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THE ROLE OF TEMPERATURE AND PH IN THE SYNTHESIS OF SILVER 1 NANOPARTICLES USING ARECA CATECHU L. SEED EXTRACT AS BIOREDUCTOR MARDIYANTO MARDIYANTO, ELSA FITRIA APRIANI\*, FITO PRATAMA HEYLKEN 3 4 Department of Pharmacy, Faculty of Mathematics and Natural Sciences, Sriwijaya University, South Sumatra, Indonesia 6 7 \*corresponding author: elsafitria@mipa.unsri.ac.id 9 Abstract 10 11 Silver nanoparticles are prepared using natural bioreductors such as Areca catechu ethanol extract,

which is rich in phenolic content. This study aims to optimize the temperature and pH conditions 12 for synthesizing silver nanoparticles using the response surface methodology (RSM)-central 13 composite design (CCD) method. The phenolic content of the Areca catechu ethanol extract was 14 111.14 ± 3.41 mg CE/g extract. Temperature and pH conditions significantly affect maximum 15 wavelength and absorbance values. The optimum condition was obtained at a temperature of 30°C 16 and a pH of 10.5. Silver nanoparticles at optimum condition had a wavelength of 423 nm, 17 absorbance was 1.148 particle size was 161.7 ± 46.1 nm, PDI was 0.286 ± 0.035, and zeta potential 18 was -16.1 ± 3.7 mV. The stability of silver nanoparticles at the optimum conditions produced is 19 relatively stable, characterized by no significant changes in organoleptic, pH, and wavelength, but 20 the absorbance value has increased. The resulting silver nanoparticles have good characteristics 21 22 and good stability.

- 23 Keywords: Areca catechu L. seed, pH, response surface methodology, silver nanoparticles,
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## 25 Introduction

The use of areca seeds is generally to overcome the decay of body tissue and to preserve animal skin. Cowhide is usually preserved with crushed areca seeds. The results of more detailed research show that areca seeds can inactivate the growth of rotting germs. Nanotechnology in the pharmaceutical field is currently developing very rapidly. Silver nanoparticles are among the most researched and set in the medical world because they have been shown to have anti- bacterial, anti-inflammatory, anti-angiogenesis, anti-fungal, antiviral, and antiplatelet activities [1-4].

The production of silver nanoparticles has shifted towards green synthesis methods that are more environmentally friendly. The green synthesis method requires the help of secondary metabolites in plants as a source of bioreductors [5]. Secondary metabolites that can act as bioreductors usually act as antioxidants, such as flavonoids, phenolics, and alkaloids [6]. Flavonoid, phenolic and alkaloid compounds contain many groups whose electrons require a partner so that they are needed to react organic-chemically way [7, 8].

The formation of silver nanoparticles through the green synthesis method can be influenced by several factors, such as temperature and pH [9]. Temperature is another critical factor affecting the formation of nanoparticles in plant extracts [10]. In general, an increase in temperature will increase nanoparticle synthesis's reaction rate and efficiency [11]. This statement is proven by research conducted by Jiang et al [12], where a temperature of 17 to 55°C will accelerate the reaction and cause the particle size to increase from 90 nm to 180 nm. Besides temperature, changes in pH will also affect the process of forming silver nanoparticles. Changes in pH will result in changes in the natural phytochemical charge contained in the extract. This effect will affect its ability to bind and reduce metal cations while synthesizing silver nanoparticles. According to research by Marciniak et al. [13], the reaction for forming silver nanoparticles runs at a pH of 6.0 to 11.0 for citric acid and 7.0 to 11.0 for malic acid. Increasing the pH can also affect the particle size of silver nanoparticles, where the higher the pH value, the smaller the particle size [14].

Based on the description above, researchers are interested in conducting research in the form of optimizing the effect of temperature and pH on lmax surface plasmon resonance (SPR) and absorbance in the manufacture of silver nanoparticles using ethanol extract of young areca nut seeds as a bioreductor agent. Optimization was carried out using the response surface methodology (RSM) - central composite design (CCD) method for temperature and pH. The temperature and pH used in this study were 30 – 70°C respectively and the pH range was 7.5 – 10.5 referring to the research by Prodjosantoso et al. [15] and Seifipour et al. [16] with slight modifications. The formula of silver nanoparticles with optimum temperature and pH will be further characterized in particle size, polydispersity index (PDI), zeta potential and thermodynamic stability tests using the cycling test method.

 Materials and Methods

Materials

The materials used in this study were Areca catechu L. seed s, AgNO3 (Emsure, Indonesia), 70% ethanol (Bratachem , Indonesia), absolute ethanol (Emsure , Indonesia), NaOH 0.1 M (Bratachem , Indonesia), catechins (Sigma-Aldrich, Singapore), and aqua deionized (Bratachem, Indonesia).

Areca catechu L. seeds were washed, dried and crushed to obtain Simplicia powder. The simplicia

Preparation of Ethanolic Extract of Areca catechu L. Seed

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69 powder was macerated using 70% ethanol solvent and stored in a dark place for 48 hours. The filtrate was then evaporated using a rotary evaporator to obtain a thick extract. Quantification of Total Phenolic Content in Extract The measurement of total phenolic content in the 71 extract was carried out based on the research of Indarti et al. [17] and Sudirman et al. [18] with 72 73 modifications. The extract is put at 10 mg into a 100 mL measuring flask, then the volume is made up to obtain a 100 mg/mL concentration. The absorbance of the sample solution was measured using a UV-Vis spectrophotometer at 281 nm. The formula can calculation the total phenolic concentration (TPC) in extract in Equation 1.

> $TPC = volume (mL) \times concentration (mg/mL) \times dilute factor$ (1) g extract

## Formula of Silver Nanoparticles

The formula for making silver nanoparticles using the ethanol extract of Areca catechu L. refers to research by Apriani et al. [19]. The concentration of silver nitrate (AgNO<sub>3</sub>) used was 3 mM and 3 mL of extract at a concentration of 10% w/v.

# Temperature and pH Optimization

Optimization of temperature and pH in the manufacture of silver nanoparticles was carried out using the response surface methodology (RSM) - central composite design (CCD) method referring to the research of Prodjosantoso et al. [15] and Seifipour et al. [16] with slight modifications. The levels used consist of low, middle, and high levels. The design of the levels for each factor, namely temperature and pH can be seen in Table I. Based on the conditions in Table I, nine formulas were produced for manufacturing silver nanoparticles which can be seen in Table II.

#### Determination of Surface Plasmon Resonance 94 95 Surface plasmon resonance was determined by observing the wavelength and absorbance of the sample solution in the range of 370 - 600 nm using a UV-Vis Spectrophotometer. The blank 96 used in this procedure is distilled water. Determination of Optimum Conditions for Silver 97 Nanoparticles Optimum conditions (temperature and pH) were determined to manufacture silver 98 99 nanoparticles by analyzing surface plasmon resonance data using the Design Expert 138 100 program. The program will examine the effect of temperature and pH factors on surface plasmon 101 resonance's wavelength response and absorbance. The program suggests optimum conditions when these conditions have a desirability value close to 1. 102 103 104 Characterization of Silver Nanoparticles at Optimum Condition 105 Silver nanoparticles under optimum conditions were subjected to further characterization, such as the determination of surface plasmon resonance, particle size, polydispersity index, and zeta 106 potential. Particle size, polydispersity index, and zeta potential were measured using a Particle 107 Size Analyzer. The sample solution is diluted using aqua deionized. Particle size and 108 polydispersity index were measured at a scattering angle of 90°, while measured the zeta potential 109 at a scattering angle of 173° [22]. 110 111 112 Stability Test of Silver Nanoparticles at Optimum Condition A stability test was carried out using the cycling test method. The cycling test was carried out at 113 temperatures 4 °C with storage at each temperature for 24 hour. The procedure was repeated for six cycles 114 [23.24]. Organoleptic observation, pH, and surface plasmon resonance were observed in cycles 0 and 6. 115 116 117 Data Analysis 118 Data analysis for optimization was performed using program Design-Expert 13<sup>®</sup>. 119 120 121 Results and Discussion 122 Areca catechu L. Seeds Extract 123 The Areca catechu L. extract obtained in this study had a distinctive odour, brown colour and thick. The total phenolic content obtained was 111.4 ± 0.411 mg CE/g extract. The results were not 124 much different from those of Zhang et al. [25]. 125

Silver Nanoparticles

Areca catechu L. seed extract is a bioreductor in forming silver nanoparticles. The silver nanoparticle preparations obtained in this study were blackish-brown in colour. Areca catechu L. seed extract can act as a bioreductor due to the presence of phenolic compounds. Phenolics can actively chelate and reduce metal ions into nanoparticles. The transformation of the phenolic tautomer from the enol to the keto form by releasing reactive hydrogen atoms can reduce metal ions to form nanoparticles [11]. An overview of the mechanism for creating silver nanoparticles from catechin compounds can be seen in Figure 1.

The formation of silver nanoparticles can be observed from surface plasmon resonance events. Surface plasmon resonance (SPR) is a free electron resonance oscillation on a metal surface layer that is excited by an incident light source [26]. The oscillation is determined by the absorption of the wavelength obtained [27]. Surface plasmon resonance (SPR) wavelengths are 400 - 450 nm due to the interaction between light and surface electrons moving from silver nanoparticles [28]. In addition, the absorbance produced at surface plasmon resonance wavelengths can describe the colour intensity and the number of silver nanoparticles formed during the synthesis process. The higher the absorbance, the more silver nanoparticles are formed. The surface plasmon resonance results obtained in this study were in the wavelength range of 424 - 431 nm and absorbance of 0.431 - 1.265 (Table III).

The maximum wavelength and absorbance data from the SPR obtained are closely related to each formula's different temperature and temperature conditions. Based on Figure 2, F3 has the highest absorbance of 1.265 while F9 has the lowest wavelength of 423 nm.

The Role of Temperature and pH to \(\lambda\) max of Silver Nanoparticles

The role of temperature and pH on the maximum wavelength of silver nanoparticles was analyzed using the Design Expert 13\* program. The initial step is to analyze the model and then analyze the response. The model analysis obtained at the maximum wavelength response. Model analyzing on lmax has an R<sup>2</sup> value of more than 0.7 namely 0.9161, which shows that 91.61%

of the data on the lmax response is influenced by temperature and pH factors. In comparison, the remaining 8.39% is an estimation error. This result is supported by the normal curve plot of residuals (Figure 3A) which shows that the data points are close to a straight line so that the data can be said to be normally distributed. Furthermore, the adjusted R<sup>2</sup> and predicted R<sup>2</sup> values obtained in the Imax model analyzing have a difference between adjusted R2 and predicted R2 values illustrates the similarity of the model obtained from the relationship between temperature and pH to the lmax response. This result is supported by the predicted vs. actual curve in Figure 3B. The predicted vs. actual curve illustrates the results of the adjusted R2 and predicted R2 values with points to the right and left of the line. The closer the distance between the dots to the right and left of the line indicates that the data values from the research are more precise than those predicted by the system, where 86.58% of the existing data already represents the population and can explain a good linear relationship. The adequate value obtained in the model analysis is more than 4, which is 11.4290. The greater the adequate precision value, the more resistant the model will be to noise (disturbance). In addition, the p-value of less than 0.05 in the model indicates that the model used significantly affects the lmax response. Based on the model analyzing, it is said that the model is good to be continued in the response analyzing process. Analyzing of the Imax response was carried out to observe the effect of each factor and the interaction between the two on the response. Response analysis was carried out by following the results of the ANOVA in the form of coefficient values and p- values of the temperature (A) and pH (B) factors. The results of the ANOVA analysis on the pH response can be seen in Table V. Based on the results of the ANOVA analysis in Table V, temperature and pH significantly affect the lmax response where the p-value obtained is less than 0.05. Factors that have a significant effect are then entered into the response equation so that the response equation lmax is accepted, namely y = 426.667 + 1.833A - 2.167B. The notation for factors A and B is (+) and (-), respectively. This notation shows that the Atemperature factor has a positive correlation, meaning that the greater the A value, the higher the resulting Imax value, In contrast, the B-pH factor has a negative correlation, meaning that the greater the pH, the smaller the resulting Imax value. This result was supported the research of Yeshchenko et al. [29], which showed that an increase in temperature causes a redshift (a shift in wavelength to a more significant value). This condition is due to the thermal volume expansion of the silver nanoparticles with increasing temperature. Thermal volume expansion is when a substance experiences a change in temperature so that the substance can expand (expand) or

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shrink (shrink) depending on the increase or decrease in temperature. Research conducted by Alqadi et al. [30] also supported the effect of pH in this study. The study showed that increasing the pH resulted in smaller lmax and was followed by the formation of smaller silver nanoparticle sizes. According to Sathishkumar et al. [31], when the pH is acidic, the aggregation of silver nanoparticles to form larger nanoparticles is believed to be preferable to the stages of creating new nanoparticles (nucleation). However at alkaline pH, it will show a large number of functional groups available for silver binding, thereby facilitating a higher number of Ag<sup>+</sup> ions to bind and subsequently form a large number of smaller diameter nanoparticles.

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The Role of Temperature and pH to Absorbance of Silver Nanoparticle

The model analysis obtained at the maximum wave-length response can be seen in Table VI. The model analysis results indicate that the model is robust and significant enough to proceed to the response analysis process. Model analysis on the absorbance response has an R<sup>2</sup> value of 0.9700, which indicates that 97% of the data on the absorbance response is influenced by temperature and pH factors. In comparison, the remaining 3% is an estimation error. This result is supported by the normal plot of the residual curve (Figure 4A), where the data points are close to a straight line so that the data can be said to be normally distributed. Furthermore, the adjusted R2 and predicted R2 values obtained in the absorbance model analysis have a difference of less than 0.2 namely 0.0697, so there is a similarity in the model obtained from the relationship between temperature and pH on the absorbance response. This result is supported by the predicted vs. actual curve in Figure 4B, which shows the distance between the points to the right and left of the near line where 95.19% of the existing data already represents the population and can explain a good linear relationship. The adeq precision value obtained in the model analysis is more than 4 namely 18.5757 so the model is robust against noise (disturbance). In addition, the p-value of less than 0.05 in the model indicates that the model used significantly affects the absorbance response. The absorbance response analysis was continued, and the results obtained can be seen in Table VII. Based on the results of the ANOVA analysis in Table VII, temperature and pH have a significant effect on the absorbance response. The absorbance response equation obtained is y =0.930 + 0.094A + 0.346B - 0.146B2. The notation on an equation for factors A and B is (+), but there is a new factor, namely B2, which has a value of (-). Factor B2, marked (-), means that when the pH used is too high, the absorbance produced will be smaller. When viewed from the p-value,

factor B has the smallest p-value, so that factor B has the greatest influence on the absorbance 220 221 222 The temperature parameter is directly proportional to the absorbance value. This shows that when there is an increase in temperature, the absorbance value of the silver nanoparticles produced 223 will increase. Based on research by Prodjosantoso et al. [15], when the temperature increases, the 224 225 formation rate of silver nanoparticles will be faster so that many silver nanoparticles are formed. The more silver nanoparticles formed, the higher the absorbance [32]. 226 227 In addition, the pH parameter is also directly proportional and has the most significant effect on the 228 absorbance value. This shows that when there is an increase in pH, the absorbance value of the silver nanoparticles will be higher. These results are supported by research by Alqadi et al. [30], 229 230 which showed that an increase in pH resulted in a higher absorbance of silver nanoparticles. In addition, Roopan et al. [33] research showed that at pH 2 no reaction to form silver nanoparticles 231 occurred, while at pH 11, the silver nanoparticles formed were highly monodispersed. Research by 232 Tagad et al. [34] also showed that at acidic nH (pH 4 and 6), there is no surface plasmon 233 resonance absorption which is characteristic of silver nanoparticles. The formation of silver 234 nanoparticles is more suitable to occur in alkaline pH conditions so that many small silver 235 nanoparticles are formed, and the resulting absorbance value will be higher [35]. However, the 236 synthesis of silver nanoparticles at extreme alkaline pH (> 12) can produce low stability and 237 238 cause aggregation [36]. 239 Optimum Condition for Silver Nanoparticles 240 241 Determination of optimum conditions is done by looking at the system's desirability value closest to 1. The desirability value close to 1 indicates that the situation meets the target desired 242 243 by the researcher. Based on the analysis results, the optimum conditions for making silver 244 nanoparticles are recommended for the system to be at a temperature of 30°C and pH of 245 10.5 with a desirability value of 0.932. Characterization of Silver Nanoparticles at Optimum

Silver nanoparticles at optimum conditions were tested for characterization, such as surface plasmon resonance, particle size, polydispersity index, and zeta potential. The results of the

characterization can be seen in Table VIII. Silver nanoparticles in the optimum condition have

an excellent \( \lambda \) max value and absorbance for silver nanoparticles. The size of the silver

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nanoparticles obtained was  $161.7 \pm 46.1$  nm. The particles size results obtained were smaller than previous studies conducted by Choi et al. [37] at 233.4 - 238 nm and Bhat et al. [38] at 553 - 610 nm. The polydispersity index value was  $0.286 \pm 0.035$  indicating that the particles size obtained was uniform, The zeta potential value obtained from the optimum formula for silver nanoparticles was  $-16.1 \pm 3.7$  mV. This results indicate that the zeta potential of the silver nanoparticle optimum formula is still relatively stable. The negative zeta potential value is due to the alkaline pH (nH 10.5) during the biosynthesis of silver nanoparticles. With an increase in pH (alkaline pH), the organic functional groups on the surface of silver nanoparticles reach a higher level of deprotonation (release of H<sup>+</sup> protons) so that the amount of negative surface charge increases simultaneously [39]. The characterization results obtained have met the requirements for nanoparticle preparations.

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### Stability of Silver Nanoparticles at Optimum Condition

The silver nanoparticles in optimum condition were tested for stability by cycling for six cycles. The stability results of silver nanoparticles can be seen in Table IX. Organoleptic results showed the formation of a slightly precipitate in the 6th cycle. The results obtained are better than the previous reference research conducted by Apriani et al. [19], which showed many deposits formed on colloidal silver nanoparticles in the 6th cycle. The difference in stability results obtained can be due to differences in the conditions for synthesizing silver nanoparticles. This study used a temperature of 30 °C and pH 10.5 as the optimum condition, while the study of Apriani et al. [19] used a temperature of 60 °C and pH 9. Research by Pinto et al. [40] showed that high temperatures can produce unstable silver nanoparticles and can cause damage or decomposition of plant secondary metabolites, which function as stabilizing agents (capping agents). Changes in the pH value in this study were also said to be relatively stable, where the pH changed from 10.50 ± 0.008 to 10.10 ± 0.008. In contrast to previous research from the same researcher, Apriani et al. [19], with the conditions of nanoparticle synthesis at a temperature of 60 °C and pH 9 the decrease in pH was significant, namely pH 9. This stable pH also affect changes in \( \lambda max \) value and absorbance. The shift in λmax in the 6th cycle was not too far and was still in the range of silver nanoparticles. In contrast, the absorbance in the 6th cycle increased, indicating that more and more silver nanoparticles were formed. This is because phenolic stability depends on pH. Phenolic compounds are in the enol form at acidic pH and are in the keto form at an alkaline

pH. The keto form of phenolic is helpful as a stabilizing agent (capping agent). In this study, the 282 283 reaction to form silver nanoparticles continued because the environmental pH was relatively stable 284 at around 10. 285 286 Conclusions 287 Temperature and pH conditions in manufacturing silver nanoparticles have been shown to affect the 288 surface plasmon resonance characteristics. The temperature has positive effect on λmax and the absorbance of SPR, while pH harms λmax and has positive impact on the absorbance of SPR. 289 The optimum temperature and pH conditions for the synthesis of silver nano- particles using 290 ethanol extract of young areca nut seeds were found at 30 °C and pH 10.5 with a surface plasmon 291 resonance silver nanoparticle λmax value of 423 nm and an absorbance of 1.148. Particle size, 292 293 polydispersity index (PDI), and zeta potential of silver nanoparticles at optimum condition were  $161.7 \pm 46.1$  nm,  $0.286 \pm 0.035$  and  $-16.1 \pm 3.7$  mV respectively. The stability of the silver 294 295 nanoparticles at the optimum conditions produced was relatively stable, characterized by not too many organoleptic changes, pH, which was still stable in the range of 10 small wavelength shifts, and 296 increased absorbance values. 297 298 Acknowledgement 299 This study was supported by PT. DKSH Indonesia (Malvern). 300 301 302 Conflict of interest The authors declare no conflict of interest. 303 304 305 306 References 1. Baharara J, Namvar F, Mousavi M, Ramezani T, Mohamad R, Anti-Angiogenesis Effect of 307 308 Biogenic Silver Nanoparticles Synthesized Using Saliva officinalis on Chick Chorioalantoic 309 Membrane (CAM). Molecules, 2014; 19: 13498-13508. 310 Mallmann EJ, Cunha FA, Castro BN, Maciel AM, Menezes EA, Fechine PB, Antifungal 311 activity of silver nanoparticles obtained by green synthesis. Rev Inst Med Trop Sao Paulo, 2015; 312 57(2): 165-167.

- 3. Tyavambiza C, Elbagory AM, Madiehe AM, Meyer M, Meyer S, The Antimicrobial and Anti-
- 314 Inflammatory Effects of Silver Nanoparticles Synthesised from Cotyledon orbiculata Aqueous
- 315 Extract. Nanomaterials, 2021; 11: 1343.
- 4. Massadeh M, Al-Masri M, Abu-Qatouseh LF, Antibacterial activity of Peganum harmala
- 317 extracts and their green silver nanoparticles against acne associated bacteria. Farmacia, 2022,
- 318 70(5): 954-963.
- 5. Roy A, Bulut O, Some S, Mandal AK, Yilmaz MD, Green synthesis of silver nanoparticles:
- 320 biomolecule- nanoparticle organizations targeting antimicrobial activity. Royal Soc Chem Adv., 2019;
- **321** 9: 2673-2702.
- 6. Wulandari IO, Pebriatin BE, Valiana V, Hadisaputra S, Ananto AD, Sabarudin A, Green
- 323 Synthesis of Silver Nanoparticles Coated by Water Soluble Chitosan and Its Potency as Non-
- 324 Alcoholic Hand Sanitizer Formulation. Materials, 2022; 15: 4641.
- 7. Jain V, Garg A, Parascandola M, Chaturvedi P, Khariwala SS, Stepanov I, Analysis of
- 326 alkaloids in Areca nut-containing products by liquid chromatography- tandem mass spectrometry. J
- 327 Agric Food Chem., 2017; 65(9): 1977-1983.
- 328 8. Sari LM, Hakim RF, Mubarak Z, Andriyanto A, Analysis of phenolic compounds and
- 329 immunomodulatory activity of areca nut extract from Aceh, Indonesia, against Staphylococcus
- 330 aureus infection in Sprague- Dawley rats. Vet World, 2020; 13(1): 134-140.
- 9. Vanlalveni C, Lallianrawna S, Biswas A, Selvaraj M, Changmai B, Rokhum SL, Green
- synthesis of silver nanoparticles using plant extracts and their antimicrobial activities: a review
- of recent literature. RSC Adv., 2021; 11(5): 2804-2837.
- 10. Das RK, Gogoi N, Bora U, Green synthesis of gold nanoparticles using Nyctanthes arbor-
- tristis flower extract. Bioprocess Biosyst Eng., 2011; 34(5): 615-619.
- 11. Makarov VV, Love AJ, Sinitsyna OV, Makarova SS, Yaminsky IV, Taliansky ME, Kalinina
- NO, "Green" nanotechnologies: synthesis of metal nanoparticles using plants. Acta Naturae,
- 338 2014; 6(1): 35-44.
- 339 12. Jiang XC, Chen WM, Chen CY, Xiong SX, Yu AB, Role of Temperature in the Growth of
- 340 Silver Nanoparticles Through a Synergetic Reduction Approach. Nanoscale Res Lett., 2011;
- 341 6(1): 32.

- 342 13. Marciniak L, Nowak M, Trojanowska A, Tylkowski B, Jastrzab R, The Effect of pH on the
- 343 Size of Silver Nanoparticles Obtained in the Reduction Reaction with Citric and Malic Acids.
- 344 Materials, 2020; 13(23): 5444.
- 345 14. Singh M, Sinha I, Mandal RK, Role of pH in the green synthesis of silver nanoparticles.
- 346 Materials Letters, 2009; 63(3-4): 425-427.
- 347 15. Prodjosantoso AK, Prawoko OS, Utomo MP, Sari LP, Green synthesis of silver
- nanoparticles using Salacca zalacca extract as reducing agent and it's antibacterial activity. Asian J
- 349 Chem., 2019; 31(12): 2804-2810.
- 350 16. Seifipour R, Nozari M, Pishkar L, Green synthesis of Silver Nanoparticles using
- 351 Tragopogon Collinus Leaf Extract and Study of Their Antibacterial Effects. JIOPM, 2020;
- 352 30(8): 2926-2936.
- 353 17. Indarti K, Apriani EF, Wibowo AE, Simanjuntak P, Antioxidant Activity of Ethanolic
- 354 Extract and Various Fractions from Green Tea (Camellia sinensis L.) Leaves. Pharmacognosy
- 355 Journal, 2019; 11(4): 771-776.
- 356 18. Sudirman S, Herpandi H, Safitri E, Apriani EF, Taqwa F, Total polyphenol and flavonoid
- 357 contents and antioxidant activities of water lettuce (Pistia stratiotes) leave extracts. Food
- 358 Research, 2022; 6: 205-210.
- 359 19. Apriani EF, Mardiyanto M, Hendrawan A, Optimization of Green Synthesis of Silver
- 360 Nanoparticles From Areca Catechu L. Seed Extract with Variations of Silver Nitrate and
- 361 Extract Concentrations Using Simplex Lattice Design Method. Farmacia, 2022; 70(5): 917-
- 362 924.
- 363 20. Priya RS, Geetha D, Ramesh PS, Antioxidant activity of chemically synthesized AgNPs and
- 364 biosynthesized Pongamia pinnata leaf extract mediated AgNPs A comparative study.
- 365 Ecotoxicol Environ Saf., 2016; 134(Pt 2): 308-318.
- 366 21. Ibrahim S, Ahmad Z, Manzoor MZ, Mujahid M, Faheem Z, Adnan A, Optimization for
- 367 biogenic microbial synthesis of silver nanoparticles through response surface methodology,
- 368 characterization, their antimicrobial, antioxidant and catalytic potential. Sci Rep., 2021; 11(1):
- 369 770.
- 370 22. Apriani EF, Rosana Y, Iskandarsyah I, Formulation, characterization, and in vitro testing
- 371 of azelaic acid ethosome-based cream against Propionibacterium acnes for the treatment of
- 372 acne. J Adv Pharm Technol Res., 2019; 10(2): 75-80.

- 373 23. Apriani EF, Miksusanti M and Fransiska N, Formulation And Optimization Peel-Off Gel
- Mask with Polyvinyl Alcohol and Gelatin Based Using Factorial Design from Banana Peel
- 375 Flour (Musa paradisiaca L) As Antioxidant. Indonesian Journal of Pharmacy, 2022; 33(2):
- 376 261-268.
- 377 24. Mardiyanto M, Apriani EF, Alfarizi MH, Formulation and In-vitro Antibacterial Activity of
- 378 Gel containing Ethanolic extract of Purple Sweet Potato Leaves (Ipomoea batatas (L.) Loaded
- 379 Poly Lactic Co- Glycolic Acid Submicroparticles against Staphylococcus aureus. Res J Pharm
- 380 Tech., 2022; 15(8): 3599-3595.
- 381 25. Zhang WM, Li B, Han L, Zhang HD, Antioxidant activities of extracts from areca (Areca
- 382 catectu L.) flower, husk and seed. Afr J Biotechnol., 2009; 8(16): 3887-3892.
- 383 26. Talabani RF, Hamad SM, Barzinjy AA, Demir U, Biosynthesis of Silver Nanoparticles and
- 384 Their Applications in Harvesting Sunlight for Solar Thermal Generation. Nanomaterials, 2021;
- 385 11(9): 2421.
- 386 27. Ider M, Abderrafi K, Eddahbi A, Ouaskit S, Kassiba A, Silver Metallic Nanoparticles with
- 387 Surface Plasmon Resonance: Synthesis and Characterizations. J Clust Sci., 2017; 28(3): 1051-
- 388 1069.
- 389 28. Usmani A, Mishra A, Jafri A, Arshad M, Siddiqui MA, Green synthesis of silver
- 390 nanocomposites of Nigella sativa seeds extract for hepatocellular carcinoma. Curr
- 391 Nanomater., 2019; 4(3): 191-200.
- Yeshchenko OA, Dmitruk IM, Alexeenko AA, Kotko AV, Verdal J, Pinchuk AO, Size
- 393 and Temperature Effects on the Surface Plasmon Resonance in Silver Nanoparticles. Plasmonics,
- 394 2012; 7(4): 685-694.
- 30. Alqadi MK, Noqtah OAA, Alzoubi FY, Alzouby J, Aljarrah K, pH effect on the aggregation
- of silver nanoparticles synthesized by chemical reduction. Mater Sci-Poland, 2014; 32(1): 107-
- 397 111.
- 398 31. Sathishkumar M, Sneha K, Yun Y, Immobilization of silver nanoparticles synthesized using
- 399 Curcuma longa tuber powder and extract on cotton cloth for bactericidal activity. Bioresour
- 400 Technol., 2010; 101(20): 7958-7965.
- 401 32. Anandalakshmi K, Venugobal J, Ramasamy V, Characterization of silver nanoparticles by
- green synthesis method using Pedalium murex leaf extract and their antibacterial activity. Appl
- 403 Nanosci., 2016; 6: 399-408.

33. Roopan SM, Rohit G, Madhumitha A, Rahuman A, Kamaraj C, Bharathi A, Suhendra TV, 404 405 Low-cost and eco-friendly phyto-synthesis of silver nanoparticles using Cocos nucifera coir 406 extract and its larvicidal activity. Ind Crop Prod., 2013; 43(1): 631-635. 34. Tagad CK, Dugasani SR, Aiyer R, Park S, Kulkarni A, Sabharwal S, Green synthesis of silver 407 nanoparticles and their application for the development of optical fiber based hydrogen peroxide 408 409 sensor. Sens Actuators B Chem., 2013; 183: 144-149. 410 35. Zuorro A, Iannone A, Natali S, Lavecchia R, Green Synthesis of Silver Nanoparticles Using 411 Bilberry and Red Currant Waste Extracts. Processes, 2019; 7(4): 193. 36. Krishnaraj C, Ramachandran R, Mohan K, Kalaichelvan PT, Optimization for rapid synthesis 412 413 of silver nanoparticles and its effect on phytopathogenic fungi. Spectrochim Acta A Mol Biomol 414 Spectrosc., 2012; 93: 95-99. 415 37. Choi JS, Jung HC, Baek YJ, Kim BY, Lee MW, Kim HD and Kim SW, Antibacterial Activity of Green-Synthesized Silver Nanoparticles Using Areca catechu Extract against Antibiotic-416 Resistant Bacteria. Nanomaterials, 2021; 11 (1): 205. 417 38. Bhat PR, Savitri VH, Laxmi PG, Jenitta EP, A Study on the Phytochemical Analysis, Silver 418 Nanoparticle Synthesis and Antibacterial Activity from Seed Extract of Areca catechu L. IJBcRR, 419 2016; 9(1): 1-9. 420 421 39. Belicky P. Ronavari A Igaz N. Szerences B. Toch IY. Pfeifer I. Kirisci M, Konya Z. Silver nanoparticles: aggregation behavior in biorelevant conditions and its impact on biological 422 activity. Int J Nanomedicine, 2019; 14: 667-687. 423 40. Pinto VV, Ferreira MJ, Silva R, Santos HA, Silva F and Pereira CM, Long time effect on 424 425 the stability of silver nanoparticles in aqueous medium: Effect of the synthesis and storage conditions. Colloids Surf A Physicochem Eng Aspects, 2010; 364(1-3): 19-25. 426 427 428 429 430 431 432 433 434

Table I Level design for temperature and pH

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1	-	Level		
	Factor	Low	Medium	Hig

		Low	Medium	
	Temperature	30	50	Г
	pH	7.5	9	Г
)				

Table II
Formula and condition of silver nanoparticles

4	4	4	
4	4	5	

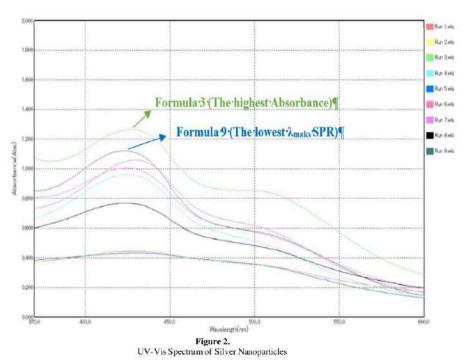
Formula	AgNO <sub>3</sub> 3 mM (mL)	Extract 10% b/v (mL)	Temp (°C)	pН
1	27	3	70	7.5
2	27	3	30	9
3	27	3	70	10.5
4	27	3	50	9
5	27	3	50	7.5
6	27	3	70	9
7	27	3	30	10.5
8	27	3	30	7.5
9	27	3	50	10.5

APPENDIX

Figure 1. Silver nanoparticle formation mechanism

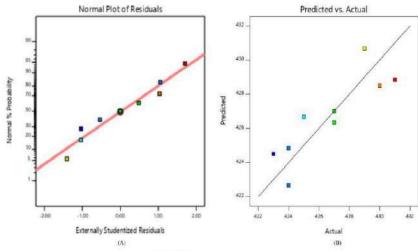
 $\begin{tabular}{ll} \textbf{Table III} \\ \textbf{Data maximum wavelength and absorbance of silver nanoparticles} \\ \end{tabular}$ 

Formula	Temperature (°C)	pН	Max. Wavelength (nm)	Absorbance
1	70	7.5	429	0.446
2	30	9	424	0.769
3	70	10.5	427	1.265
4	50	9	425	0.962
5	50	7.5	431	0.431
6	70	9	430	1.059
7	30	10.5	424	1.004
8	30	7.5	427	0.435
9	50	10.5	423	1.121



476 Table IV
 477 Model Analyzing of λmax Responses

Response	5		Paramet	er	
261130 #41101 GHZ	$\mathbb{R}^2$	Adjusted R <sup>2</sup>	Predicted R <sup>2</sup>	Adequate precision	p-value
Max. Wavelength	0.9161	0.8658	0.6980	11.4290	0.0192



 $\label{eq:Figure 3.} \textbf{Normal Plot of residual (A) and predicted $\it ws.$ actual (B) curves of $\lambda$max}$ 

Table V
Response analyzing of λmax

Responses	Intercept (nm)	A	В
λmaks SPR	426.667	1.833	-2.167
p-values		0.0397*	0.0213*

A: Temperature; B: pH; \*factors that have a significant effect (p-value < 0.05)

**Table VI**Model analysis of absorbance responses

Response	5		Paramete	er	
	R <sup>2</sup>	Adjusted R <sup>2</sup>	Predicted R <sup>2</sup>	Adequate precision	p-value
Absorbance	0.9700	0.9519	0.8822	18,5757	0.0003

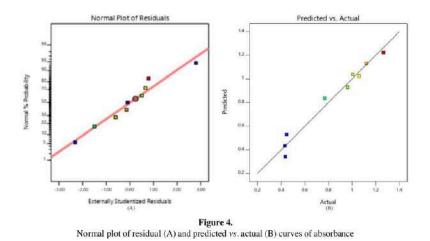


Table VII
Response analysis of absorbance

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Response	Intercept (nm)	A	В	<b>B</b> <sup>2</sup>
Absorbance	0.930	0.094	0.346	-0.146
p-values		0.0232*	0.0001*	0.0333*

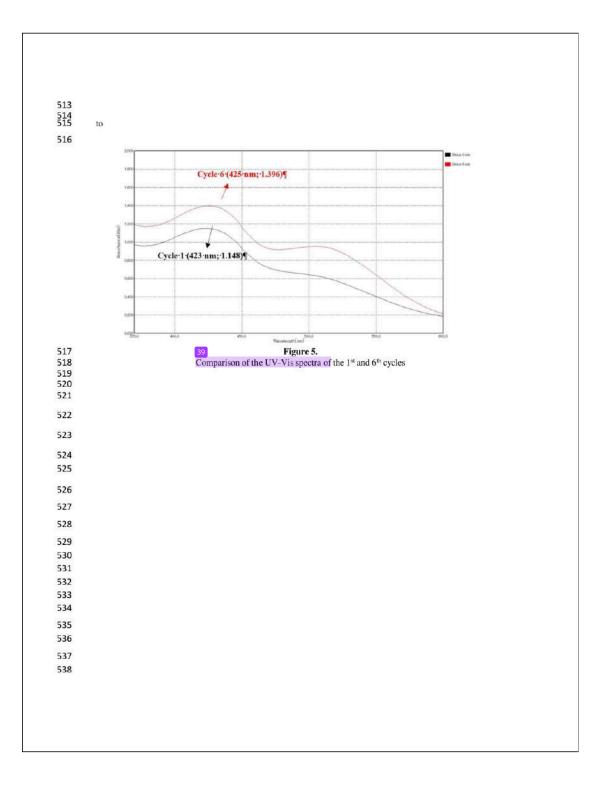
A: Temperature; B: pH; "factors that have a significant effect (p-value < 0.05

Table VIII Characterization results

Parameter	Mean ± SD
λmax (nm)	$423 \pm 2.156$
Absorbance particle	$1.148 \pm 0.076$
Size (nm)	$161.7 \pm 46.1$
PDI (index)	$0.286 \pm 0.035$
Zeta potential (mV)	$-16.1 \pm 3.7$

Table IX The Stability Results

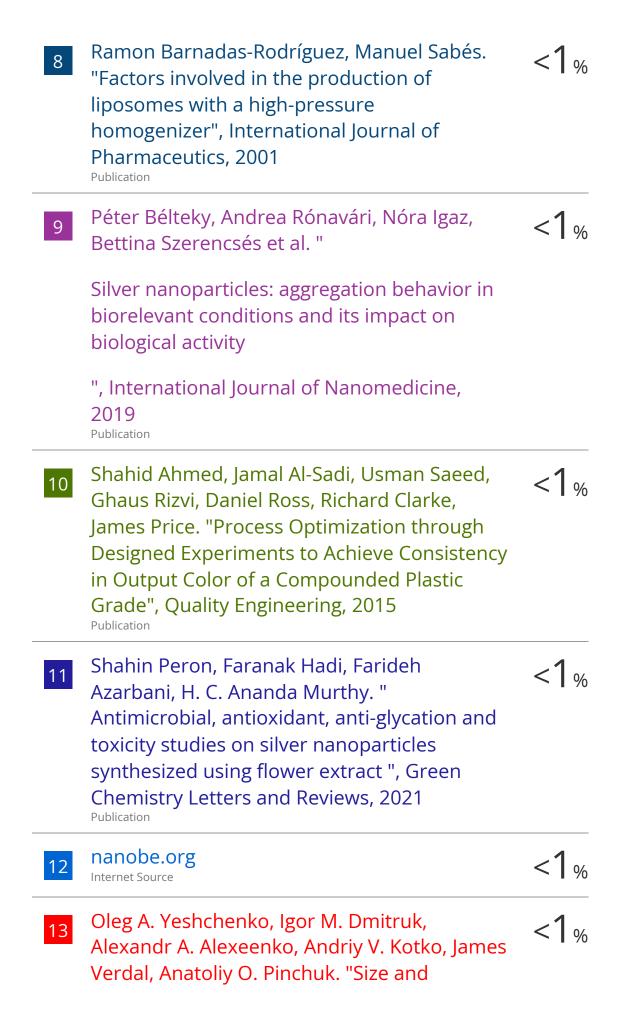
Cycle	Sedimentation	pН	λmax (nm)	Absorbance
1	None	$10.50 \pm 0.008$	423	1.148
6	Slight	$10.10 \pm 0.008$	425	1.396



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