SJ. Tissue remodeling in eosinophilic esophagitis. *Am J Physiol Gastrointest Liver Physiol.* 2012;303(11).
68. Khan S, Guo X, Liu T, Iqbal M, Jiang K, Zhu L, et al. An Update on Eosinophilic Esophagitis: Etiological Factors, Coexisting Diseases, and Complications. *Digestion.* 2021;102(3):342–56.
69. Doyle AD, Masuda MY, Kita H, Wright BL. Eosinophils in Eosinophilic Esophagitis: The Road to Fibrostenosis is Paved With Good Intentions. *Front Immunol.* 2020;11(December):1–13.
70. Abdulnour-Nakhoul SM, Al-Tawil Y, Gyftopoulos AA, Brown KL, Hansen M, Butcher KF, et al. Alterations in junctional proteins, inflammatory mediators and extracellular matrix molecules in eosinophilic esophagitis. *Clinical Immunology.* 2013;148(2):265–78.
71. Muir AB, Wang JX, Nakagawa H. Epithelial-stromal crosstalk and fibrosis in eosinophilic esophagitis. *J Gastroenterol.* 2019;54(1):10–8.
72. Beppu L, Yang T, Luk M, Newbury RO, Palmquist J, Dohil R, et al. MMPs-2 and-14 are elevated in eosinophilic esophagitis and reduced following topical corticosteroid therapy. *J Pediatr Gastroenterol Nutr.* 2015;61(2):194–9.
73. Runge TM, Eluri S, Woosley JT, Shaheen NJ, Dellon ES. Control of inflammation decreases the need for subsequent esophageal dilation in

- patients with eosinophilic esophagitis. Diseases of the Esophagus. 2017;30(7):1–7.
74. Cabral-Pacheco GA, Garza-Veloz I, Castruita-De la Rosa C, Ramirez-Acuña JM, Perez-Romero BA, Guerrero-Rodriguez JF, et al. The Roles of Matrix Metalloproteinases and Their Inhibitors in Human Diseases. *Int J Mol Sci.* 2020 Dec 20;21(24):9739.
 75. Khokha R, Murthy A, Weiss A. Metalloproteinases and their natural inhibitors in inflammation and immunity. *Nat Rev Immunol.* 2013;13(9):649–65.
 76. Hu X, li J, Fu M, Zhao X, Wang W. The JAK/STAT signaling pathway: from bench to clinic. Vol. 6, *Signal Transduction and Targeted Therapy*. Springer Nature; 2021.
 77. Hu Q, Bian Q, Rong D, Wang L, Song J, Huang HS, et al. JAK/STAT pathway: Extracellular signals, diseases, immunity, and therapeutic regimens. Vol. 11, *Frontiers in Bioengineering and Biotechnology*. Frontiers Media S.A.; 2023.
 78. Fey D, Croucher DR, Kolch W, Kholodenko BN. Crosstalk and signaling switches in mitogen-activated protein kinase cascades. *Front Physiol.* 2012;3 SEP.
 79. Marx F, Training O, Training P, Darin C, Training RO, Kimberly M, et al. Molecular Biology of the Cell. Igars 2014. 2014. 1–5 p.
 80. Fanjul-Fernández M, Folgueras AR, Cabrera S, López-Otín C. Matrix metalloproteinases: Evolution, gene regulation and functional analysis in mouse models. *Biochim Biophys Acta Mol Cell Res.* 2010;1803(1):3–19.
 81. Cui J, Zhang M, Zhang YQ, Xu ZH. JNK pathway: Diseases and therapeutic potential. *Acta Pharmacol Sin.* 2007;28(5):601–8.
 82. Weston CR, Davis RJ. The JNK signal transduction pathway. *Curr Opin Cell Biol.* 2007;19(2):142–9.
 83. Davis RJ. Signal transduction by the JNK group of MAP kinases. *Cell.* 2000;103(2):239–52.
 84. Cho JW, Cho SY, Lee SR, Lee KS. Onion extract and quercetin induce matrix metalloproteinase-1 in vitro and in vivo. *Int J Mol Med.* 2010 Jan 22;25(3):521–7.
 85. Chen J, Qin S, Liu S, Zhong K, Jing Y, Wu X, et al. Targeting matrix metalloproteinases in diabetic wound healing. *Front Immunol.* 2023;14(February):1–19.
 86. Nee LE, McMorrow T, Campbell E, Slattery C, Ryan MP. TNF- α and IL-1 β -mediated regulation of MMP-9 and TIMP-1 in renal proximal tubular cells. Vol. 66, *Kidney International*. 2004. p. 1376–86.
 87. Bertrand-Philippe M, Ruddell RG, Arthur MJP, Thomas J, Mungalsingh N, Mann DA. Regulation of tissue inhibitor of metalloproteinase 1 gene transcription by RUNX1 and RUNX2. *Journal of Biological Chemistry.* 2004;279(23):24530–9.
 88. Arpino V, Brock M, Gill SE. The role of TIMPs in regulation of extracellular matrix proteolysis. *Matrix Biology.* 2015;44–46:247–54.

89. Gutiérrez-Fernández A, Inada M, Balbín M, Fueyo A, Pitiot AS, Astudillo A, et al. Increased inflammation delays wound healing in mice deficient in collagenase-2 (MMP-8). *The FASEB Journal.* 2007;21(10):2580–91.
90. Naim A, Baig MS. Matrix metalloproteinase-8 (MMP-8) regulates the activation of hepatic stellate cells (HSCs) through the ERK-mediated pathway. *Mol Cell Biochem.* 2020 Apr 27;467(1–2):107–16.
91. Vincenti MP, Brinckerhoff CE. Transcriptional regulation of collagenase (MMP-1, MMP-13) genes in arthritis: Integration of complex signaling pathways for the recruitment of gene-specific transcription factors. *Arthritis Res.* 2002;4(3):157–64.
92. Bian F, Wang C, Tukler-Henriksson J, Pflugfelder SC, Camodeca C, Nuti E, et al. MMP-8 Is Critical for Dexamethasone Therapy in Alkali-Burned Corneas Under Dry Eye Conditions. *J Cell Physiol.* 2016 Nov 1;231(11):2506–16.
93. Sondergaard BC, Schultz N, Madsen SH, Bay-Jensen AC, Kassem M, Karsdal MA. MAPKs are essential upstream signaling pathways in proteolytic cartilage degradation - divergence in pathways leading to aggrecanase and MMP-mediated articular cartilage degradation. Vol. 18, *Osteoarthritis and Cartilage.* 2010. p. 279–88.
94. Prihadi JC, Sugandi S, Siregar NC, Soejono G, Harahap A. Imbalance in extracellular matrix degradation in urethral stricture. *Res Rep Urol.* 2018;10:227–32.
95. Wilkinson HN, Hardman MJ. Wound healing: cellular mechanisms and pathological outcomes: Cellular Mechanisms of Wound Repair. *Open Biol.* 2020;10(9).
96. Rodrigues M, Kosaric N, Bonham CA, Gurtner GC. Wound healing: A cellular perspective. *Physiol Rev.* 2019;99(1):665–706.
97. Sorg H, Tilkorn DJ, Hager S, Hauser J, Mirastschijski U. Skin Wound Healing: An Update on the Current Knowledge and Concepts. *European Surgical Research.* 2017;58(1–2):81–94.
98. Tottoli EM, Dorati R, Genta I, Chiesa E, Pisani S, Conti B. Skin wound healing process and new emerging technologies for skin wound care and regeneration. *Pharmaceutics.* 2020;12(8):1–30.
99. Miron RJ, Chai J, Fujioka-Kobayashi M, Sculean A, Zhang Y. Evaluation of 24 protocols for the production of platelet-rich fibrin. *BMC Oral Health.* 2020;20(1):1–13.
100. Reksodiputro MH, Harahap AR, Setiawan L, Yosia M. A Modified Preparation Method of Ideal Platelet-Rich Fibrin Matrix From Whole Blood. *Front Med (Lausanne).* 2021;8(August):1–9.
101. Castro AB, Andrade C, Li X, Pinto N, Teughels W, Quirynen M. Impact of g force and timing on the characteristics of platelet-rich fibrin matrices. *Sci Rep.* 2021 Mar 16;11(1):6038.
102. Dashore S, Chouhan K, Nanda S, Sharma A. Platelet-rich fibrin, preparation and use in dermatology. *Indian Dermatol Online J.* 2021;12(7):55.

103. Isobe K, Suzuki M, Watanabe T, Kitamura Y, Suzuki T, Kawabata H, et al. Platelet-rich fibrin prepared from stored whole-blood samples. *Int J Implant Dent.* 2017;3(1):0–6.
104. Saluja H, Dehane V, Mahindra U. Platelet-Rich fibrin: A second generation platelet concentrate and a new friend of oral and maxillofacial surgeons. *Ann Maxillofac Surg.* 2011;1(1):53.
105. Akyildiz S, Soluk-tekkesin ÂM, Keskin-yalcin B, Unsal ÂG, Yildiz O, Ozcan I, et al. Acceleration of Fracture Healing in Experimental Model: Platelet-Rich Fibrin or Hyaluronic Acid? *Journal of Craniofacial Surgery.* 2018 Oct;29(7):1794–8.
106. Li S, Yang H, Duan Q, Bao H, Li A, Li W, et al. A comparative study of the effects of platelet-rich fibrin, concentrated growth factor and platelet-poor plasma on the healing of tooth extraction sockets in rabbits. *BMC Oral Health.* 2022 Mar 23;22(1):87.
107. Ondur E, Balcioglu NB, Tekkesin MS, Guzel O, Ersanli S, Bolukbasi Balcioglu N, et al. Effects of Platelet-Rich Fibrin on Hard Tissue Healing: A Histomorphometric Crossover Trial in Sheep. *Materials.* 2020 Apr 4;13(7):1695.
108. Böyükbaş N, Yeniyol S, Tekkesin MS, Altunatmaz K, Böyükbaş N, Yeniyol S, et al. The Use of Platelet-Rich Fibrin in Combination With Biphasic Calcium Phosphate in the Treatment of Bone Defects: A Histologic and Histomorphometric Study. *Current Therapeutic Research.* 2013 Dec;75:15–21.
109. Shafei F, Khoshzaban A, Taleghani F, Tehranchi M, Tayeed MH. The Effect of Low-Level Laser Therapy in Combination with Leukocyte- and Platelet-Rich Fibrin on Bone Regeneration in Rabbits' Calvarial Defects: Histologic and Histomorphometric Studies. *Cell J.* 2022;24(6):346–52.
110. Salih SI, Al-falahi NH, Saliem AH, Abedsalih AN. Effectiveness of platelet-rich fibrin matrix treated with silver nanoparticles in fracture healing in rabbit model. *Vet World.* 2018 Jul;11(7):944–52.
111. Tayşı M, Atalay B, Çankaya B, Yıldırım S. Effects of single- and double-layered resorbable membranes and platelet-rich fibrin on bone healing. *Clin Oral Investig.* 2018 May 27;22(4):1689–95.
112. Demirel E, Yıldız K, Çadirci K, Aygün H, Şenocak E, Gündoğdu B. Effect of platelet-rich fibrin on epidural fibrosis and comparison to ADCON® Gel and hyaluronic acid. *Acta Orthop Traumatol Turc.* 2018 Nov;52(6):469–74.
113. Wong CC, Huang YM, Chen C hwa, Lin F huei, Yeh YY, Bai MY. Cytokine and Growth Factor Delivery from Implanted Platelet-Rich Fibrin Enhances Rabbit Achilles Tendon Healing. *Int J Mol Sci.* 2020 May 2;21(9):3221.
114. Dietrich F, L Duré G, P Klein C, F Bampi V, V Padoin A, D Silva V, et al. Platelet-Rich Fibrin Promotes an Accelerated Healing of Achilles Tendon When Compared to Platelet-Rich Plasma in Rat. *World J Plast Surg.* 2015;4(2):101–9.
115. Chuang MHH, Ho LHH, Kuo TFF, Sheu SYY, Liu YHH, Lin PCC, et al. Regenerative Potential of Platelet-Rich Fibrin Releasate Combined with

- Adipose Tissue-Derived Stem Cells in a Rat Sciatic Nerve Injury Model. *Cell Transplant.* 2020 Jan 1;29:096368972091943.
116. Kornsuthisopon C, Pirarat N, Osathanon T, Kalpravidh C. Autologous platelet-rich fibrin stimulates canine periodontal regeneration. *Sci Rep.* 2020 Feb 5;10(1):1850.
 117. Ensari N, Gür ÖE, Öztürk MT, Süren D, Selçuk ÖT, Osma Ü. The effect of platelet-rich fibrin membrane on the repair of perforated tympanic membrane: an experimental study. *Acta Otolaryngol.* 2017 Jul 3;137(7):695–9.
 118. Sun CK, Zhen YY, Leu S, Tsai TH, Chang LT, Sheu JJ, et al. Direct implantation versus platelet-rich fibrin-embedded adipose-derived mesenchymal stem cells in treating rat acute myocardial infarction. *Int J Cardiol.* 2014 May;173(3):410–23.
 119. Chen Y, Niu Z, Xue Y, Yuan F, Fu Y, Bai N. Improvement in the repair of defects in maxillofacial soft tissue in irradiated minipigs by a mixture of adipose-derived stem cells and platelet-rich fibrin. *British Journal of Oral and Maxillofacial Surgery.* 2014 Oct;52(8):740–5.
 120. Wang J, Le K, Guo X, Yan F, Guo Y, Zhang T, et al. Platelet-rich fibrin prevents postoperative intestinal adhesion. *J Biomed Mater Res A.* 2020 May;108(5):1077–85.
 121. Güler İ, Billur D, Aydin S, Kocatürk S. Efficacy of platelet-rich fibrin matrix on viability of diced cartilage grafts in a rabbit model. *Laryngoscope.* 2015 Mar;125(3):E104–11.
 122. Nica O, Popa DG, Grecu AF, Ciucă EM, Ciurea ME. Effects of Platelet Rich Fibrin on Full Thickness Skin Grafts in the Rat Model-Planimetry Results. *Curr Health Sci J.* 2019;45(3):278–84.
 123. Duan X, Lin Z, Lin X, Wang Z, Wu Y, Ji M, et al. Study of platelet-rich fibrin combined with rat periodontal ligament stem cells in periodontal tissue regeneration. *J Cell Mol Med.* 2018;22(2):1047–55.
 124. Gupta S, Gupta P, Raman N, Singh A, Shah A, Ramola V. Comparison between Different Combinations of Alendronate, Platelet-rich Fibrin, Hydroxyapatite in Bone Regeneration in Endodontic Surgeries Using Cone-beam Computed Tomography. *J Contemp Dent Pract.* 2022 Jun 24;23(3):337–42.
 125. Revathy Ns, Kannan R, Karthik R, Kumar MS, Munshi MI, Vijay R. Comparative study on alveolar bone healing in postextraction socket versus healing aided with autologous platelet-rich fibrin following surgical removal of bilateral mandibular impacted third molar tooth: A radiographic evaluation. *Natl J Maxillofac Surg.* 2018;9(2):140.
 126. Eid A, Mancino D, Rekab MS, Haikel Y, Kharouf N. Effectiveness of Three Agents in Pulpotomy Treatment of Permanent Molars with Incomplete Root Development: A Randomized Controlled Trial. *Healthcare.* 2022 Feb 25;10(3):431.
 127. Sari H, Karaketir S, Kumral TL, Akgun MF, Gurpinar B, Hancı D, et al. The effect of platelet-rich fibrin (PRF) on wound healing, adhesion, and

- hemostasis after endoscopic sinus surgery in patients with nasal polyposis. *Am J Otolaryngol.* 2021 Sep;42(5):103010.
128. Matz EL, Pearlman AM, Terlecki RP. Safety and feasibility of platelet rich fibrin matrix injections for treatment of common urologic conditions. *Investig Clin Urol.* 2018;59(1):61.
 129. Reksodiputro MH, Hutaarak SM, Koento T, Fardizza F, Hakim RYR, Audindra S, et al. Randomised clinical trial: Effect of administering platelet-rich fibrin to autologous fat tissue in injection laryngoplasty for vocal cord paralysis. *Annals of Medicine and Surgery.* 2021 Aug;68(71):102564.
 130. Yuvasri G, Rai R. Comparison of efficacy of autologous platelet-rich fibrin versus Unna's paste dressing in chronic venous leg ulcers: A comparative study. *Indian Dermatol Online J.* 2020;11(1):58.
 131. Singampalli Z, Rajan YRD, Hemanth Rathod R, RajLaxmi PLS. The Efficacy of Platelet-Rich Fibrin in the Management of Chronic Nonhealing Ulcers of the Lower Limb. *Cureus.* 2022 Jul 13;14(7):10–6.
 132. Xue X, Bian Y, Yang M, Wei W, Meng L, Zhang Q, et al. Evaluation of injectable platelet-rich fibrin produced by a simple twice-centrifugation method combined with vacuum sealing drainage technology in the treatment of chronic refractory wounds. *Front Bioeng Biotechnol.* 2022 Oct 28;10(October):1–10.
 133. Kartika RW, Alwi I, Suyatna FD, Yunir E, Waspadji S, Immanuel S, et al. The role of VEGF, PDGF and IL-6 on diabetic foot ulcer after Platelet Rich Fibrin + hyaluronic therapy. *Heliyon.* 2021 Sep;7(9):e07934.
 134. Blatt S, Krüger M, Kämmerer PW, Thiem DGEE, Mattheis P, Eisenbeiß AKK, et al. Non-Interventional Prospective Observational Study of Platelet Rich Fibrin as a Therapy Adjunctive in Patients with Medication-Related Osteonecrosis of the Jaw. *J Clin Med.* 2022 Jan 28;11(3):682.
 135. Waldner M, Ismail T, Lunger A, Klein HJ, Schweizer R, Alan O, et al. Evolution of a concept with enzymatic debridement and autologous in situ cell and platelet-rich fibrin therapy (BrokerF). *Scars Burn Heal.* 2022 Jan 6;8:205951312110523.
 136. Soares CS, Dias IR, Pires MA, Carvalho PP. Canine-Origin Platelet-Rich Fibrin as an Effective Biomaterial for Wound Healing in Domestic Cats: A Preliminary Study. *Vet Sci.* 2021 Sep 30;8(10):213.
 137. Bucur M, Constantin C, Neagu M, Zurac S, Dinca O, Vladan C, et al. Alveolar blood clots and platelet-rich fibrin induce in vitro fibroblast proliferation and migration. *Exp Ther Med.* 2018 Dec 6:982–9.
 138. Wang Z, Mudalal M, Sun Y, Liu Y, Wang J, Wang Y, et al. The Effects of Leukocyte-Platelet Rich Fibrin (L-PRF) on Suppression of the Expressions of the Pro-Inflammatory Cytokines, and Proliferation of Schwann Cell, and Neurotrophic Factors. *Sci Rep.* 2020 Feb 12;10(1):2421.
 139. Nagaraja S, Mathew S, Rajaram R, Pushpalatha C, Abraham A, Chandanala S. Evaluation of histological and pH changes in platelet-rich fibrin and platelet-rich fibrin matrix: A In vitro study. *Contemp Clin Dent.* 2019;10(4):652.

140. Talebi Ardakani MR, Meimandi M, Shaker R, Golmohammadi S. The Effect of Platelet-Rich Fibrin (PRF), Plasma Rich in Growth Factors (PRGF), and Enamel Matrix Proteins (Emdogain) on Migration of Human Gingival Fibroblasts. *J Dent (Shiraz)*. 2019;20(4):232–9.
141. Wang Z, Han L, Sun T, Wang W, Li X, Wu B. Preparation and effect of lyophilized platelet-rich fibrin on the osteogenic potential of bone marrow mesenchymal stem cells in vitro and in vivo. *Heliyon*. 2019 Oct;5(10):e02739.
142. Xu FTT, Liang ZJJ, Li HMM, Peng QLL, Huang MHH, Li DQDHQH, et al. Ginsenoside Rg1 and platelet-rich fibrin enhance human breast adipose-derived stem cells function for soft tissue regeneration. *Oncotarget*. 2016 Jun 7;7(23):35390–403.
143. Bayer A, Wijaya B, Möbus L, Rademacher F, Rodewald M, Tohidnezhad M, et al. Platelet-Released Growth Factors and Platelet-Rich Fibrin Induce Expression of Factors Involved in Extracellular Matrix Organization in Human Keratinocytes. *Int J Mol Sci*. 2020 Jun 20;21(12):4404.
144. Jasineviciute I, Grigas J, Ziukaite G, Pautienius A, Razukevicius D, Zymantiene J, et al. Peripheral mononuclear cells composition in platelet-rich fibrin in canines with chronic conditions. *Sci Rep*. 2022 Oct 19;12(1):17426.
145. Göral A, Aslan C, Bolat Küçükzeybek B, Işlk D, Hoşnute M, Durgun M, et al. Platelet-Rich Fibrin Improves the Viability of Diced Cartilage Grafts in a Rabbit Model. *Aesthet Surg J*. 2016 Apr 9;36(4):NP153–62.
146. Yu P, Zhai Z, Lu H, Jin X, Yang X, Qi Z. Platelet-Rich Fibrin Improves Fat Graft Survival Possibly by Promoting Angiogenesis and Adipogenesis, Inhibiting Apoptosis, and Regulating Collagen Production. *Aesthet Surg J*. 2020 Aug 14;40(9):NP530–45.
147. Schoem, SR.; Rosbe KW; BS. Aerodigestive Foreign Bodies and Caustic Ingestion. In: Cummings Otolaryngology Head and Neck Surgery. 2015. p. 3184–94.
148. Shedoeva A, Leavesley D, Upton Z, Fan C. Wound Healing and the Use of Medicinal Plants. Evidence-Based Complementary and Alternative Medicine. 2019 Sep 22;2019(Figure 1):1–30.
149. Hammond JW, Hinton RY, Curl LA, Muriel JM, Lovering RM. Use of autologous platelet-rich plasma to treat muscle strain injuries. *American Journal of Sports Medicine*. 2009;37(6):1135–42.
150. Reksodiputro M, Harahap A. Mengenal Lebih Dalam Produk Konsentrat Trombosit. 1st ed. Reksodiputro L, Mufida T, editors. Jakarta: UI Publishing; 2021.
151. Kobayashi M, Kawase T, Horimizu M, Okuda K, Wolff LF, Yoshie H. A proposed protocol for the standardized preparation of PRF membranes for clinical use. *Biologicals*. 2012;40(5):323–9.
152. Knight BE, Kozlowski N, Havelin J, King T, Crocker SJ, Young EE, et al. TIMP-1 Attenuates the Development of Inflammatory Pain Through MMP-Dependent and Receptor-Mediated Cell Signaling Mechanisms. *Front Mol Neurosci*. 2019 Sep 20;12(September):1–16.

153. Akay MA, Akduman M, Tataroğlu AÇ, Eraldemir C, Kum T, Vural Ç, et al. Evaluation of the efficacy of Hypericum perforatum (St. John's wort) oil in the prevention of stricture due to esophageal corrosive burns. *Esophagus*. 2019;16(4):352–61.
154. Berthet B, Di Costanzo J, Arnaud C, Choux R, Assadourian R. Influence of epidermal growth factor and interferon γ on healing of oesophageal corrosive burns in the rat. *Journal of British Surgery*. 1994 Mar 1;81(3):395–8.
155. Sengupta P. The Laboratory Rat: Relating Its Age With Human's. *Int J Prev Med*. 2013 Jun;4(6):624–30.
156. Chornenka NM, Raetska YB, Dranitsina AS, Kalmukova OO, Beregova T V., Dzerzhinsky ME, et al. Molecular Genetic and Cytological Features of Healing in Esophageal Alkaline Burns and When Melanin is Administered. *Cytol Genet*. 2020;54(4):333–40.
157. Donovan J, Brown P. Blood Collection. *Curr Protoc Immunol*. 2006 Jun;73(1):1–9.
158. Kargarpour Z, Panahipour L, Miron RJ, Gruber R. Fibrinogen Concentrations in Liquid PRF Using Various Centrifugation Protocols. *Molecules*. 2022 Mar 22;27(7):2043.
159. Oztan MO, Arslan FD, Oztan S, Diniz G, Koyluoglu G. Effects of topical application of platelet-rich plasma on esophageal stricture and oxidative stress after caustic burn in rats: Is autologous treatment possible? *J Pediatr Surg*. 2019;54(7):1397–404.
160. Kalkan Y, Tumkaya L, Akdogan RA, Yucel AF, Tomak Y, Sehitoglu İ, et al. A novel model approach for esophageal burns in rats. *Toxicol Ind Health*. 2015 Jul 1;31(7):595–601.
161. Rigalli A, Loreto VE Di. *Experimental Surgical Models in the Laboratory Rat*. Boca Raton, USA: CRC Press; 2009.
162. Flecknell P. Rodent analgesia: Assessment and therapeutics. Vol. 232, *Veterinary Journal*. Elsevier Ltd.; 2018. p. 70–7.
163. Hawk CT, Leary SL. *Formulary For Laboratory Animals*. 2nd ed. Ames: Iowa State University Press; 2005. 167 p.
164. Cevik M, Demir T, Karadag CA, Ketani MA, Celik H, Kaplan DS, et al. Preliminary study of efficacy of hyaluronic acid on caustic esophageal burns in an experimental rat model. *J Pediatr Surg*. 2013;48(4):716–23.
165. Crowe AR, Yue W. Semi-quantitative Determination of Protein Expression Using Immunohistochemistry Staining and Analysis. *Bio Protoc*. 2019 Dec 20;9(24).
166. Gehanno P, Guedon C. Inhibition of Experimental Esophageal Lye Strictures by Penicillamine. *Archives of Otolaryngology - Head and Neck Surgery*. 1981 Mar 1;107(3):145–7.
167. Liu AJ, Richardson MA. Effects of N-Acetylcysteine on Experimentally Induced Esophageal Lye Injury. *Annals of Otology, Rhinology & Laryngology*. 1985 Sep 1;94(5):477–82.
168. Okata Y, Hisamatsu C, Hasegawa T, Nishijima E, Okita Y. Development of a model of benign esophageal stricture in rats: the optimal concentration of

- sodium hydroxide for stricture formation. *Pediatr Surg Int.* 2011 Jan 24;27(1):73–80.
169. Wu R, Fu M, Tao HM, Dong T, Fan WT, Zhao LL, et al. Benign esophageal stricture model construction and mechanism exploration. *Sci Rep.* 2023 Jul 20;13(1):11769.
 170. Bakan V, Cirali H, Kartal S. A corrosive oesophageal burn model in rats: Double-lumen central venous catheter usage. *Afr J Paediatr Surg.* 2015;12(4):247–50.
 171. Aciksari K, Yanar HT, Hepgul G, Ertekin C, Gunay K, Ozcelik DN, et al. 183 Effect of Beta-aminopropionitrile on the Prevention of Fibrosis in Alkali Burns of the Esophagus: An Experimental Study. *Ann Emerg Med.* 2012;60(4):S66.
 172. Numanoğlu KV, Tatlı D, Bektaş S, Er E. Efficacy of keratinocyte growth factor (palifermin) for the treatment of caustic esophageal burns. *Exp Ther Med.* 2014;8(4):1087–91.
 173. Senturk E, Pabucu E, Sen S, Unsal C. Comparison of mitomycin-c and heparin affects in experimental corrosive esophagitis on rats. *Int J Pediatr Otorhinolaryngol.* 2011 Jun;75(6):785–9.
 174. Kalkan Y, Tumkaya L, Akdogan RA, Yucel AF, Tomak Y, Sehitoglu İ, et al. A novel model approach for esophageal burns in rats: A comparison of three methods. *Toxicol Ind Health.* 2015;31(7):595–601.
 175. Sharp P, Villano J. The laboratory rat, second edition. Vol. i, The Laboratory Rat, Second Edition. 2012. 1–359 p.
 176. Treuting PM, Dintzis SM. Comparative Anatomy and Histology: A Mouse and Human Atlas. In: Comparative Anatomy and Histology: A Mouse and Human Atlas. Academic Press; 2012. p. 46–51.
 177. Millar AJW, Cox SG. Caustic injury of the oesophagus. *Pediatr Surg Int.* 2015;31(2):111–21.
 178. Rieder F, Biancani P, Harnett K, Yerian L, Falk GW. Inflammatory mediators in gastroesophageal reflux disease: impact on esophageal motility, fibrosis, and carcinogenesis. *Am J Physiol Gastrointest Liver Physiol.* 2010 May;298(5):G571–81.
 179. Calabrese EJ, Baldwin LA. Hormesis: The Dose-Response Revolution. Vol. 43, Annual Review of Pharmacology and Toxicology. 2003. p. 175–97.
 180. Kendig EL, Le HH, Belcher SM. Defining hormesis: Evaluation of a complex concentration response phenomenon. Vol. 29, International Journal of Toxicology. 2010. p. 235–46.
 181. Calabrese EJ. Hormetic dose-response relationships in immunology: Occurrence, quantitative features of the dose response, mechanistic foundations, and clinical implications. Vol. 35, Critical Reviews in Toxicology. 2005. p. 89–295.
 182. Calabrese EJ. Getting the dose-response wrong: Why hormesis became marginalized and the threshold model accepted. Vol. 83, Archives of Toxicology. 2009. p. 227–47.

183. Calabrese EJ, Dhawan G, Kapoor R, Agathokleous E, Calabrese V. Hormesis: wound healing and fibroblasts. Vol. 184, Pharmacological Research. Academic Press; 2022.
184. Kobayashi E, Flückiger L, Fujioka-Kobayashi M, Sawada K, Sculean A, Schaller B, et al. Comparative release of growth factors from PRP, PRF, and advanced-PRF. *Clin Oral Investig*. 2016 Dec 1;20(9):2353–60.
185. Baca-Gonzalez L, Serrano Zamora R, Rancan L, González Fernández-Tresguerres F, Fernández-Tresguerres I, López-Pintor RM, et al. Plasma rich in growth factors (PRGF) and leukocyte-platelet rich fibrin (L-PRF): comparative release of growth factors and biological effect on osteoblasts. *Int J Implant Dent*. 2022 Oct 3;8(1).
186. Masuki H, Okudera T, Watanabe T, Suzuki M, Nishiyama K, Okudera H, et al. Growth factor and pro-inflammatory cytokine contents in platelet-rich plasma (PRP), plasma rich in growth factors (PRGF), advanced platelet-rich fibrin (A-PRF), and concentrated growth factors (CGF). *Int J Implant Dent*. 2016 Dec;2(1).
187. Calabrese EJ, Kapoor R, Dhawan G, Calabrese V. Hormesis mediates platelet-rich plasma and wound healing. Vol. 31, Wound Repair and Regeneration. John Wiley and Sons Inc; 2023. p. 56–68.
188. Hirshoren N, Eliashar R. Wound-healing modulation in upper airway stenosis-myths and facts. Wiley InterScience (www.interscience.wiley.com). 2008;111–26.
189. Calabrese EJ, Giordano JJ, Kozumbo WJ, Leak RK, Bhatia TN. Hormesis mediates dose-sensitive shifts in macrophage activation patterns. Vol. 137, Pharmacological Research. Academic Press; 2018. p. 236–49.
190. Zhang Q, Pi J, Woods CG, Jarabek AM, Clewell HJ, Andersen ME. Hormesis and adaptive cellular control systems. *Dose-Response*. 2008;6(2):196–208.
191. Bauer M, Ermolaeva M, Singer M, Wetzker R, Soares MP. Hormesis as an adaptive response to infection. Vol. 30, Trends in Molecular Medicine. Elsevier Ltd; 2024. p. 633–41.
192. Sah RL, Trippel SB, Grodzinsky AJ. Differential effects of serum, insulin-like growth factor-I, and fibroblast growth factor-2 on the maintenance of cartilage physical properties during long-term culture. *Journal of Orthopaedic Research [Internet]*. 1996 Jan 18;14(1):44–52. Available from: <https://onlinelibrary.wiley.com/doi/10.1002/jor.1100140109>
193. Elson EL, Qian H, Fee JA, Wakatsuki T. A model for positive feedback control of the transformation of fibroblasts to myofibroblasts. Vol. 144, Progress in Biophysics and Molecular Biology. Elsevier Ltd; 2019. p. 30–40.
194. Kanodia J, Chai D, Vollmer J, Kim J, Raue A, Finn G, et al. Deciphering the mechanism behind Fibroblast Growth Factor (FGF) induced biphasic signal-response profiles. *Cell Communication and Signaling*. 2014 May 15;12(1).
195. Lee KS. Platelet-rich plasma induces increased expression of G1 cell cycle regulators, type I collagen, and matrix metalloproteinase-1 in human skin fibroblasts. *Int J Mol Med*. 2011 Sep 30;

196. Talebi Ardakani MR, Meimandi M, Shaker R, Golmohammadi S. The Effect of Platelet-Rich Fibrin (PRF), Plasma Rich in Growth Factors (PRGF), and Enamel Matrix Proteins (Emdogain) on Migration of Human Gingival Fibroblasts. *J Dent (Shiraz)*. 2019;20(4):232–9.
197. Shi HX, Lin C, Lin BB, Wang ZG, Zhang HY, Wu FZ, et al. The Anti-Scar Effects of Basic Fibroblast Growth Factor on the Wound Repair In Vitro and In Vivo. Xu A, editor. *PLoS One*. 2013 Apr 2;8(4):e59966.