## JURNAL SYARAT KHUSUS

# THE ROLE OF TEMPERATURE AND pH IN THE SYNTHESIS OF SILVER NANOPARTICLES USING ARECA CATECHU L. SEED EXTRACT AS BIOREDUCTOR

### MARDIYANTO MARDIYANTO, ELSA FITRIA APRIANI, FITO PRATAMA HEYLKEN

🖹 🧱 Farmacia 🗙 +						0	~ -	đ
	www.scimagojr.com/journalsearch.php?q=218108	tip=sid&clean=0	图 ¾ ☆	Q Search			Ģ	ំំំំ
*								
9 0								
	COUNTRY	SUBJECT AREA AND CATEGORY	PUBLISHER	SJR 2024				
	Romania	Pharmacology, Toxicology and Pharmaceutics	Romanian Society for Pharmaceutical Sciences	0.251	Q3			
	Universities and research institutions in Romania	<ul> <li>Pharmacology, Toxicology and Pharmaceutics</li> </ul>						
	Media Ranking in Romania	(miscellaneous)		H-INDEX				
				33				
	PUBLICATION TYPE	ISSN	COVERAGE	INFORMATION				
	Journals	00148237	1945-1946, 1973-1987, 2003, 2008-2024	Homepage	10.1 X - 10.			
				How to publish in	this journal			
					negerin sedana			
	SCOPE							
	FARMACIA publishes original resea	rch papers, invited topical reviews and	editorial commentaries and news, with	emphasis on conce	ptual novelty			
	and scientific quality. Main research personalized medicine, nanostructu	n areas are focused on: pharmacology, ires, nutraceuticals, biochemistry and b	toxicology, medicinal chemistry, biophar piotechnology, Manuscripts submitted to	rmacy, drug design, the Journal are on	drug delivery, ly accepted after			
	the peer review precess. The papers humans, and the International guide	s should have not been published in an elines as accepted principles for the us	y other journal. The recommendations o e of experimental animals should be Ads	f the Declaration of by clicklo	Helsinki, for			
	86				Authomatical Coffue			
\$	Ő			<u> </u>	/ Mathematical Softwa	ne		1
	Q Sear	rch 🛛 🞯 🖬 🔹 🥠 🕻	a 🖾 📲 🖾 🔕 💆 📮 🕯	9		^ O 🖾		1:20 Pi 6/29/202
10.00 #				6	4G+ 4		7.4	
13:29 ₩				C	) till ann a till (	י יב ווי	4%	
	com/courcoid/219	10			L 1	ា	:	
	5.COIII/SOUICEIU/210	10				٣	•	
Farmacia					CiteScore 2024		()	
Years currently cover	red by Scopus: from 194	5 to 1946 from 1973 to 1	987 2003 from 2008 to 2	025	2.2			
Publisher: Romania	in Society for Pharmaceu	tical Sciences	567, 2005, 110111 2000 to 2	025				
ISSN: 0014-8237	in boolety for Finannacea				SJR 2024			$\bullet$
Subject area: (Pharm	acology, Toxicology and Pharmace	utics: General Pharmacoloay. Toxic	cology and Pharmaceutics		0.251		U	
Source type: lourne	1							
					SNIP 2024		-	
View all documents >	Set document alert 📃	Save to source list			0.350		()	
CiteScore CiteSco	re rank & trend Scopy	is content covergae						

3:28 🍄 released. V	'iew CiteScore methodology >				🎯 🖼 👫 🛄 📶	<mark>ແໄ ທີ </mark> ອີ 74 <sub>%</sub>	
	1 result	▲ Download	Scopus Source List	: (i) Learn m	iore about Sco	opus Source List	t
^	Source title V	CiteScore V	Highest percentile ↓	View metr Citations 2021-24 $\downarrow$	ics for year: Documents 2021-24 ↓	2024 ♥ % Cited ↓ >	-
5	☐ 1 Farmacia	2.2	48% 40/77 General Pharmacology, Toxicology and Pharmaceutics	1.318	599	63	_

#### **ORIGINAL ARTICLE**

### THE ROLE OF TEMPERATURE AND pH IN THE SYNTHESIS OF SILVER NANOPARTICLES USING *ARECA CATECHU* L. SEED EXTRACT AS BIOREDUCTOR

#### MARDIYANTO MARDIYANTO, ELSA FITRIA APRIANI\*, FITO PRATAMA HEYLKEN

Department of Pharmacy, Faculty of Mathematics and Natural Sciences, Sriwijaya University, South Sumatra, Indonesia

\*corresponding author: elsafitria@mipa.unsri.ac.id

Manuscript received: October 2022

#### Abstract

Silver nanoparticles are prepared using natural bioreductors such as *Areca catechu* ethanol extract, which is rich in phenolic content. This study aims to optimize the temperature and pH conditions for synthesizing silver nanoparticles using the response surface methodology (RSM) - central composite design (CCD) method. The phenolic content of the *Areca catechu* ethanol extract was 111.14  $\pm$  3.41 mg CE/g extract. Temperature and pH conditions significantly affect maximum wavelength and absorbance values. The optimum condition was obtained at a temperature of 30oC and a pH of 10.5. Silver nanoparticles at optimum conditions had a wavelength of 423 nm, absorbance was 1.148, particle size was 161.7  $\pm$  46.1 nm, PDI was 0.286  $\pm$  0.035, and zeta potential was - 16.1  $\pm$  3.7 mV. The stability of silver nanoparticles at the optimum conditions produced is relatively stable, characterized by no significant changes in organoleptic, pH, and wavelength, but the absorbance value has increased. The resulting silver nanoparticles have good characteristics and good stability.

#### Rezumat

Nanoparticulele de argint pot fi preparate folosind bioreducători naturali, cum este extractul etanolic de *Areca catechu*, bogat în fenoli. În cadrul acestui studiu ne-am propus optimizarea condițiilor de temperatură și pH pentru sintetizarea nanoparticulelor de argint folosind metoda suprafeței de răspuns (MSR). Conținutul fenolic al extractului etanolic de *Areca catechu* a fost de 111,14  $\pm$  3,41 mg/g extract. Temperatura și pH-ul afectează semnificativ reacția, condițiile optime de formare a nanoparticulelor fiind la 30°C și un pH de 10,5. Stabilitatea nanoparticulelor de argint obținute este relativ optimală, caracterizată prin faptul că nu există modificări semnificative în ceea ce privește proprietățile organoleptice și pH-ul.

Keywords: Areca catechu L. seed, pH, response surface methodology, silver nanoparticles, temperature

#### Introduction

Nanotechnology in the pharmaceutical field is currently developing very rapidly. Silver nanoparticles are among the most researched and set in the medical world because they have been shown to have anti-bacterial, anti-inflammatory, anti-angiogenesis, anti-fungal, antiviral, and antiplatelet activities [1-4].

The production of silver nanoparticles has shifted towards green synthesis methods that are more environmentally friendly. The green synthesis method requires the help of secondary metabolites in plants as a source of bioreductors [5]. Secondary metabolites that can act as bioreductors usually act as antioxidants, such as flavonoids, phenolics, and alkaloids [6]. *Areca catechu* L. is a plant rich in alkaloid compounds such as guvacine, guvacoline, arecaidine, and arecoline and also rich in polyphenolic compounds such as catechin and quercetin [7, 8].

The formation of silver nanoparticles through the green synthesis method can be influenced by several factors, such as temperature and pH [9]. Temperature is another critical factor affecting the formation of nanoparticles in plant extracts [10]. In general, an

increase in temperature will increase nanoparticle synthesis's reaction rate and efficiency [11]. This statement is proven by research conducted by Jiang et al [12], where a temperature of 17 to 55°C will accelerate the reaction and cause the particle size to increase from 90 nm to 180 nm. Besides temperature, changes in pH will also affect the process of forming silver nanoparticles. Changes in pH will result in changes in the natural phytochemical charge contained in the extract. This effect will affect its ability to bind and reduce metal cations while synthesizing silver nanoparticles. According to research by Marciniak et al. [13], the reaction for forming silver nanoparticles runs at a pH of 6.0 to 11.0 for citric acid and 7.0 to 11.0 for malic acid. Increasing the pH can also affect the particle size of silver nanoparticles, where the higher the pH value, the smaller the particle size [14]. Based on the description above, researchers are interested in conducting research in the form of optimizing the effect of temperature and pH on  $\lambda$ max surface plasmon resonance (SPR) and absorbance in the manufacture of silver nanoparticles using ethanol extract of young areca nut seeds as a bioreductor agent.

Optimization was carried out using the response surface methodology (RSM) - central composite design (CCD) method for temperature and pH. The temperature and pH used in this study were 30 - $70^{\circ}$ C, respectively, and the pH range was 7.5 - 10.5, referring to the research by Prodjosantoso *et al.* [15] and Seifipour *et al.* [16] with slight modifications. The formula of silver nanoparticles with optimum temperature and pH will be further characterized in particle size, polydispersity index (PDI), zeta potential and thermodynamic stability tests using the cycling test method.

#### **Materials and Methods**

#### Materials

The materials used in this study were *Areca catechu L*. seed s, AgNO3 (Emsure<sup>®</sup>, Indonesia), 70% ethanol (Bratachem<sup>®</sup>, Indonesia), absolute ethanol (Emsure<sup>®</sup>, Indonesia), NaOH 0.1 M (Bratachem<sup>®</sup>, Indonesia), catechins (Sigma-Aldrich<sup>®</sup>, Singapore), and aqua deionized (Bratachem<sup>®</sup>, Indonesia).

*Preparation of Ethanolic Extract of Areca catechu* L. Seed

*Areca catechu* L. seeds were washed, dried and crushed to obtain Simplicia powder. The simplicia powder was macerated using 70% ethanol solvent and stored in a dark place for 48 hours. The filtrate was then evaporated using a rotary evaporator to obtain a thick extract.

Determination of Total Phenolic Content in Extract The measurement of total phenolic content in the extract was carried out based on the research of Indarti *et al.* [17] and Sudirman *et al.* [18] with modifications. The standard used in this procedure is catechin. The extract is put at 10 mg into a 100 mL measuring flask, then the volume is made up to obtain a 100  $\mu$ g/mL concentration. The absorbance of the sample solution was measured using a UV-Vis Spectrophotometer at 281 nm. The formula can calculate the total phenolic content in the extract in Equation 1.

 $TPC = \frac{Volume \ (mL)x \ Concentration \ \left(\frac{mg}{mL}\right)x \ dilute \ factor}{g \ Extract}. (1)$ 

#### Formula of Silver Nanoparticles

The formula for making silver nanoparticles using the ethanol extract of *Areca catechu* L. refers to research by Apriani *et al.* [19]. The concentration of silver nitrate (AgNO<sub>3</sub>) used was 3 mM and 3 mL of extract at a concentration of 10% w/v with a ratio of 1:9.

#### Temperature and pH Optimization

Optimization of temperature and pH in the manufacture of silver nanoparticles was carried out using the response surface methodology (RSM) - central composite design (CCD) method referring to the research of Prodjosantoso *et al.* [15] and Seifipour *et al.* [16] with slight modifications. The levels used consist of low, middle, and high levels. The design of the levels for each factor, namely temperature and pH can be seen in Table I. Based on the conditions in Table I, nine formulas were produced for manufacturing silver nanoparticles which can be seen in Table II.

#### Table I

Level design for temperature and pH

Factor	Level				
ractor	Low	Medium	High		
Temperature	30	50	70		
рН	7.5	9	10.5		

#### Table II

Formula and condition of silver nanoparticles

Formula	AgNO <sub>3</sub> 3 mM (mL)	Extract 10% b/v (mL)	Temp (°C)	pН
1	27	3	70	7.5
2	27	3	30	9
3	27	3	70	10.5
4	27	3	50	9
5	27	3	50	7.5
6	27	3	70	9
7	27	3	30	10.5
8	27	3	30	7.5
9	27	3	50	10.5

#### Preparation of Silver Nanoparticles

As much as 3 mL of 10% w/v young areca seed ethanol extract was added drop by drop in 27 mL of 3 mM silver nitrate solution at 750 rpm for 15 minutes [19]. The solution was heated and maintained at the temperature and pH according to Table II using 0.1 M NaOH. The formation of yellow-brown, reddishbrown, or dark brown colour indicated the formation of silver nanoparticles [20, 21].

#### Determination of Surface Plasmon Resonance Surface plasmon resonance was determined by observing the wavelength and absorbance of the sample solution in the range of 370 - 600 nm using a UV-Vis Spectrophotometer. The blank used in this procedure is distilled water.

Determination of Optimum Conditions for Silver Nanoparticles

Optimum conditions (temperature and pH) were determined to manufacture silver nanoparticles by analyzing surface plasmon resonance data using the Design Expert 13<sup>®</sup> program. The program will examine the effect of temperature and pH factors on surface plasmon resonance's wavelength response and absorbance. The program suggests optimum conditions when these conditions have a desirability value close to 1.

#### Characterization of Silver Nanoparticles at Optimum Condition

Silver nanoparticles under optimum conditions were subjected to further characterization, such as the determination of surface plasmon resonance, particle size, polydispersity index, and zeta potential. Particle size, polydispersity index, and zeta potential were measured using a Particle Size Analyzer. The sample solution is diluted using aqua deionized. Particle size and polydispersity index were measured at a scattering angle of 90°, while measured the zeta potential at a scattering angle of 173° [22].

# Stability Test of Silver Nanoparticles at Optimum Condition

A stability test was carried out using the cycling test method. The cycling test was carried out at temperatures 4°C and 40°C with storage at each temperature for 24 hours. The procedure was repeated for six cycles [23, 24]. Organoleptic observations, pH, and surface plasmon resonances were observed in cycles 0 and 6. *Data Analysis* 

Data analysis for optimization was performed using program Design-Expert 13<sup>®</sup>.

#### **Results and Discussion**

#### Areca catechu L. Seeds Extract

The Areca catechu L. extract obtained in this study had a distinctive odour, brown colour and thick. The total phenolic content obtained was  $111.4 \pm 0.411$ mg CE/g extract. The results were not much different from those of Zhang *et al.* [25], who got a total phenolic content of  $111.14 \pm 3.41$  mg CE/g extract. *Silver Nanoparticles* 

Areca catechu L. seed extract is a bioreductor in forming silver nanoparticles. The silver nanoparticle preparations obtained in this study were blackishbrown in colour. Areca catechu L. seed extract can act as a bioreductor due to the presence of phenolic compounds. Phenolics can actively chelate and reduce metal ions into nanoparticles due to the presence of carbonyl groups or phi ( $\pi$ ) electrons. The transformation of the phenolic tautomer from the enol to the keto form by releasing reactive hydrogen atoms can reduce metal ions to form nanoparticles [11]. An overview of the mechanism for creating silver nanoparticles from catechin compounds can be seen in Figure 1.



Figure 1. Silver nanoparticle formation mechanism

The formation of silver nanoparticles can be observed from surface plasmon resonance events. Surface plasmon resonance (SPR) is a free electron resonance oscillation on a metal surface layer that is excited by an incident light source [26]. The oscillation is determined by the absorption of the wavelength obtained [27]. Surface plasmon resonance (SPR) wavelengths are 400 - 450 nm due to the interaction between light and surface electrons moving from silver nanoparticles [28]. In addition, the absorbance produced at surface plasmon resonance wavelengths can describe the colour intensity and the number of silver nanoparticles formed during the synthesis process. The higher the absorbance, the more silver nanoparticles are formed. The surface plasmon resonance results obtained in this study were in the wavelength range of 424 - 431 nm and absorbance of 0.431 - 1.265 (Table III).

#### Table III

Data maximum wavelength and absorbance of si						
Formula	Temperature (°C)	pН	Max. Wavelength (nm)	Absorbance		
1	70	7.5	429	0.446		
2	30	9	424	0.769		
3	70	10.5	427	1.265		
4	50	9	425	0.962		
5	50	7.5	431	0.431		
6	70	9	430	1.059		
7	30	10.5	424	1.004		
8	30	7.5	427	0.435		
9	50	10.5	423	1.121		

The maximum wavelength and absorbance data from the SPR obtained are closely related to each formula's different temperature and temperature conditions. Based on Figure 2, F3 has the highest absorbance of 1.265 while F9 has the lowest wavelength of 423 nm.



UV-Vis Spectrum of Silver Nanoparticles

*The Role of Temperature and pH to λmax of Silver Nanoparticle* 

The role of temperature and pH on the maximum wavelength of silver nanoparticles was analyzed using

the Design Expert 13<sup>®</sup> program. The initial step is to analyze the model and then analyze the response. The model analysis obtained at the maximum wavelength response can be seen in Table IV.

# Table IV Model Analysis of λmax Responses

Response		Parameter					
	R <sup>2</sup>	Adjusted R <sup>2</sup>	Predicted R <sup>2</sup>	Adequate precision	p-value		
Max. Wavelength	0.9161	0.8658	0.6980	11.4290	0.0192		

Model analysis on  $\lambda$ max has an R<sup>2</sup> value of more than 0.7, namely 0.9161, which shows that 91.61% of the data on the  $\lambda$ max response is influenced by temperature and pH factors. In comparison, the remaining 8.39% is an estimation error. This result is supported by the normal curve plot of residuals (Figure 3A) which shows that the data points are close to a straight line so that the data can be said to be normally distributed. Furthermore, the adjusted R<sup>2</sup> and predicted R<sup>2</sup> values obtained in the  $\lambda$ max model analysis have a difference of less than 0.2 namely 0.1678. The difference between adjusted R2 and predicted R2 values illustrates the similarity of the model obtained from the relationship between temperature and pH to the  $\lambda$ max response. This result is supported by the predicted vs. actual curve in Figure 3B. The predicted vs. actual curve

illustrates the results of the adjusted R<sup>2</sup> and predicted  $R^2$  values with points to the right and left of the line. The closer the distance between the dots to the right and left of the line indicates that the data values from the research are more precise than those predicted by the system, where 86.58% of the existing data already represents the population and can explain a good linear relationship. The adequate value obtained in the model analysis is more than 4, which is 11.4290. The greater the adequate precision value, the more resistant the model will be to noise (disturbance). In addition, the p-value of less than 0.05 in the model indicates that the model used significantly affects the  $\lambda$ max response. Based on the model analysis, it is said that the model is good to be continued in the response analysis process.



Normal Plot of residual (A) and predicted vs. actual (B) curves of  $\lambda$  max

Analysis of the  $\lambda$ max response was carried out to observe the effect of each factor and the interaction between the two on the response. Response analysis was carried out by following the results of the ANOVA in the form of coefficient values and pvalues of the temperature (A) and pH (B) factors. The results of the ANOVA analysis on the pH response can be seen in Table V.

	Т	ab	le	V
--	---	----	----	---

	Respo	onse analys	sis of $\lambda$ max
Respon	Intercept (nm)	A	В
λmaks SPR	426.667	1.833	-2.167
p-values		0.0397*	0.0213*

A: Temperature; B: pH; \*factors that have a significant effect (p-value < 0.05)

Based on the results of the ANOVA analysis in Table V, temperature and pH significantly affect the  $\lambda$ max response where the p-value obtained is less than 0.05. Factors that have a significant effect are then entered into the response equation so that the response equation  $\lambda$ max is accepted, namely y = 426.667 + 1.833A - 2.167B. The notation for factors A and B is (+) and (-), respectively. This notation shows that the A-temperature factor has a positive correlation, meaning that the greater the A value, the higher the resulting  $\lambda$ max value. In contrast, the B-pH factor has a negative correlation, meaning that the greater the pH, the smaller the resulting  $\lambda$ max value. This result is supported by the research of Yeshchenko *et al.* [29], which showed that an increase in temperature causes a redshift (a

shift in wavelength to a more significant value). This condition is due to the thermal volume expansion of the silver nanoparticles with increasing temperature. Thermal volume expansion is when a substance experiences a change in temperature so that the substance can expand (expand) or shrink (shrink) depending on the increase or decrease in temperature. Research conducted by Alqadi *et al.* [30] also supported the effect of pH in this study. The study showed that increasing the pH resulted in a smaller  $\lambda$ max and was followed by the formation of smaller silver nanoparticle sizes. According to Sathishkumar *et al.* [31], when the pH is acidic, the aggregation of silver nanoparticles to form larger nanoparticles is believed to be preferable

to the stages of creating new nanoparticles (nucleation). However, at alkaline pH, it will show a large number of functional groups available for silver binding, thereby facilitating a higher number of  $Ag^+$  ions to bind and subsequently form a large number of smaller diameter nanoparticles.

*The Role of Temperature and pH to Absorbance of Silver Nanoparticle* 

The model analysis obtained at the maximum wavelength response can be seen in Table VI. The model analysis results indicate that the model is robust and significant enough to proceed to the response analysis process.

#### Table VI

Model analysis of absorbance responses

Response	Parameter					
	R <sup>2</sup>	Adjusted R <sup>2</sup>	Predicted R <sup>2</sup>	Adequate precision	p-value	
Absorbance	0.9700	0.9519	0.8822	18.5757	0.0003	

Model analysis on the absorbance response has an  $R^2$  value of 0.9700, which indicates that 97% of the data on the absorbance response is influenced by temperature and pH factors. In comparison, the remaining 3% is an estimation error. This result is supported by the normal plot of the residual curve (Figure 4A), where the data points are close to a straight line so that the data can be said to be normally distributed.

Furthermore, the adjusted  $R^2$  and predicted  $R^2$  values obtained in the absorbance model analysis have a difference of less than 0.2 namely 0.0697, so there is a similarity in the model obtained from the relationship between temperature and pH on the absorbance response. This result is supported by the predicted vs. actual curve in Figure 4B, which shows the distance between the points to the right and left of the near line where 95.19% of the existing data already represents the population and can explain a good linear relationship. The adeq precision value obtained in the model analysis is more than 4 namely 18.5757 so the model is robust against noise (disturbance). In addition, the p-value of less than 0.05 in the model indicates that the model used significantly affects the absorbance response.



Figure 4.

Normal plot of residual (A) and predicted vs. actual (B) curves of absorbance

The absorbance response analysis was continued, and the results obtained can be seen in Table VII. Based on the results of the ANOVA analysis in Table VII, temperature and pH have a significant effect on the absorbance response.

FARMACIA, 2023, Vol. 71, 2

Response analysis of absorbance						
Response	Intercept (nm)	A	В	<b>B</b> <sup>2</sup>		
Absorbance	0.930	0.094	0.346	-0.146		
p-values		0.0232*	0.0001*	0.0333*		
			1 1.01	00		

.

Table VII

A: Temperature; B: pH; \*factors that have a significant effect (p-value < 0.05)

The absorbance response equation obtained is y =0.930 + 0.094A + 0.346B - 0.146B2. The notation on an equation for factors A and B is (+), but there is a new factor, namely B2, which has a value of (-). Factor B2, marked (-), means that when the pH used is too high, the absorbance produced will be smaller. When viewed from the p-value, factor B has the smallest p-value, so that factor B has the greatest influence on the absorbance response.

The temperature parameter is directly proportional to the absorbance value. This shows that when there is an increase in temperature, the absorbance value of the silver nanoparticles produced will increase. Based on research by Prodjosantoso et al. [15], when the temperature increases, the formation rate of silver nanoparticles will be faster so that many silver nanoparticles are formed. The more silver nanoparticles formed, the higher the absorbance [32].

In addition, the pH parameter is also directly proportional and has the most significant effect on the absorbance value. This shows that when there is an increase in pH, the absorbance value of the silver nanoparticles will be higher. These results are supported by research by Alqadi et al. [30], which showed that an increase in pH resulted in a higher absorbance of silver nanoparticles. In addition, Roopan et al. [33] research showed that at pH 2 no reaction to form silver nanoparticles occurred, while at pH 11, the silver nanoparticles formed were highly monodispersed. Research by Tagad et al. [34] also showed that at acidic pH (pH 4 and 6), there is no surface plasmon resonance absorption which is characteristic of silver nanoparticles. The formation of silver nanoparticles is more suitable to occur in alkaline pH conditions so that many small silver nanoparticles are formed, and the resulting absorbance value will be higher [35]. However, the synthesis of silver nanoparticles at extreme alkaline pH (> 12) can produce low stability and cause aggregation [36].

**Optimum Condition for Silver Nanoparticles** Determination of optimum conditions is done by looking at the system's desirability value closest to 1. The desirability value close to 1 indicates that the situation meets the target desired by the researcher. Based on the analysis results, the optimum conditions for making silver nanoparticles are recommended for the system to be at a temperature of 30°C and pH of 10.5 with a desirability value of 0.932.

Characterization of Silver Nanoparticles at Optimum Condition

Silver nanoparticles at optimum conditions were tested for characterization, such as surface plasmon resonance, particle size, polydispersity index, and zeta potential. The results of the characterization can be seen in Table VIII.

	Tabl				
(	Characterization	results			
Parameter	Mean ± SD				
Amax (nm)	$423\pm2.156$				
Absorbance	$1.148\pm0.076$				
Particle size (nm)	$161.7\pm46.1$				
PDI	$0.286\pm0.035$				
Zeta Potential (mV)	$-16.1 \pm 3.7$				

Silver nanoparticles in the optimum condition have an excellent  $\lambda$  max value and absorbance for silver nanoparticles. The size of the silver nanoparticles obtained was  $161.7 \pm 46.1$  nm. The particle size results obtained were smaller than previous studies conducted by Choi et al. [37] at 233.4 - 238 nm and Bhat et al. [38] at 553 - 610 nm. The polydispersity index value was  $0.286 \pm 0.035$ , indicating that the particle size obtained was uniform. The zeta potential value obtained from the optimum formula for silver nanoparticles is -  $16.1 \pm 3.7$  mV. These results indicate that the zeta potential of the silver nanoparticle optimum formula is still relatively stable. The negative zeta potential value is due to the alkaline pH (pH 10.5) during the biosynthesis of silver nanoparticles. With an increase in pH (alkaline pH), the organic functional groups on the surface of silver nanoparticles reach a higher level of deprotonation (release of H<sup>+</sup> protons) so that the amount of negative surface charge increases simultaneously [39]. The characterization results obtained have met the requirements for nanoparticle preparations.

Stability of Silver Nanoparticles at Optimum Condition The silver nanoparticles in optimum condition were tested for stability by cycling for six cycles. The stability results of silver nanoparticles can be seen in Table IX.

> Table IX The stability results

Cycle	Sedimentation	pН	λmax (nm)	Absorbance
1	None	$10.50\pm0.008$	423	1.148
6	Slight	$10.10\pm0.008$	425	1.396

ılts

Organoleptic results showed the formation of a slightly precipitate in the 6<sup>th</sup> cycle. The results obtained are better than the previous reference research conducted by Apriani *et al.* [19], which showed many deposits formed on colloidal silver nanoparticles in the 6th cycle. The difference in stability results obtained can be due to differences in the conditions for synthesizing silver nanoparticles. This study used a temperature of  $30^{\circ}$ C and pH 10.5 as the optimum conditions, while the study of Apriani *et al.* [19] used a temperature of  $60^{\circ}$ C and pH 9. Research by Pinto *et al.* [40] showed that high temperatures can produce unstable silver nanoparticles and can cause damage or decomposition of plant secondary metabolites, which function as stabilizing agents (capping agents).

Changes in the pH value in this study were also said to be relatively stable, where the pH changed from  $10.50 \pm 0.008$  to  $10.10 \pm 0.008$ . In contrast to

previous research from the same researcher, Apriani et al. [19], with the conditions of nanoparticle synthesis at a temperature of 60°C and pH 9, the decrease in pH was significant, namely pH 9.00  $\pm$  0.10 to 7.97  $\pm$ 0.03. This stable pH also affects changes in  $\lambda$ max values and absorbance. The shift in  $\lambda$ max in the 6<sup>th</sup> cycle was not too far and was still in the range of silver nanoparticles. In contrast, the absorbance in the 6<sup>th</sup> cycle increased, indicating that more and more silver nanoparticles were formed, as shown in Figure 5. This is because phenolic stability depends on pH. Phenolic compounds are in the enol form at acidic pH and are in the keto form at an alkaline pH. The keto form of phenolic is helpful as a stabilizing agent (capping agent). In this study, the reaction to form silver nanoparticles continued because the environmental pH was relatively stable at around 10.



Comparison of the UV-Vis spectra of the 1<sup>st</sup> and 6<sup>th</sup> cycles

#### Conclusions

Temperature and pH conditions in manufacturing silver nanoparticles have been shown to affect the surface plasmon resonance characteristics. The temperature has a positive effect on  $\lambda$ max and the absorbance of SPR, while pH harms  $\lambda$ max and has a positive impact on the absorbance of SPR. The optimum temperature and pH conditions for the synthesis of silver nanoparticles using ethanol extract of young areca nut seeds were found at 30°C and pH 10.5 with a surface plasmon resonance silver nanoparticle  $\lambda$ max value of 423 nm and an absorbance of 1.148. Particle size, polydispersity index (PDI), and zeta potential of silver nanoparticles at optimum conditions were  $161.7 \pm 46.1$ nm,  $0.286 \pm 0.035$ , and  $-16.1 \pm 3.7$  mV, respectively. The stability of the silver nanoparticles at the optimum conditions produced was relatively stable, characterized

by not too many organoleptic changes, pH, which was still stable in the range of 10 small wavelength shifts, and increased absorbance values.

#### Acknowledgement

This study was supported by PT. DKSH Indonesia (Malvern).

#### **Conflict of interest**

The authors declare no conflict of interest.

#### References

 Baharara J, Namvar F, Mousavi M, Ramezani T, Mohamad R, Anti-Angiogenesis Effect of Biogenic Silver Nanoparticles Synthesized Using *Saliva officinalis* on Chick Chorioalantoic Membrane (CAM). *Molecules*, 2014; 19: 13498-13508.

- Mallmann EJ, Cunha FA, Castro BN, Maciel AM, Menezes EA, Fechine PB, Antifungal activity of silver nanoparticles obtained by green synthesis. *Rev Inst Med Trop Sao Paulo*, 2015; 57(2): 165-167.
- Tyavambiza C, Elbagory AM, Madiehe AM, Meyer M, Meyer S, The Antimicrobial and Anti-Inflammatory Effects of Silver Nanoparticles Synthesised from *Cotyledon orbiculata* Aqueous Extract. *Nanomaterials*, 2021; 11: 1343.
- Massadeh M, Al-Masri M, Abu-Qatouseh LF, Antibacterial activity of *Peganum harmala* extracts and their green silver nanoparticles against acne associated bacteria. *Farmacia*, 2022, 70(5): 954-963.
- Roy A, Bulut O, Some S, Mandal AK, Yilmaz MD, Green synthesis of silver nanoparticles: biomoleculenanoparticle organizations targeting antimicrobial activity. *Royal Soc Chem Adv.*, 2019; 9: 2673-2702.
- Wulandari IO, Pebriatin BE, Valiana V, Hadisaputra S, Ananto AD, Sabarudin A, Green Synthesis of Silver Nanoparticles Coated by Water Soluble Chitosan and Its Potency as Non-Alcoholic Hand Sanitizer Formulation. *Materials*, 2022; 15: 4641.
- Jain V, Garg A, Parascandola M, Chaturvedi P, Khariwala SS, Stepanov I, Analysis of alkaloids in *Areca* nut-containing products by liquid chromatographytandem mass spectrometry. *J Agric Food Chem.*, 2017; 65(9): 1977-1983.
- Sari LM, Hakim RF, Mubarak Z, Andriyanto A, Analysis of phenolic compounds and immunomodulatory activity of areca nut extract from Aceh, Indonesia, against *Staphylococcus aureus* infection in Sprague-Dawley rats. *Vet World*, 2020; 13(1): 134-140.
- Vanlalveni C, Lallianrawna S, Biswas A, Selvaraj M, Changmai B, Rokhum SL, Green synthesis of silver nanoparticles using plant extracts and their antimicrobial activities: a review of recent literature. *RSC Adv.*, 2021; 11(5): 2804-2837.
- Das RK, Gogoi N, Bora U, Green synthesis of gold nanoparticles using Nyctanthes arbor-tristis flower extract. *Bioprocess Biosyst Eng.*, 2011; 34(5): 615-619.
- Makarov VV, Love AJ, Sinitsyna OV, Makarova SS, Yaminsky IV, Taliansky ME, Kalinina NO, "Green" nanotechnologies: synthesis of metal nanoparticles using plants. *Acta Naturae*, 2014; 6(1): 35-44.
- Jiang XC, Chen WM, Chen CY, Xiong SX, Yu AB, Role of Temperature in the Growth of Silver Nanoparticles Through a Synergetic Reduction Approach. *Nanoscale Res Lett.*, 2011; 6(1): 32.
- Marciniak L, Nowak M, Trojanowska A, Tylkowski B, Jastrzab R, The Effect of pH on the Size of Silver Nanoparticles Obtained in the Reduction Reaction with Citric and Malic Acids. *Materials*, 2020; 13(23): 5444.
- 14. Singh M, Sinha I, Mandal RK, Role of pH in the green synthesis of silver nanoparticles. *Materials Letters*, 2009; 63(3-4): 425-427.
- Prodjosantoso AK, Prawoko OS, Utomo MP, Sari LP, Green synthesis of silver nanoparticles using *Salacca zalacca* extract as reducing agent and it's antibacterial activity. *Asian J Chem.*, 2019; 31(12): 2804-2810.
- 16. Seifipour R, Nozari M, Pishkar L, Green synthesis of Silver Nanoparticles using Tragopogon Collinus

Leaf Extract and Study of Their Antibacterial Effects. *JIOPM*, 2020; 30(8): 2926-2936.

- Indarti K, Apriani EF, Wibowo AE, Simanjuntak P, Antioxidant Activity of Ethanolic Extract and Various Fractions from Green Tea (*Camellia sinensis* L.) Leaves. *Pharmacognosy Journal*, 2019; 11(4): 771-776.
- Sudirman S, Herpandi H, Safitri E, Apriani EF, Taqwa F, Total polyphenol and flavonoid contents and antioxidant activities of water lettuce (*Pistia* stratiotes) leave extracts. Food Research, 2022; 6: 205-210.
- Apriani EF, Mardiyanto M, Hendrawan A, Optimization of Green Synthesis of Silver Nanoparticles From *Areca Catechu* L. Seed Extract with Variations of Silver Nitrate and Extract Concentrations Using Simplex Lattice Design Method. *Farmacia*, 2022; 70(5): 917-924.
- Priya RS, Geetha D, Ramesh PS, Antioxidant activity of chemically synthesized AgNPs and biosynthesized *Pongamia pinnata* leaf extract mediated AgNPs - A comparative study. *Ecotoxicol Environ Saf.*, 2016; 134(Pt 2): 308-318.
- Ibrahim S, Ahmad Z, Manzoor MZ, Mujahid M, Faheem Z, Adnan A, Optimization for biogenic microbial synthesis of silver nanoparticles through response surface methodology, characterization, their antimicrobial, antioxidant and catalytic potential. *Sci Rep.*, 2021; 11(1): 770.
- Apriani EF, Rosana Y, Iskandarsyah I, Formulation, characterization, and in vitro testing of azelaic acid ethosome-based cream against *Propionibacterium acnes* for the treatment of acne. *J Adv Pharm Technol Res.*, 2019; 10(2): 75-80.
- 23. Apriani EF, Miksusanti M and Fransiska N, Formulation And Optimization Peel-Off Gel Mask with Polyvinyl Alcohol and Gelatin Based Using Factorial Design from Banana Peel Flour (*Musa* paradisiaca L) As Antioxidant. Indonesian Journal of Pharmacy, 2022; 33(2): 261-268.
- Mardiyanto M, Apriani EF, Alfarizi MH, Formulation and *In-vitro* Antibacterial Activity of Gel containing Ethanolic extract of Purple Sweet Potato Leaves (*Ipomoea batatas* (L.) Loaded Poly Lactic Co-Glycolic Acid Submicroparticles against *Staphylococcus aureus*. *Res J Pharm Tech.*, 2022; 15(8): 3599-3595.
- Zhang WM, Li B, Han L, Zhang HD, Antioxidant activities of extracts from areca (*Areca catectu* L.) flower, husk and seed. *Afr J Biotechnol.*, 2009; 8(16): 3887-3892.
- Talabani RF, Hamad SM, Barzinjy AA, Demir U, Biosynthesis of Silver Nanoparticles and Their Applications in Harvesting Sunlight for Solar Thermal Generation. *Nanomaterials*, 2021; 11(9): 2421.
- Ider M, Abderrafi K, Eddahbi A, Ouaskit S, Kassiba A, Silver Metallic Nanoparticles with Surface Plasmon Resonance: Synthesis and Characterizations. *J Clust Sci.*, 2017; 28(3): 1051-1069.
- Usmani A, Mishra A, Jafri A, Arshad M, Siddiqui MA, Green synthesis of silver nanocomposites of *Nigella sativa* seeds extract for hepatocellular carcinoma. *Curr Nanomater.*, 2019; 4(3): 191-200.
- 29. Yeshchenko OA, Dmitruk IM, Alexeenko AA, Kotko AV, Verdal J, Pinchuk AO, Size and

Temperature Effects on the Surface Plasmon Resonance in Silver Nanoparticles. *Plasmonics*, 2012; 7(4): 685-694.

- Alqadi MK, Noqtah OAA, Alzoubi FY, Alzouby J, Aljarrah K, pH effect on the aggregation of silver nanoparticles synthesized by chemical reduction. *Mater Sci-Poland*, 2014; 32(1): 107-111.
- Sathishkumar M, Sneha K, Yun Y, Immobilization of silver nanoparticles synthesized using *Curcuma longa* tuber powder and extract on cotton cloth for bactericidal activity. *Bioresour Technol.*, 2010; 101(20): 7958-7965.
- Anandalakshmi K, Venugobal J, Ramasamy V, Characterization of silver nanoparticles by green synthesis method using *Pedalium murex* leaf extract and their antibacterial activity. *Appl Nanosci.*, 2016; 6: 399-408.
- 33. Roopan SM, Rohit G, Madhumitha A, Rahuman A, Kamaraj C, Bharathi A, Suhendra TV, Low-cost and eco-friendly phyto-synthesis of silver nanoparticles using *Cocos nucifera* coir extract and its larvicidal activity. *Ind Crop Prod.*, 2013; 43(1): 631-635.
- 34. Tagad CK, Dugasani SR, Aiyer R, Park S, Kulkarni A, Sabharwal S, Green synthesis of silver nanoparticles and their application for the development of optical fiber based hydrogen peroxide sensor. *Sens Actuators B Chem.*, 2013; 183: 144-149.

- Zuorro A, Iannone A, Natali S, Lavecchia R, Green Synthesis of Silver Nanoparticles Using Bilberry and Red Currant Waste Extracts. *Processes*, 2019; 7(4): 193.
- Krishnaraj C, Ramachandran R, Mohan K, Kalaichelvan PT, Optimization for rapid synthesis of silver nanoparticles and its effect on phytopathogenic fungi. *Spectrochim Acta A Mol Biomol Spectrosc.*, 2012; 93: 95-99.
- Choi JS, Jung HC, Baek YJ, Kim BY, Lee MW, Kim HD and Kim SW, Antibacterial Activity of Green-Synthesized Silver Nanoparticles Using *Areca catechu* Extract against Antibiotic-Resistant Bacteria. *Nanomaterials*, 2021; 11 (1): 205.
- Bhat PR, Savitri VH, Laxmi PG, Jenitta EP, A Study on the Phytochemical Analysis, Silver Nanoparticle Synthesis and Antibacterial Activity from Seed Extract of *Areca catechu L. IJBcRR*, 2016; 9(1): 1-9.
- Bélteky P, Rónavári A, Igaz N, Szerencsés B, Tóth IY, Pfeiffer I, Kiricsi M, Kónya Z, Silver nanoparticles: aggregation behavior in biorelevant conditions and its impact on biological activity. *Int J Nanomedicine*, 2019; 14: 667-687.
- 40. Pinto VV, Ferreira MJ, Silva R, Santos HA, Silva F and Pereira CM, Long time effect on the stability of silver nanoparticles in aqueous medium: Effect of the synthesis and storage conditions. *Colloids Surf A Physicochem Eng Aspects*, 2010; 364(1-3): 19-25.