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	501	THE ROLE OF TEMPERATURE AND pH IN THE SYNTHESIS OF SILVER NANOPARTICLES USING ARECA CATECHU L. SEED EXTRACT AS BIOREDUCTOR	2023-01-27 06:03:52	New Submission	



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AUTHOR	Mardiyanto Mardiyanto
NAME	
MANUSCRIPT	THE ROLE OF TEMPERATURE AND pH IN THE SYNTHESIS
TITLE	OF SILVER NANOPARTICLES USING ARECA CATECHU L.
	SEED EXTRACT AS BIOREDUCTOR

### **Reviewer** A

- 1. At the beginning of the introduction just focus on silver NPs.
- 2. line 27 there is no relationship between the first sentence and the next.
- 3. Line 36 what are the details of the active compound?
- 4. Line 73 write what is the standard compound used.
- 5. Line 84 add the ratio number.
- 6. Line 133 why does the reduction reaction occur.
- 7. Line 156 refers to which Tab?
- 8. Line 249-250 subtitle is included in previous sentence ?
- 9. Line 280 refers to which Fig?

### **Reviewer B**

- 1. The first sentence in introduction is not necessary
- 2. What kind of the active compound in sample ?
- 3. Please write the standard for quantification
- 4. Please write in short sentence regarding the reason does the reduction reaction AgNO<sub>3</sub> to Ag
- 5. Some number of Table is not mentioned

### Sebelum Riview

1	THE ROLE OF TEMPERATURE AND pH IN THE SYNTHESIS OF SILVER
2	NANOPARTICLES USING ARECA CATECHU L. SEED EXTRACT AS BIOREDUCTOR
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10	Abstract
11	Silver nanoparticles are prepared using natural bioreductors such as Areca catechu ethanol extract,
12	which is rich in phenolic content. This study aims to optimize the temperature and pH conditions
13	for synthesizing silver nanoparticles using the response surface methodology (RSM)-central
14	composite design (CCD) method. The phenolic content of the Areca catechu ethanol extract was
15	111.14 $\pm$ 3.41 mg CE/g extract. Temperature and pH conditions significantly affect maximum
16	wavelength and absorbance values. The optimum condition was obtained at a temperature of $30^{\circ}C$
17	and a pH of 10.5. Silver nanoparticles at optimum condition had a wavelength of 423 nm,
18	absorbance was 1.148 particle size was 161.7 $\pm$ 46.1 nm, PDI was 0.286 $\pm$ 0.035, and zeta potential
19	was -16.1 $\pm$ 3.7 mV. The stability of silver nanoparticles at the optimum conditions produced is
20	relatively stable, characterized by no significant changes in organoleptic, pH, and wavelength, but
21	the absorbance value has increased. The resulting silver nanoparticles have good characteristics
22	and good stability.
23	Keywords: Areca catechu L. seed, pH, response surface methodology, silver nanoparticles,

24 temperature

### 25 Introduction

The use of areca seeds is generally to overcome the decay of body tissue and to preserve animal skin. Cowhide is usually preserved with crushed areca seeds. The results of more detailed research show that areca seeds can inactivate the growth of rotting germs. Nanotechnology in the pharmaceutical field is currently developing very rapidly. Silver nanoparticles are among the most researched and set in the medical world because they have been shown to have anti- bacterial, anti-inflammatory, antiangiogenesis, anti- fungal, antiviral, and antiplatelet activities [1-4]. The production of silver nanoparticles has shifted towards green synthesis methods that are more environmentally friendly. The green synthesis method requires the help of secondary metabolites in plants as a source of bioreductors [5]. Secondary metabolites that can act as bioreductors usually act as antioxidants, such as flavonoids, phenolics, and alkaloids [6]. Flavonoid, phenolic and alkaloid compounds contain many groups whose electrons require a partner so that they are needed to react organic-chemically way [7, 8].

The formation of silver nanoparticles through the green synthesis method can be influenced 38 by several factors, such as temperature and pH [9]. Temperature is another critical factor affecting 39 the formation of nanoparticles in plant extracts [10]. In general, an increase in temperature will 40 increase nanoparticle synthesis's reaction rate and efficiency [11]. This statement is proven by research 41 42 conducted by Jiang et al [12], where a temperature of 17 to 55°C will accelerate the reaction and cause the particle size to increase from 90 nm to 180 nm. Besides temperature, changes in pH will also affect 43 the process of forming silver nanoparticles. Changes in pH will result in changes in the natural 44 45 phytochemical charge contained in the extract. This effect will affect its ability to bind and reduce metal cations while synthesizing silver nanoparticles. According to research by Marciniak et al. [13], the 46 47 reaction for forming silver nanoparticles runs at a pH of 6.0 to 11.0 for citric acid and 7.0 to 11.0 for malic acid. Increasing the pH can also affect the particle size of silver nanoparticles, where the higher 48 the pH value, the smaller the particle size [14]. 49

Based on the description above, researchers are interested in conducting research in the form of 50 optimizing the effect of temperature and pH on lmax surface plasmon resonance (SPR) and absorbance 51 52 in the manufacture of silver nanoparticles using ethanol extract of young areca nut seeds as a bioreductor agent. Optimization was carried out using the response surface methodology (RSM) -53 central composite design (CCD) method for temperature and pH. The temperature and pH used in this 54 study were  $30 - 70^{\circ}$ C respectively and the pH range was 7.5 - 10.5 referring to the research by 55 Prodjosantoso et al. [15] and Seifipour et al. [16] with slight modifications. The formula of silver 56 57 nanoparticles with optimum temperature and pH will be further characterized in particle size, polydispersity index (PDI), zeta potential and thermodynamic stability tests using the cycling test 58 method. 59

- 61 Materials and Methods
- 62 Materials

The materials used in this study were Areca catechu L. seed s, AgNO<sub>3</sub> (Emsure, Indonesia), 70% 63 ethanol (Bratachem, Indonesia), absolute ethanol (Emsure, Indonesia), NaOH 0.1 M (Bratachem, 64 Indonesia), catechins (Sigma-Aldrich, Singapore), and aqua deionized (Bratachem, Indonesia).

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#### 67 Preparation of Ethanolic Extract of Areca catechu L. Seed

Areca catechu L. seeds were washed, dried and crushed to obtain Simplicia powder. The simplicia 68 powder was macerated using 70% ethanol solvent and stored in a dark place for 48 hours. The filtrate 69 was then evaporated using a rotary evaporator to obtain a thick extract. 70

71 Quantification of Total Phenolic Content in Extract The measurement of total phenolic content in the extract was carried out based on the research of Indarti et al. [17] and Sudirman et al. [18] with 72 modifications. The extract is put at 10 mg into a 100 mL measuring flask, then the volume is made up 73 to obtain a 100 mg/mL concentration. The absorbance of the sample solution was measured using a 74 UV-Vis spectrophotometer at 281 nm. The formula can calculation the total phenolic concentration 75 76 (TPC) in extract in Equation 1.

TPC = volume (mL) x concentration (mg/mL) x dilute factor

q extract

(1)

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- 78
- 79
- 80

#### **Formula of Silver Nanoparticles** 81

82 The formula for making silver nanoparticles using the ethanol extract of Areca catechu L. refers to research by Apriani et al. [19]. The concentration of silver nitrate (AgNO<sub>3</sub>) used was 3 mM 83 and 3 mL of extract at a concentration of 10% w/v. 84

85

#### Temperature and pH Optimization 86

Optimization of temperature and pH in the manufacture of silver nanoparticles was carried out using 87 the response surface methodology (RSM) - central composite design (CCD) method referring to the 88 research of Prodjosantoso et al. [15] and Seifipour et al. [16] with slight modifications. The levels 89 used consist of low, middle, and high levels. The design of the levels for each factor, namely 90 91 temperature and pH can be seen in Table I. Based on the conditions in Table I, nine formulas were produced for manufacturing silver nanoparticles which can be seen in Table II. 92

### 94 Determination of Surface Plasmon Resonance

Surface plasmon resonance was determined by observing the wavelength and absorbance of the 95 sample solution in the range of 370 - 600 nm using a UV-Vis Spectrophotometer. The blank 96 used in this procedure is distilled water. Determination of Optimum Conditions for Silver 97 Nanoparticles Optimum conditions (temperature and pH) were determined to manufacture silver 98 nanoparticles by analyzing surface plasmon resonance data using the Design Expert 13<sup>®</sup> 99 program. The program will examine the effect of temperature and pH factors on surface plasmon 100 resonance's wavelength response and absorbance. The program suggests optimum conditions when 101 102 these conditions have a desirability value close to 1.

103

### 104 Characterization of Silver Nanoparticles at Optimum Condition

105 Silver nanoparticles under optimum conditions were subjected to further characterization, such 106 as the determination of surface plasmon resonance, particle size, polydispersity index, and zeta 107 potential. Particle size, polydispersity index, and zeta potential were measured using a Particle 108 Size Analyzer. The sample solution is diluted using aqua deionized. Particle size and 109 polydispersity index were measured at a scattering angle of 90°, while measured the zeta potential 110 at a scattering angle of 173° [22].

111

### 112 Stability Test of Silver Nanoparticles at Optimum Condition

A stability test was carried out using the cycling test method. The cycling test was carried out at temperatures 4 °C with storage at each temperature for 24 hour. The procedure was repeated for six cycles [23.24]. Organoleptic observation, pH, and surface plasmon resonance were observed in cycles 0 and 6.

116

### 117 Data Analysis

- 118 Data analysis for optimization was performed using program Design-Expert 13<sup>®</sup>.
- 119
- 120

### 121 Results and Discussion

- 122 Areca catechu L. Seeds Extract
- 123 The *Areca catechu* L. extract obtained in this study had a distinctive odour, brown colour and
- thick. The total phenolic content obtained was  $111.4 \pm 0.411$  mg CE/g extract. The results were not
- 125 much different from those of Zhang *et al.* [25].

126 127

### 128 Silver Nanoparticles

Areca catechu L. seed extract is a bioreductor in forming silver nanoparticles. The silver nanoparticle preparations obtained in this study were blackish- brown in colour. Areca catechu L. seed extract can act as a bioreductor due to the presence of phenolic compounds. Phenolics can actively chelate and reduce metal ions into nanoparticles. The transformation of the phenolic tautomer from the enol to the keto form by releasing reactive hydrogen atoms can reduce metal ions to form nanoparticles [11]. An overview of the mechanism for creating silver nanoparticles from catechin compounds can be seen in Figure 1.

136

The formation of silver nanoparticles can be observed from surface plasmon resonance events. 137 Surface plasmon resonance (SPR) is a free electron resonance oscillation on a metal surface layer 138 that is excited by an incident light source [26]. The oscillation is determined by the absorption of the 139 wavelength obtained [27]. Surface plasmon resonance (SPR) wavelengths are 400 - 450 nm due 140 to the interaction between light and surface electrons moving from silver nanoparticles [28]. In 141 addition, the absorbance produced at surface plasmon resonance wavelengths can describe the 142 colour intensity and the number of silver nanoparticles formed during the synthesis process. The 143 144 higher the absorbance, the more silver nanoparticles are formed. The surface plasmon resonance results obtained in this study were in the wavelength range of 424 - 431 nm and absorbance of 145 0.431 - 1.265 (Table III). 146

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148 The maximum wavelength and absorbance data from the SPR obtained are closely related to 149 each formula's different temperature and temperature conditions. Based on Figure 2, F3 has 150 the highest absorbance of 1.265 while F9 has the lowest wavelength of 423 nm.

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### 152 The Role of Temperature and pH to λmax of Silver Nanoparticles

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The role of temperature and pH on the maximum wavelength of silver nanoparticles was analyzed using the Design Expert 13<sup>®</sup> program. The initial step is to analyze the model and then analyze the response. The model analysis obtained at the maximum wavelength response. Model analyzing on lmax has an R<sup>2</sup> value of more than 0.7 namely 0.9161, which shows that 91.61%

of the data on the lmax response is influenced by temperature and pH factors. In comparison, the 158 remaining 8.39% is an estimation error. This result is supported by the normal curve plot of 159 residuals (Figure 3A) which shows that the data points are close to a straight line so that the data can 160 be said to be normally distributed. Furthermore, the adjusted  $R^2$  and predicted  $R^2$  values obtained 161 in the lmax model analyzing have a difference between adjusted R<sup>2</sup> and predicted R<sup>2</sup> values illustrates 162 the similarity of the model obtained from the relationship between temperature and pH to the lmax 163 response. This result is supported by the predicted vs. actual curve in Figure 3B. The predicted 164 vs. actual curve illustrates the results of the adjusted  $R^2$  and predicted  $R^2$  values with points to the 165 right and left of the line. The closer the distance between the dots to the right and left of the line 166 indicates that the data values from the research are more precise than those predicted by the 167 system, where 86.58% of the existing data already represents the population and can explain a 168 169 good linear relationship. The adequate value obtained in the model analysis is more than 4, which 170 is 11.4290. The greater the adequate precision value, the more resistant the model will be to noise 171 (disturbance). In addition, the p-value of less than 0.05 in the model indicates that the model used significantly affects the lmax response. Based on the model analyzing, it is said that the model is 172 good to be continued in the response analyzing process. Analyzing of the lmax response was 173 174 carried out to observe the effect of each factor and the interaction between the two on the response. Response analysis was carried out by following the results of the ANOVA in the form 175 of coefficient values and p- values of the temperature (A) and pH (B) factors. The results of the 176 ANOVA analysis on the pH response can be seen in Table V. Based on the results of the ANOVA 177 analysis in Table V, temperature and pH significantly affect the lmax response where the p-value 178 obtained is less than 0.05. Factors that have a significant effect are then entered into the response 179 equation so that the response equation lmax is accepted, namely y = 426.667 + 1.833 A - 2.167 B. 180 The notation for factors A and B is (+) and (-), respectively. This notation shows that the A-181 182 temperature factor has a positive correlation, meaning that the greater the A value, the higher the resulting lmax value, In contrast, the B-pH factor has a negative correlation, meaning that the 183 greater the pH, the smaller the resulting lmax value. This result was supported the research of 184 Yeshchenko et al. [29], which showed that an increase in temperature causes a redshift (a shift 185 in wavelength to a more significant value). This condition is due to the thermal volume expansion 186 of the silver nanoparticles with increasing temperature. Thermal volume expansion is when a 187 substance experiences a change in temperature so that the substance can expand (expand) or 188

shrink (shrink) depending on the increase or decrease in temperature. Research conducted by Alqadi 189 et al. [30] also supported the effect of pH in this study. The study showed that increasing the pH 190 resulted in smaller lmax and was followed by the formation of smaller silver nanoparticle sizes. 191 According to Sathishkumar et al. [31], when the pH is acidic, the aggregation of silver nanoparticles 192 to form larger nanoparticles is believed to be preferable to the stages of creating new nanoparticles 193 (nucleation). However, at alkaline pH, it will show a large number of functional groups available for 194 silver binding, thereby facilitating a higher number of Ag<sup>+</sup> ions to bind and subsequently form a 195 large number of smaller diameter nanoparticles. 196

197

### 198 The Role of Temperature and pH to Absorbance of Silver Nanoparticle

199 The model analysis obtained at the maximum wave- length response can be seen in Table VI. The model analysis results indicate that the model is robust and significant enough to proceed to the 200 response analysis process. Model analysis on the absorbance response has an R<sup>2</sup> value of 0.9700, 201 which indicates that 97% of the data on the absorbance response is influenced by temperature and pH 202 factors. In comparison, the remaining 3% is an estimation error. This result is supported by the 203 normal plot of the residual curve (Figure 4A), where the data points are close to a straight line so 204 205 that the data can be said to be normally distributed. Furthermore, the adjusted R<sup>2</sup> and predicted  $R^2$  values obtained in the absorbance model analysis have a difference of less than 0.2 namely 206 0.0697, so there is a similarity in the model obtained from the relationship between 207 temperature and pH on the absorbance response. This result is supported by the predicted vs. 208 209 actual curve in Figure 4B, which shows the distance between the points to the right and left of the 210 near line where 95.19% of the existing data already represents the population and can explain a good linear relationship. The adeq precision value obtained in the model analysis is more than 4 211 212 namely 18.5757 so the model is robust against noise (disturbance). In addition, the p-value of less than 0.05 in the model indicates that the model used significantly affects the absorbance response. The 213 absorbance response analysis was continued, and the results obtained can be seen in Table VII. 214 Based on the results of the ANOVA analysis in Table VII, temperature and pH have a 215 significant effect on the absorbance response. The absorbance response equation obtained is y = 216 0.930 + 0.094A + 0.346B - 0.146B2. The notation on an equation for factors A and B is (+), but 217 218 there is a new factor, namely B2, which has a value of (-). Factor B2, marked (-), means that when the pH used is too high, the absorbance produced will be smaller. When viewed from the p-value, 219

factor B has the smallest p-value, so that factor B has the greatest influence on the absorbanceresponse.

The temperature parameter is directly proportional to the absorbance value. This shows that when there is an increase in temperature, the absorbance value of the silver nanoparticles produced will increase. Based on research by Prodjosantoso *et al.* [15], when the temperature increases, the formation rate of silver nanoparticles will be faster so that many silver nanoparticles are formed. The more silver nanoparticles formed, the higher the absorbance [32].

In addition, the pH parameter is also directly proportional and has the most significant effect on the 227 228 absorbance value. This shows that when there is an increase in pH, the absorbance value of the silver nanoparticles will be higher. These results are supported by research by Alqadi et al. [30], 229 which showed that an increase in pH resulted in a higher absorbance of silver nanoparticles. In 230 231 addition, Roopan et al. [33] research showed that at pH 2 no reaction to form silver nanoparticles 232 occurred, while at pH 11, the silver nanoparticles formed were highly monodispersed. Research by 233 Tagad et al. [34] also showed that at acidic pH (pH 4 and 6), there is no surface plasmon resonance absorption which is characteristic of silver nanoparticles. The formation of silver 234 nanoparticles is more suitable to occur in alkaline pH conditions so that many small silver 235 236 nanoparticles are formed, and the resulting absorbance value will be higher [35]. However, the synthesis of silver nanoparticles at extreme alkaline pH (> 12) can produce low stability and 237 cause aggregation [36]. 238

239

### 240 Optimum Condition for Silver Nanoparticles

Determination of optimum conditions is done by looking at the system's desirability value closest to 1. The desirability value close to 1 indicates that the situation meets the target desired by the researcher. Based on the analysis results, the optimum conditions for making silver nanoparticles are recommended for the system to be at a temperature of 30°C and pH of

10.5 with a desirability value of 0.932. *Characterization of Silver Nanoparticles at Optimum* 

246 Condition

Silver nanoparticles at optimum conditions were tested for characterization, such as surface plasmon resonance, particle size, polydispersity index, and zeta potential. The results of the characterization can be seen in Table VIII. Silver nanoparticles in the optimum condition have an excellent  $\lambda$ max value and absorbance for silver nanoparticles. The size of the silver 251 nanoparticles obtained was  $161.7 \pm 46.1$  nm. The particles size results obtained were smaller than previous studies conducted by Choi et al. [37] at 233.4 – 238 nm and Bhat et al. [38] at 252 553 - 610 nm. The polydispersity index value was  $0.286 \pm 0.035$  indicating that the particles 253 size obtained was uniform. The zeta potential value obtained from the optimum formula for 254 255 silver nanoparticles was  $-16.1 \pm 3.7$  mV. This results indicate that the zeta potential of the silver nanoparticle optimum formula is still relatively stable. The negative zeta potential value is due 256 to the alkaline pH (pH 10.5) during the biosynthesis of silver nanoparticles. With an increase in 257 pH (alkaline pH), the organic functional groups on the surface of silver nanoparticles reach a 258 259 higher level of deprotonation (release of H<sup>+</sup> protons) so that the amount of negative surface charge increases simultaneously [39]. The characterization results obtained have met the requirements 260 for nanoparticle preparations. 261

262

### 263 Stability of Silver Nanoparticles at Optimum Condition

The silver nanoparticles in optimum condition were tested for stability by cycling for six cycles. 264 The stability results of silver nanoparticles can be seen in Table IX. Organoleptic results showed 265 the formation of a slightly precipitate in the  $6^{th}$  cycle. The results obtained are better than the 266 previous reference research conducted by Apriani et al. [19], which showed many deposits 267 formed on colloidal silver nanoparticles in the 6th cycle. The difference in stability results 268 269 obtained can be due to differences in the conditions for synthesizing silver nanoparticles. This study used a temperature of 30 °C and pH 10.5 as the optimum condition, while the study of Apriani et 270 al. [19] used a temperature of 60 °C and pH 9. Research by Pinto et al. [40] showed that high 271 temperatures can produce unstable silver nanoparticles and can cause damage or decomposition 272 of plant secondary metabolites, which function as stabilizing agents (capping agents). Changes 273 274 in the pH value in this study were also said to be relatively stable, where the pH changed from  $10.50 \pm 0.008$  to  $10.10 \pm 0.008$ . In contrast to previous research from the same researcher, Apriani 275 et al. [19], with the conditions of nanoparticle synthesis at a temperature of 60 °C and pH 9 the 276 decrease in pH was significant, namely pH 9. This stable pH also affect changes in  $\lambda$ max value and 277 absorbance. The shift in  $\lambda$  max in the 6<sup>th</sup> cycle was not too far and was still in the range of silver 278 nanoparticles. In contrast, the absorbance in the 6<sup>th</sup> cycle increased, indicating that more and 279 more silver nanoparticles were formed. This is because phenolic stability depends on pH. 280 Phenolic compounds are in the enol form at acidic pH and are in the keto form at an alkaline 281

pH. The keto form of phenolic is helpful as a stabilizing agent (capping agent). In this study, the
reaction to form silver nanoparticles continued because the environmental pH was relatively stable
at around 10.

285

### 286 Conclusions

287 Temperature and pH conditions in manufacturing silver nanoparticles have been shown to affect the surface plasmon resonance characteristics. The temperature has positive effect on  $\lambda$ max and the 288 289 absorbance of SPR, while pH harms  $\lambda$  max and has positive impact on the absorbance of SPR. The optimum temperature and pH conditions for the synthesis of silver nano- particles using 290 ethanol extract of young areca nut seeds were found at 30 °C and pH 10.5 with a surface plasmon 291 resonance silver nanoparticle  $\lambda$ max value of 423 nm and an absorbance of 1.148. Particle size, 292 polydispersity index (PDI), and zeta potential of silver nanoparticles at optimum condition were 293  $161.7 \pm 46.1$  nm,  $0.286 \pm 0.035$  and  $-16.1 \pm 3.7$  mV respectively. The stability of the silver 294 295 nanoparticles at the optimum conditions produced was relatively stable, characterized by not too many 296 organoleptic changes, pH, which was still stable in the range of 10 small wavelength shifts, and increased absorbance values. 297

298

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- 301

### **302 Conflict of interest**

- 303 The authors declare no conflict of interest.
- 304
- 305

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## APPENDIX

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### 

### Level design for temperature and pH

Table I

Table II

### 

Fastar		Level				
Factor	Low	Medium	High			
Temperature	30	50	70			
pН	7.5	9	10.5			

### 

### 

### 

Formula and condition of silver nanoparticles

Formula	AgNO <sub>3</sub> 3 mM (mL)	Extract 10% b/v (mL)	Temp (°C)	pН
1	27	3	70	7.5
2	27	3	30	9
3	27	3	70	10.5
4	27	3	50	9
5	27	3	50	7.5
6	27	3	70	9
7	27	3	30	10.5
8	27	3	30	7.5
9	27	3	50	10.5





- 449 450 451 452 453 454 455 456 457 458 459 460 461 462

#### Table III

### Data maximum wavelength and absorbance of silver nanoparticles

Formula	Temperature (°C)	pН	Max. Wavelength (nm)	Absorbance
1	70	7.5	429	0.446
2	30	9	424	0.769
3	70	10.5	427	1.265
4	50	9	425	0.962
5	50	7.5	431	0.431
6	70	9	430	1.059
7	30	10.5	424	1.004
8	30	7.5	427	0.435
9	50	10.5	423	1.121



Table IV Model Analyzing of  $\lambda$ max Responses

Response	Parameter				
	R <sup>2</sup>	Adjusted R <sup>2</sup>	Predicted R <sup>2</sup>	Adequate precision	p-value
Max. Wavelength	0.9161	0.8658	0.6980	11.4290	0.0192





### 

480

Table V

484	Response analyzing of $\lambda$ max
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Responses	Intercept (nm)	Α	В
λmaks SPR	426.667	1.833	-2.167
p-values		0.0397*	0.0213*
A: Temperature; B:	pH; *factors that ]	have a signific	ant effect (p-v

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### 

**Table VI** Model analysis of absorbance responses 

Response		Parameter					
	R <sup>2</sup>	Adjusted R <sup>2</sup>	Predicted R <sup>2</sup>	Adequate precision	p-value		
Absorbance	0.9700	0.9519	0.8822	18.5757	0.0003		





1	THE ROLE OF TEMPERATURE AND pH IN THE SYNTHESIS OF SILVER
2	NANOPARTICLES USING ARECA CATECHU L. SEED EXTRACT AS BIOREDUCTOR
3	
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5	
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8	
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10	
11	Abstract
12	Silver nanoparticles are prepared using natural bioreductors such as Areca catechu ethanol extract,

which is rich in phenolic content. This study aims to optimize the temperature and pH conditions 13 for synthesizing silver nanoparticles using the response surface methodology (RSM)-central 14 composite design (CCD) method. The phenolic content of the Areca catechu ethanol extract was 15  $111.14 \pm 3.41$  mg CE/g extract. Temperature and pH conditions significantly affect maximum 16 17 wavelength and absorbance values. The optimum condition was obtained at a temperature of 30°C and a pH of 10.5. Silver nanoparticles at optimum condition had a wavelength of 423 nm, 18 19 absorbance was 1.148 particle size was  $161.7 \pm 46.1$  nm, PDI was  $0.286 \pm 0.035$ , and zeta potential was  $-16.1 \pm 3.7$  mV. The stability of silver nanoparticles at the optimum conditions produced is 20 relatively stable, characterized by no significant changes in organoleptic, pH, and wavelength, but 21 22 the absorbance value has increased. The resulting silver nanoparticles have good characteristics and good stability. 23 Keywords: Areca catechu L. seed, pH, response surface methodology, silver nanoparticles, 24

- 25 temperature
- 26 Introduction
- 27
- 28 The use of areca seeds is generally to overcome the decay of body tissue and to preserve animal
- 29 skin. Cowhide is usually preserved with crushed areca seeds. The results of more detailed research show
- 30 that areca seeds can inactivate the growth of rotting germs. Nanotechnology in the pharmaceutical field
- 31 is currently developing very rapidly. Silver nanoparticles are among the most researched and set in

the medical world because they have been shown to have anti- bacterial, anti-inflammatory, antiangiogenesis, anti- fungal, antiviral, and antiplatelet activities [1-4].

The production of silver nanoparticles has shifted towards green synthesis methods that are more environmentally friendly. The green synthesis method requires the help of secondary metabolites in plants as a source of bioreductors [5]. Secondary metabolites that can act as bioreductors usually act as antioxidants, such as flavonoids, phenolics, and alkaloids [6]. *Areca catechu* L. is a plant rich in alkaloid compounds such as guvacine, guvacoline, arecaidine, and arecoline and also rich in polyphenolic compounds such as catechin and quercetin [7, 8].

The formation of silver nanoparticles through the green synthesis method can be influenced 40 by several factors, such as temperature and pH [9]. Temperature is another critical factor affecting 41 the formation of nanoparticles in plant extracts [10]. In general, an increase in temperature will 42 increase nanoparticle synthesis's reaction rate and efficiency [11]. This statement is proven by research 43 conducted by Jiang et al [12], where a temperature of 17 to 55°C will accelerate the reaction and cause 44 the particle size to increase from 90 nm to 180 nm. Besides temperature, changes in pH will also affect 45 the process of forming silver nanoparticles. Changes in pH will result in changes in the natural 46 47 phytochemical charge contained in the extract. This effect will affect its ability to bind and reduce metal cations while synthesizing silver nanoparticles. According to research by Marciniak et al. [13], the 48 reaction for forming silver nanoparticles runs at a pH of 6.0 to 11.0 for citric acid and 7.0 to 11.0 for 49 malic acid. Increasing the pH can also affect the particle size of silver nanoparticles, where the higher 50 51 the pH value, the smaller the particle size [14].

Based on the description above, researchers are interested in conducting research in the form of 52 optimizing the effect of temperature and pH on lmax surface plasmon resonance (SPR) and absorbance 53 in the manufacture of silver nanoparticles using ethanol extract of young areca nut seeds as a 54 55 bioreductor agent. Optimization was carried out using the response surface methodology (RSM) central composite design (CCD) method for temperature and pH. The temperature and pH used in this 56 study were  $30 - 70^{\circ}$ C respectively and the pH range was 7.5 - 10.5 referring to the research by 57 Prodjosantoso et al. [15] and Seifipour et al. [16] with slight modifications. The formula of silver 58 nanoparticles with optimum temperature and pH will be further characterized in particle size, 59 polydispersity index (PDI), zeta potential and thermodynamic stability tests using the cycling test 60 61 method.

### 63 Materials and Methods

### 64 Materials

65 The materials used in this study were Areca catechu L. seed s, AgNO<sub>3</sub> (Emsure , Indonesia), 70%

- ethanol (Bratachem, Indonesia), absolute ethanol (Emsure, Indonesia), NaOH 0.1 M (Bratachem,
- 67 Indonesia), catechins (Sigma-Aldrich, Singapore), and aqua deionized (Bratachem, Indonesia).
- 68

### 69 Preparation of Ethanolic Extract of *Areca catechu* L. Seed

70 Areca catechu L. seeds were washed, dried and crushed to obtain Simplicia powder. The simplicia

powder was macerated using 70% ethanol solvent and stored in a dark place for 48 hours. The filtrate

72 was then evaporated using a rotary evaporator to obtain a thick extract.

73 Quantification of Total Phenolic Content in Extract

74 The measurement of total phenolic content in the extract was carried out based on the research of Indarti

et al. [17] and Sudirman et al. [18] with modifications. The standard used in this procedure is catechin.

The extract is put at 10 mg into a 100 mL measuring flask, then the volume is made up to obtain a 100

- mg/mL concentration. The absorbance of the sample solution was measured using a UV-Vis
  spectrophotometer at 281 nm. The formula can calculation the total phenolic concentration (TPC) in
  extract in Equation 1.
- 80
- 81 82

# $TPC = \underline{volume (mL) \ x \ concentration (mq/mL) \ x \ dilute \ factor} \qquad (1)$ $g \ extract$

83

### 84 Formula of Silver Nanoparticles

The formula for making silver nanoparticles using the ethanol extract of *Areca catechu* L. refers to research by Apriani *et al.* [19]. The concentration of silver nitrate (AgNO<sub>3</sub>) used was 3 mM and 3 mL of extract at a concentration of 10% w/v with a ratio of 1:9.

88

### 89 Temperature and pH Optimization

Optimization of temperature and pH in the manufacture of silver nanoparticles was carried out using the response surface methodology (RSM) - central composite design (CCD) method referring to the research of Prodjosantoso *et al.* [15] and Seifipour *et al.* [16] with slight modifications. The levels used consist of low, middle, and high levels. The design of the levels for each factor, namely temperature and pH can be seen in Table I. Based on the conditions in Table I, nine formulas
were produced for manufacturing silver nanoparticles which can be seen in Table II.

96

### 97 Determination of Surface Plasmon Resonance

Surface plasmon resonance was determined by observing the wavelength and absorbance of the 98 sample solution in the range of 370 - 600 nm using a UV-Vis Spectrophotometer. The blank 99 used in this procedure is distilled water. Determination of Optimum Conditions for Silver 100 Nanoparticles Optimum conditions (temperature and pH) were determined to manufacture silver 101 nanoparticles by analyzing surface plasmon resonance data using the Design Expert 13® 102 program. The program will examine the effect of temperature and pH factors on surface plasmon 103 104 resonance's wavelength response and absorbance. The program suggests optimum conditions when these conditions have a desirability value close to 1. 105

106

### 107 Characterization of Silver Nanoparticles at Optimum Condition

Silver nanoparticles under optimum conditions were subjected to further characterization, such as the determination of surface plasmon resonance, particle size, polydispersity index, and zeta potential. Particle size, polydispersity index, and zeta potential were measured using a Particle Size Analyzer. The sample solution is diluted using aqua deionized. Particle size and polydispersity index were measured at a scattering angle of 90°, while measured the zeta potential at a scattering angle of 173° [22].

114

### 115 Stability Test of Silver Nanoparticles at Optimum Condition

A stability test was carried out using the cycling test method. The cycling test was carried out at temperatures 4 °C with storage at each temperature for 24 hour. The procedure was repeated for six cycles [23.24]. Organoleptic observation, pH, and surface plasmon resonance were observed in cycles 0 and 6.

120 Data Analysis

121 Data analysis for optimization was performed using program Design-Expert 13<sup>®</sup>.

- 122
- 123
- 124 **Results and Discussion**

### 125 Areca catechu L. Seeds Extract

126 The *Areca catechu* L. extract obtained in this study had a distinctive odour, brown colour and 127 thick. The total phenolic content obtained was  $111.4 \pm 0.411$  mg CE/g extract. The results were not 128 much different from those of Zhang *et al.* [25].

129 130

### 131 Silver Nanoparticles

132 Areca catechu L. seed extract is a bioreductor in forming silver nanoparticles. The silver nanoparticle preparations obtained in this study were blackish- brown in colour. Areca catechu 133 L. seed extract can act as a bioreductor due to the presence of phenolic compounds. Phenolics 134 135 can actively chelate and reduce metal ions into nanoparticles due to the presence of carbonyl group or phi (p) electrons. The transformation of the phenolic tautomer from the enol to the keto 136 form by releasing reactive hydrogen atoms can reduce metal ions to form nanoparticles [11]. An 137 overview of the mechanism for creating silver nanoparticles from catechin compounds can be 138 seen in Figure 1. 139

140

The formation of silver nanoparticles can be observed from surface plasmon resonance events. 141 Surface plasmon resonance (SPR) is a free electron resonance oscillation on a metal surface layer 142 that is excited by an incident light source [26]. The oscillation is determined by the absorption of the 143 wavelength obtained [27]. Surface plasmon resonance (SPR) wavelengths are 400 - 450 nm due 144 to the interaction between light and surface electrons moving from silver nanoparticles [28]. In 145 addition, the absorbance produced at surface plasmon resonance wavelengths can describe the 146 colour intensity and the number of silver nanoparticles formed during the synthesis process. The 147 higher the absorbance, the more silver nanoparticles are formed. The surface plasmon resonance 148 results obtained in this study were in the wavelength range of 424 - 431 nm and absorbance of 149 150 0.431 - 1.265 (Table III).

151

The maximum wavelength and absorbance data from the SPR obtained are closely related to each formula's different temperature and temperature conditions. Based on Figure 2, F3 has the highest absorbance of 1.265 while F9 has the lowest wavelength of 423 nm.

155

156 The Role of Temperature and pH to λmax of Silver Nanoparticles

157

The role of temperature and pH on the maximum wavelength of silver nanoparticles was 158 analyzed using the Design Expert 13<sup>®</sup> program. The initial step is to analyze the model and then 159 analyze the response. The model analysis obtained at the maximum wavelength response can be 160 seen in Table IV. Model analyzing on lmax has an R<sup>2</sup> value of more than 0.7 namely 0.9161, 161 which shows that 91.61% of the data on the lmax response is influenced by temperature and pH 162 factors. In comparison, the remaining 8.39% is an estimation error. This result is supported by 163 the normal curve plot of residuals (Figure 3A) which shows that the data points are close to a straight 164 line so that the data can be said to be normally distributed. Furthermore, the adjusted R<sup>2</sup> and 165 predicted  $R^2$  values obtained in the lmax model analyzing have a difference between adjusted  $R^2$ 166 and predicted  $R^2$  values illustrates the similarity of the model obtained from the relationship between 167 temperature and pH to the lmax response. This result is supported by the predicted vs. actual curve 168 in Figure 3B. The predicted vs. actual curve illustrates the results of the adjusted R<sup>2</sup> and predicted 169  $R^2$  values with points to the right and left of the line. The closer the distance between the dots to 170 the right and left of the line indicates that the data values from the research are more precise than 171 those predicted by the system, where 86.58% of the existing data already represents the population 172 and can explain a good linear relationship. The adequate value obtained in the model analysis is 173 more than 4, which is 11.4290. The greater the adequate precision value, the more resistant the 174 model will be to noise (disturbance). In addition, the p-value of less than 0.05 in the model 175 indicates that the model used significantly affects the lmax response. Based on the model 176 analyzing, it is said that the model is good to be continued in the response analyzing process. 177 Analyzing of the  $\lambda$ max response was carried out to observe the effect of each factor and the 178 interaction between the two on the response. Response analysis was carried out by following the 179 180 results of the ANOVA in the form of coefficient values and p-values of the temperature (A) and pH (B) factors. The results of the ANOVA analysis on the pH response can be seen in Table V. 181 Based on the results of the ANOVA analysis in Table V, temperature and pH significantly affect the 182 lmax response where the p-value obtained is less than 0.05. Factors that have a significant 183 effect are then entered into the response equation so that the response equation  $\lambda$  max is accepted, 184 namely y = 426.667 + 1.833A - 2.167B. The notation for factors A and B is (+) and (-), 185 respectively. This notation shows that the A-temperature factor has a positive correlation, meaning 186 187 that the greater the A value, the higher the resulting lmax value, In contrast, the B-pH factor has

188 a negative correlation, meaning that the greater the pH, the smaller the resulting  $\lambda$  max value. This result was supported the research of Yeshchenko et al. [29], which showed that an increase in 189 temperature causes a redshift (a shift in wavelength to a more significant value). This condition 190 is due to the thermal volume expansion of the silver nanoparticles with increasing temperature. 191 Thermal volume expansion is when a substance experiences a change in temperature so that the 192 substance can expand (expand) or shrink (shrink) depending on the increase or decrease in 193 temperature. Research conducted by Alqadi et al. [30] also supported the effect of pH in this study. 194 The study showed that increasing the pH resulted in smaller lmax and was followed by the formation 195 of smaller silver nanoparticle sizes. According to Sathishkumar et al. [31], when the pH is acidic, the 196 aggregation of silver nanoparticles to form larger nanoparticles is believed to be preferable to the 197 198 stages of creating new nanoparticles (nucleation). However, at alkaline pH, it will show a large number of functional groups available for silver binding, thereby facilitating a higher number of Ag<sup>+</sup> 199 ions to bind and subsequently form a large number of smaller diameter nanoparticles. 200

201

### 202 The Role of Temperature and pH to Absorbance of Silver Nanoparticle

203 The model analysis obtained at the maximum wave- length response can be seen in Table VI. The model analysis results indicate that the model is robust and significant enough to proceed to the 204 response analysis process. Model analysis on the absorbance response has an R<sup>2</sup> value of 0.9700. 205 which indicates that 97% of the data on the absorbance response is influenced by temperature and pH 206 factors. In comparison, the remaining 3% is an estimation error. This result is supported by the 207 208 normal plot of the residual curve (Figure 4A), where the data points are close to a straight line so that the data can be said to be normally distributed. Furthermore, the adjusted R<sup>2</sup> and predicted 209  $R^2$  values obtained in the absorbance model analysis have a difference of less than 0.2 namely 210 211 0.0697, so there is a similarity in the model obtained from the relationship between temperature and pH on the absorbance response. This result is supported by the predicted vs. 212 213 actual curve in Figure 4B, which shows the distance between the points to the right and left of the near line where 95.19% of the existing data already represents the population and can explain a 214 good linear relationship. The adeq precision value obtained in the model analysis is more than 4 215 216 namely 18.5757 so the model is robust against noise (disturbance). In addition, the p-value of less than 0.05 in the model indicates that the model used significantly affects the absorbance response. The 217 absorbance response analysis was continued, and the results obtained can be seen in Table VII. 218

significant effect on the absorbance response. The absorbance response equation obtained is y =
0.930 + 0.094A + 0.346B - 0.146B2. The notation on an equation for factors A and B is (+), but
there is a new factor, namely B2, which has a value of (-). Factor B2, marked (-), means that when
the pH used is too high, the absorbance produced will be smaller. When viewed from the p-value,
factor B has the smallest p-value, so that factor B has the greatest influence on the absorbance

Based on the results of the ANOVA analysis in Table VII, temperature and pH have a

response.

219

- The temperature parameter is directly proportional to the absorbance value. This shows that when there is an increase in temperature, the absorbance value of the silver nanoparticles produced will increase. Based on research by Prodjosantoso *et al.* [15], when the temperature increases, the formation rate of silver nanoparticles will be faster so that many silver nanoparticles are formed.
- 230 The more silver nanoparticles formed, the higher the absorbance [32].
- 231 In addition, the pH parameter is also directly proportional and has the most significant effect on the 232 absorbance value. This shows that when there is an increase in pH, the absorbance value of the silver nanoparticles will be higher. These results are supported by research by Alqadi et al. [30], 233 which showed that an increase in pH resulted in a higher absorbance of silver nanoparticles. In 234 235 addition, Roopan et al. [33] research showed that at pH 2 no reaction to form silver nanoparticles occurred, while at pH 11, the silver nanoparticles formed were highly monodispersed. Research by 236 237 Tagad et al. [34] also showed that at acidic pH (pH 4 and 6), there is no surface plasmon resonance absorption which is characteristic of silver nanoparticles. The formation of silver 238 nanoparticles is more suitable to occur in alkaline pH conditions so that many small silver 239 240 nanoparticles are formed, and the resulting absorbance value will be higher [35]. However, the synthesis of silver nanoparticles at extreme alkaline pH (> 12) can produce low stability and 241 cause aggregation [36]. 242
- 243

### 244 Optimum Condition for Silver Nanoparticles

Determination of optimum conditions is done by looking at the system's desirability value closest to 1. The desirability value close to 1 indicates that the situation meets the target desired by the researcher. Based on the analysis results, the optimum conditions for making silver nanoparticles are recommended for the system to be at a temperature of 30°C and pH of

249 10.5 with a desirability value of 0.932.

### 251 Characterization of Silver Nanoparticles at Optimum Condition

Silver nanoparticles at optimum conditions were tested for characterization, such as surface plasmon 252 resonance, particle size, polydispersity index, and zeta potential. The results of the 253 characterization can be seen in Table VIII. Silver nanoparticles in the optimum condition have 254 255 an excellent  $\lambda$  max value and absorbance for silver nanoparticles. The size of the silver nanoparticles obtained was  $161.7 \pm 46.1$  nm. The particles size results obtained were smaller 256 than previous studies conducted by Choi et al. [37] at 233.4 – 238 nm and Bhat et al. [38] at 257 258 553 - 610 nm. The polydispersity index value was  $0.286 \pm 0.035$  indicating that the particles 259 size obtained was uniform, The zeta potential value obtained from the optimum formula for silver nanoparticles was  $-16.1 \pm 3.7$  mV. This results indicate that the zeta potential of the silver 260 nanoparticle optimum formula is still relatively stable. The negative zeta potential value is due 261 to the alkaline pH (pH 10.5) during the biosynthesis of silver nanoparticles. With an increase in 262 263 pH (alkaline pH), the organic functional groups on the surface of silver nanoparticles reach a 264 higher level of deprotonation (release of H<sup>+</sup> protons) so that the amount of negative surface charge increases simultaneously [39]. The characterization results obtained have met the requirements 265 266 for nanoparticle preparations.

267

### 268 Stability of Silver Nanoparticles at Optimum Condition

The silver nanoparticles in optimum condition were tested for stability by cycling for six cycles. 269 The stability results of silver nanoparticles can be seen in Table IX. Organoleptic results showed 270 271 the formation of a slightly precipitate in the  $6^{th}$  cycle. The results obtained are better than the previous reference research conducted by Apriani et al. [19], which showed many deposits 272 273 formed on colloidal silver nanoparticles in the 6th cycle. The difference in stability results obtained can be due to differences in the conditions for synthesizing silver nanoparticles. This study 274 used a temperature of 30 °C and pH 10.5 as the optimum condition, while the study of Apriani et 275 al. [19] used a temperature of 60 °C and pH 9. Research by Pinto et al. [40] showed that high 276 temperatures can produce unstable silver nanoparticles and can cause damage or decomposition 277 of plant secondary metabolites, which function as stabilizing agents (capping agents). Changes 278 279 in the pH value in this study were also said to be relatively stable, where the pH changed from  $10.50 \pm 0.008$  to  $10.10 \pm 0.008$ . In contrast to previous research from the same researcher, Apriani 280

et al. [19], with the conditions of nanoparticle synthesis at a temperature of 60 °C and pH 9 the 281 decrease in pH was significant, namely pH 9. This stable pH also affect changes in  $\lambda$ max value and 282 absorbance. The shift in  $\lambda$  max in the 6<sup>th</sup> cycle was not too far and was still in the range of silver 283 nanoparticles. In contrast, the absorbance in the 6<sup>th</sup> cycle increased, indicating that more and 284 more silver nanoparticles were formed, as shown in Figure 5. This is because phenolic stability 285 depends on pH. Phenolic compounds are in the enol form at acidic pH and are in the keto form 286 287 at an alkaline pH. The keto form of phenolic is helpful as a stabilizing agent (capping agent). In this study, the reaction to form silver nanoparticles continued because the environmental pH was 288 relatively stable at around 10. 289

290

### 291 Conclusions

Temperature and pH conditions in manufacturing silver nanoparticles have been shown to affect the 292 surface plasmon resonance characteristics. The temperature has positive effect on  $\lambda$ max and the 293 absorbance of SPR, while pH harms  $\lambda$  max and has positive impact on the absorbance of SPR. 294 The optimum temperature and pH conditions for the synthesis of silver nano- particles using 295 296 ethanol extract of young areca nut seeds were found at 30 °C and pH 10.5 with a surface plasmon resonance silver nanoparticle  $\lambda$ max value of 423 nm and an absorbance of 1.148. Particle size, 297 polydispersity index (PDI), and zeta potential of silver nanoparticles at optimum condition were 298  $161.7 \pm 46.1$  nm,  $0.286 \pm 0.035$  and  $-16.1 \pm 3.7$  mV respectively. The stability of the silver 299 300 nanoparticles at the optimum conditions produced was relatively stable, characterized by not too many organoleptic changes, pH, which was still stable in the range of 10 small wavelength shifts, and 301 increased absorbance values. 302

303

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- 306

### **307 Conflict of interest**

- 308 The authors declare no conflict of interest.
- 309
- 310
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#### Table III

Data maximum wavelength and absorbance of silver nanoparticles
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Formula	Temperature (°C)	pН	Max. Wavelength (nm)	Absorbance
1	70	7.5	429	0.446
2	30	9	424	0.769
3	70	10.5	427	1.265
4	50	9	425	0.962
5	50	7.5	431	0.431
6	70	9	430	1.059
7	30	10.5	424	1.004
8	30	7.5	427	0.435
9	50	10.5	423	1.121



Table IV Model Analyzing of  $\lambda$ max Responses

Response	-	Parameter			
	R <sup>2</sup>	Adjusted R <sup>2</sup>	Predicted R <sup>2</sup>	Adequate precision	p-value
Max. Wavelength	0.9161	0.8658	0.6980	11.4290	0.0192





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#### Table V Response analyzing of $\lambda max$

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Responses	Intercept (nm)	Α	В
<b>λmaks SPR</b>	426.667	1.833	-2.167
p-values		0.0397*	0.0213*

490	A: Temperature; B: pH; *factors that have a significant effect (p-value $< 0.05$ )
491	
400	

#### Table VI

Model analysis of absorbance responses 

Response		Parameter			
	R <sup>2</sup>	Adjusted R <sup>2</sup>	Predicted R <sup>2</sup>	Adequate precision	p-value
Absorbance	0.9700	0.9519	0.8822	18.5757	0.0003





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