

DISERTASI

**EKSPLORASI JAMUR ENDOFITIK TANAMAN CIPLUKAN
(*Physalis angulata* L.) YANG BERPOTENSI SEBAGAI
SUMBER ANTIOKSIDAN DAN ANTIBAKTERI**

**Diajukan untuk memenuhi salah satu syarat memperoleh gelar
Doktor pada Program Studi Doktor Ilmu MIPA Fakultas MIPA
Universitas Sriwijaya**



BUDI EKO WAHYUDI

NIM 08013622328006

**PROGRAM STUDI DOKTOR ILMU MIPA
FAKULTAS MATEMATIKA DAN ILMU PENGETAHUAN ALAM
UNIVERSITAS SRIWIJAYA**

2025

HALAMAN PENGESAHAN

EKSPLORASI JAMUR ENDOFITIK TANAMAN CIPLUKAN (*Physalis angulata* L.) YANG BERPOTENSI SEBAGAI SUMBER ANTIOKSIDAN DAN ANTIBAKTERI

DISERTASI

**Diajukan untuk Melengkapi Salah Satu Syarat
Memperoleh Gelar Doktor Bidang Ilmu MIPA**

**OLEH
BUDI EKO WAHYUDI
NIM. 08013622328006**

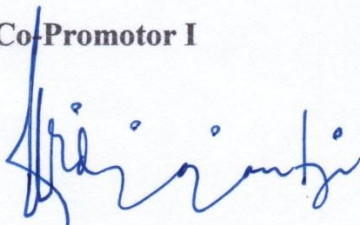
Palembang, 15 September 2025

Promotor



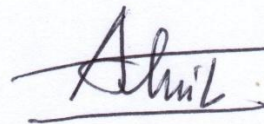
**Prof. Dr. Elfita, M.Si.
NIP. 196903261994122001**

Co-Promotor I



**Prof. Dr. Hary Widjajanti, M.Si.
NIP. 196112121987102001**

Co-Promotor II



**Prof. Dr. Salni, M.Si.
NIP. 196608231993031002**

**Mengetahui,
Dekan Fakultas Matematika dan Ilmu Pengetahuan Alam**



**Prof. Hormansyah, S.Si., M.Si., Ph.D.
NIP. 197111191997021001**

HALAMAN PERSETUJUAN

Karya tulis ilmiah berupa Disertasi ini dengan judul “Eksplorasi Jamur Endofitik Tanaman Ciplukan (*Physalis angulata* L.) yang Berpotensi Sebagai Sumber Antioksidan dan Antibakteri” telah dipertahankan di hadapan Tim Penguji Karya Ilmiah Fakultas Matematika dan Ilmu Pengetahuan Alam Program Studi Doktor Ilmu Matematika dan Ilmu Pengetahuan Alam Universitas Sriwijaya pada tanggal 15 September 2025

Palembang, 15 September 2025

Tim Penguji Karya Tulis Ilmiah berupa Disertasi

Ketua:

Prof. Dr. Elfita, M.Si.

NIP. 196903261994122001

Anggota:

1. Prof. Dr. Hary Widjajanti, M.Si.

NIP. 196112121987102001

2. Prof. Dr. Salni, M.Si.

NIP. 196608231993031002

3. Prof. Dr. Andria Agusta

NIP. 196908161994031003

4. Dr. rer. nat. Mardiyanto, M.Si.

NIP. 197103101998021002

5. Prof. Dr. Arum Setiawan, S.Si., M.Si., C.EIA

NIP. 197211221998031001

Mengetahui,

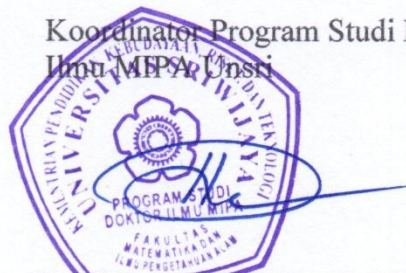
Dekan Fakultas MIPA Unsri



Prof. Hermansyah, S.Si., M.Si., Ph.D

NIP. 197111191997021001

Koordinator Program Studi Doktor
Ilmu MIPA Unsri



Prof. Drs. Dedi Rohendi, M.T., Ph.D.

NIP. 196704191993031001

HALAMAN PERNYATAAN INTEGRITAS

Yang bertanda tangan di bawah ini:

Nama : Budi Eko Wahyudi

NIM : 08013622328006

Judul : Eksplorasi Jamur Endofitik Tanaman Ciplukan (*Physalis angulata* L.) yang Berpotensi sebagai Sumber Antioksidan dan Antibakteri

Menyatakan bahwa Disertasi saya merupakan hasil karya sendiri didampingi tim Promotor dan Co-Promotor dan bukan hasil penjiplakan/plagiat. Apabila ditemukan unsur penjiplakan/plagiat dalam Disertasi ini, maka saya bersedia menerima sanksi akademik dari Universitas Sriwijaya sesuai aturan yang berlaku.

Demikian pernyataan ini saya buat dalam keadaan sadar dan tanpa ada paksaan dari siapapun.

Palembang, 15 September 2025



Budi Eko Wahyudi

NIM. 08013622328006

HALAMAN PERNYATAAN PERSETUJUAN PUBLIKASI

Yang bertanda tangan di bawah ini:

Nama : Budi Eko Wahyudi
NIM : 08013622328006
Tempat, Tanggal Lahir : Muba, 03 Juni 1988
Judul : Eksplorasi Jamur Endofitik Tanaman
Ciplukan (*Physalis angulata* L.) yang
Berpotensi sebagai Sumber Antioksidan
dan Antibakteri

Memberikan izin kepada Promotor dan Co-Promotor dan Universitas Sriwijaya untuk mempublikasikan hasil penelitian saya untuk kepentingan institusi apabila dalam 1 (satu) tahun tidak mempublikasikan karya penelitian saya. Dalam kasus ini saya setuju untuk menempatkan Pembimbing sebagai penulis korespondensi (Corresponding author).

Demikianlah pernyataan ini saya buat dalam keadaan sadar dan tanpa ada paksaan dari siapapun.

Palembang, 15 September 2025



Budi Eko Wahyudi

NIM. 08013622328006

HALAMAN PERSEMBAHAN

ومن لم يذق مر التعلم ساعة, تجرع ذل الجهل طول حياته

“Barang siapa yang tidak merasakan pahitnya belajar sesaat, maka dia akan merasakan hinanya kebodohan seumur hidupnya”

(Muhammad bin Idris Asy-Syafi’i)

Disertasi ini ku persembahkan untuk:

Orangtua dan Mertua yang telah mendo’akan dalam setiap langkah
Istri dan anakku yang selalu memberikan semangat, mendo’akan dan mengiringi
setiap perjuangan

Adikku yang selalu memberikan dukungan

Sahabat-sahabatku yang selalu setia membantu dan memberi dukungan

KATA PENGANTAR

Alhamdulillah, segala puji bagi Allah SWT atas berkat rahmat dan karunia-Nya, penulisan Disertasi yang berjudul “Eksplorasi Jamur Endofitik Tanaman Ciplukan (*Physalis angulata* L.) yang Berpotensi sebagai Sumber Antioksidan dan Antibakteri” dapat diselesaikan.

Pada kesempatan ini, penulis mengucapkan terima kasih yang sebesar-besarnya kepada Prof. Dr. Elfita, M.Si. sebagai Promotor, Prof. Dr. Hary Widjajanti, M.Si. sebagai Co-Promotor I dan Prof. Dr. Salni, M.Si. sebagai Co-Promotor II yang telah banyak meluangkan waktu untuk memberikan bimbingan selama penulisan disertasi ini. Penulis juga mengucapkan terima kasih kepada:

1. Prof. Dr. Taufiq Marwa, S.E., M.Si. selaku Rektor Universitas Sriwijaya
2. Prof. Hermansyah, S.Si., M.Si., Ph.D. selaku Dekan Fakultas Matematika dan Ilmu Pengetahuan Alam Universitas Sriwijaya
3. Prof. Drs. Dedi Rohendi, M.T., Ph.D., selaku Koordinator Program Studi Doktor Ilmu Matematika dan Ilmu Pengetahuan Alam Fakultas MIPA Universitas Sriwijaya.
4. Seluruh Dosen dan Staf Administrasi Program Studi Doktor Ilmu Matematika dan Ilmu Pengetahuan Alam Universitas Sriwijaya.
5. Universitas Sriwijaya yang telah membiayai kuliah melalui Beasiswa Tenaga Kependidikan dan penelitian melalui Skema Hibah Penelitian Fundamental dan Hibah Profesi
6. Teladanku dalam pendidikan dan pekerjaan, Ibu Dra. Lucia Maria Santoso, M.Si. dan Bapak Dr. Riyanto, S.Pd., M.Si., serta seluruh dosen Program Studi Pendidikan Biologi FKIP Unsri
7. Orang tuaku (Edi Mulyono dan Siti Nur Badriyah), mertuaku (Heni Setiawati), istriku (Yulvi Restalita), anakku (Azam Khalid Kencana), adikku (Dwi Angga Kusuma) dan sahabatku (Rian Oktiansyah, Noviyanto, Sakinah, Daniel Alfarado, Faried Arba'i) yang sudah banyak membantu, mendo'akan dan mendukung sehingga pendidikan doktor ini bisa diselesaikan.

Dalam Disertasi ini, penulis menyadari masih banyak kekurangan dan keterbatasan ilmu. Maka dari itu, penulis menerima saran dan kritik yang membangun untuk perbaikan di masa mendatang. Penulis berharap, Disertasi ini dapat diterima dan bermanfaat bagi masyarakat.

Palembang, 15 September 2025
Penulis

RINGKASAN

EKSPLORASI JAMUR ENDOFITIK TANAMAN CIPLUKAN (*Physalis angulata* L.) YANG BERPOTENSI SEBAGAI SUMBER ANTIOKSIDAN DAN ANTIBAKTERI

Karya Tulis Ilmiah berupa Disertasi, September 2025

Budi Eko Wahyudi, Dibimbing oleh Prof. Dr. Elfita, M.Si., Prof. Dr. Hary Widjajanti, M.Si., Prof. Dr. Salni, M.Si.

Program Studi Doktor Ilmu MIPA, Fakultas Matematika dan Ilmu Pengetahuan Alam, Universitas Sriwijaya

viii, 252 halaman, 70 gambar, 29 tabel, 4 lampiran

Tanaman ciplukan (*Physalis angulata* L.) telah dimanfaatkan oleh masyarakat Indonesia dan dunia sebagai obat tradisional untuk mengatasi diabetes, gusi berdarah, hipertensi, bisul, atau luka infeksi, karena efek farmakologi yang kuat sebagai antioksidan, antibakteri, hepatoprotektor, antifibrotik dan antidiare. Senyawa bioaktif tanaman ciplukan dapat diperoleh melalui jamur endofitik dengan keuntungan budidaya yang lebih singkat dan kemampuan jamur dalam menghasilkan metabolit sekunder yang mirip atau lebih baik dari inangnya. Penelitian ini bertujuan untuk menganalisis karakter fenotip dan mengidentifikasi jamur endofitik, menganalisis aktivitas antioksidan dan antibakteri ekstrak jamur endofitik, menganalisis karakterisasi dan menentukan struktur senyawa murni serta bioaktivitas dari senyawa murni tersebut. Isolasi, kultivasi dan ekstraksi jamur endofitik dilakukan untuk mengidentifikasi jenis jamur endofitik dan memperoleh ekstrak jamur endofitik. Jamur endofitik terpilih diidentifikasi secara molekuler, kemudian dianalisis menggunakan BLAST NCBI dan disusun pohon filogenetiknya menggunakan BioEdit 2.2, MEGA 11 dan iTOL. Uji aktivitas antioksidan menggunakan metode DPPH untuk ekstrak jamur endofitik dan senyawa murni. Uji aktivitas antibakteri menggunakan metode difusi Kirby-Bauer untuk ekstrak jamur endofitik dan senyawa murni. Senyawa murni di analisis menggunakan spektroskopi NMR dan LCMS/MS, dan penentuan struktur menggunakan ChemDraw. Berdasarkan hasil penelitian, diperoleh 47 jamur endofitik yang diisolasi dari akar (CA1-CA10), batang (CB1-CB5), daun (CD1-CD9), tangkai buah (CT1-CT6), kelopak buah (EP1-EP8) dan perikarp buah (CH1-CH2, BP1-BP7) yang terbagi menjadi 21 genera yaitu *Aspergillus*, *Bispora*, *Cladorrhinum*, *Cylindrocarpon*, *Diaporthe*, *Fusarium*, *Helicocephalum*, *Hyalodendron*, *Lasiodiplodia*, *Mortierella*, *Mucor*, *Nigrospora*, *Paecilomyces*, *Papulaspora*, *Penicillium*, *Periconia*, *Phialophora*, *Pythium*, *Trichocladium*, *Trichoderma* dan *Verticillium*. Isolasi senyawa murni dari ekstrak jamur endofitik menggunakan kromatografi kolom pada 6 isolat dengan aktivitas antioksidan dan antibakteri yang potensial. Karakterisasi senyawa menggunakan spektroskopi ¹H-NMR, ¹³C-NMR, HSQC, HMBC dan LCMS/MS. Diperoleh senyawa murni 10-hydroxy-benzoisochromen-1-one yang berasal dari isolat CA3 dan CB1. Senyawa

7-hydroxy-benzochromen-6-one dari isolat CA3, CA7 dan CB1. Senyawa *4,5,6-trihydroxy-2',3',5',6-tetramethyl-[3,4'-bipyranylidene]-2-one* dari isolat CB5 dan CH1. Senyawa *2-hydroxy-5-((6-methylheptyl)oxy)phenyl)(phenyl)methanone* dari isolat CD1. Senyawa tersebut diuji aktivitas antioksidan, secara berurutan menunjukkan IC₅₀, dimulai dari senyawa *10-hydroxy-benzoisochromen-1-one* (IC₅₀ 262,68 dan 264,09 µg/mL), *7-hydroxy-benzochromen-6-one* (IC₅₀ 259,11 µg/mL, 256,13 µg/mL dan 258,20 µg/mL), *4,5,6-trihydroxy-2',3',5',6-tetramethyl-[3,4'-bipyranylidene]-2-one* (IC₅₀ 23,82 µg/mL dan 24,40 µg/mL), dan senyawa murni *2-hydroxy-5-((6-methylheptyl)oxy)phenyl)(phenyl)methanone* (IC₅₀ 137,07 µg/mL). Uji aktivitas antibakteri senyawa murni *10-hydroxy-benzoisochromen-1-one*, *7-hydroxy-benzochromen-6-one* dan *4,5,6-trihydroxy-2',3',5',6-tetramethyl-[3,4'-bipyranylidene]-2-one*, menunjukkan nilai KHM ≤64 µg/mL (kuat) pada semua bakteri uji. Selanjutnya senyawa murni *2-hydroxy-5-((6-methylheptyl)oxy)phenyl)(phenyl)methanone*, menunjukkan nilai KHM 125 µg/mL (sedang) pada bakteri *S. typhi* dan 250 µg/mL (lemah) pada bakteri uji lainnya. Berdasarkan struktur penyusunnya, semua senyawa murni tersebut merupakan golongan senyawa fenolat, kecuali senyawa dari isolat CB5 dan CH1 yang merupakan golongan bipyranylidene. Uji molekuler dari jamur endofit dengan aktivitas antioksidan dan antibakteri potensial terhadap 8 isolat, teridentifikasi spesies *Trichoderma virens* (CA3), *Papulaspora equi* (CA9), *Paecilomyces variotii* (CB5), *Lasiodiplodia theobromae* (CD1), *Fusarium equiseti* (CD8), *Diaporthe eucalyptorum* (CT5), *Penicillium citrinum* (EP6) dan *Penicillium chermesinum* (BP2). Hasil penelitian ini menunjukkan bahwa jamur endofitik yang diisolasi dari berbagai organ tanaman ciplukan memiliki potensi untuk dikembangkan menjadi antioksidan dan antibakteri alami.

Kata kunci : antibakteri, antioksidan, jamur endofitik, *Physalis angulata* L.

Kepustakaan : 500 (2019-2025)

SUMMARY

EXPLORATION OF ENDOPHYTIC FUNGI FROM CIPLUKAN (*Physalis angulata* L.) WITH POTENTIAL AS A SOURCE OF ANTIOXIDANTS AND ANTIBACTERIAL AGENTS

Doctoral Dissertation, September 2025

Budi Eko Wahyudi; Supervised by Prof. Dr. Elfita, M.Si., Prof. Dr. Hary Widjajanti, M.Si., Prof. Dr. Salni, M.Si.

Doctoral Program in Natural Sciences, Faculty of Mathematics and Natural Sciences, Sriwijaya University

viii, 250 pages, 70 figures, 29 tables, 4 appendices

Physalis angulata L. (commonly known as ciplukan) has been traditionally utilized in Indonesia and several other countries as a medicinal plant for the treatment of diabetes, bleeding gums, hypertension, boils, and infected wounds, owing to its strong pharmacological properties as an antioxidant, antibacterial, hepatoprotective, antifibrotic, and antidiarrheal agent. The bioactive compounds of *P. angulata* can also be obtained from endophytic fungi, which provide advantages such as shorter cultivation periods and the ability to produce secondary metabolites similar to, or even more potent than, those of the host plant. This study aimed to analyze the phenotypic characteristics and identify endophytic fungi, evaluate the antioxidant and antibacterial activities of their extracts, characterize and elucidate the structure of purified compounds, and assess their bioactivities. Isolation, cultivation, and extraction of endophytic fungi were conducted to identify fungal species and obtain crude extracts. Selected endophytic fungi were identified molecularly, analyzed using NCBI BLAST, and their phylogenetic relationships reconstructed with BioEdit 2.2, MEGA 11, and iTOL. Antioxidant activity was evaluated using the DPPH method, while antibacterial activity was tested by the Kirby–Bauer diffusion method, for both crude extracts and purified compounds. Purified compounds were analyzed by NMR spectroscopy and LCMS/MS, and their structures determined using ChemDraw. A total of 47 endophytic fungi were isolated from different organs of *P. angulata*, including roots (CA1–CA10), stems (CB1–CB5), leaves (CD1–CD9), fruit stalks (CT1–CT6), calyces (EP1–EP8), and pericarps (CH1–CH2, BP1–BP7). These isolates were assigned to 21 genera: *Aspergillus*, *Bispora*, *Cladorrhinum*, *Cylindrocarpon*, *Diaporthe*, *Fusarium*, *Helicocephalum*, *Hyalodendron*, *Lasiodiplodia*, *Mortierella*, *Mucor*, *Nigrospora*, *Paecilomyces*, *Papulaspora*, *Penicillium*, *Periconia*, *Phialophora*, *Pythium*, *Trichocladium*, *Trichoderma*, and *Verticillium*. Purification of bioactive compounds from six isolates with promising antioxidant and antibacterial potential was performed using column chromatography. Structural characterization was carried out by ¹H-NMR, ¹³C-NMR, HSQC, HMBC and LCMS/MS analysis. The purified compounds obtained were 10-hydroxy-benzoisochromen-1-one from isolates CA3 and CB1. 7-hydroxy-benzochromen-6-one from isolates CA3, CA7,

and CB1. 4,5,6-trihydroxy-2',3',5',6-tetramethyl-[3,4'-bipyranylidene]-2-one from isolates CB5 and CH1. 2-hydroxy-5-((6-methylheptyl)oxy)phenyl(phenyl)methanone from isolate CD1. The antioxidant activities of the isolated compounds were evaluated based on their IC₅₀ values. The results, in order, were as follows: 10-hydroxy-benzoisochromen-1-one (IC₅₀ 262.68 and 264.09 µg/mL), 7-hydroxy-benzochromen-6-one (IC₅₀ 259.11, 256.13, and 258.20 µg/mL), 4,5,6-trihydroxy-2',3',5',6-tetramethyl-[3,4'-bipyranylidene]-2-one (IC₅₀ 23.82 and 24.40 µg/mL), and 2-hydroxy-5-((6-methylheptyl)oxy)phenyl(phenyl)methanone (IC₅₀ 137.07 µg/mL). Antibacterial assays showed that 10-hydroxy-benzoisochromen-1-one, 7-hydroxy-benzochromen-6-one, and 4,5,6-trihydroxy-2',3',5',6-tetramethyl-[3,4'-bipyranylidene]-2-one exhibited strong inhibitory effects against all tested bacteria, with minimum inhibitory concentration (MIC) values ≤64 µg/mL. In contrast, 2-hydroxy-5-((6-methylheptyl)oxy)phenyl(phenyl)methanone displayed moderate antibacterial activity against *S. typhi* (MIC = 125 µg/mL) and weak activity against the other tested bacteria (MIC = 250 µg/mL). Structurally, all purified compounds belonged to the phenolic class, except those isolated from CB5 and CH1, which were classified as bipyranylidene derivatives. Molecular identification of eight bioactive isolates revealed them as *Trichoderma virens* (CA3), *Papulaspora equi* (CA9), *Paecilomyces variotii* (CB5), *Lasiodiplodia theobromae* (CD1), *Fusarium equiseti* (CD8), *Diaporthe eucalyptorum* (CT5), *Penicillium citrinum* (EP6), and *Penicillium chermesinum* (BP2). The findings of this study demonstrate that endophytic fungi isolated from different organs of *P. angulata* have significant potential to be developed as natural sources of antioxidant and antibacterial agents.

Keywords : antibacterial, antioxidant, endophytic fungi, *Physalis angulata* L.
Citation : 500 (2019-2025)

DAFTAR ISI

	Halaman
HALAMAN PENGESAHAN.....	ii
HALAMAN PERSETUJUAN.....	iii
HALAMAN PERNYATAAN INTEGRITAS.....	iv
HALAMAN PERNYATAAN PERSETUJUAN PUBLIKASI.....	v
HALAMAN PERSEMBAHAN.....	vi
KATA PENGANTAR.....	vii
RINGKASAN	viii
SUMMARY	x
DAFTAR ISI.....	xii
DAFTAR GAMBAR.....	xiv
DAFTAR TABEL.....	xvi
BAB 1 PENDAHULUAN	18
1.1 Latar Belakang	18
1.2 Rumusan Masalah.....	22
1.3 Hipotesis	23
1.4 Tujuan Penelitian	23
1.5 Kebaruan Penelitian	24
1.6 Manfaat Penelitian	25
1.7 Kerangka Pikir Penelitian	25
1.8 State of The Art.....	26
BAB 2 TINJAUAN PUSTAKA.....	28
2.1 Tanaman Ciplukan	28
2.2 Khasiat Tanaman Ciplukan.....	30
2.3 Metabolit Sekunder	32
2.4 Jamur endofitik	37
2.5 Radikal Bebas	44
2.6 Antioksidan	45
2.7 Bakteri Patogen.....	46
2.8 Antibakteri	52
BAB 3 METODE PENELITIAN	54
3.1 Waktu dan Tempat Penelitian	54
3.2 Alat dan Bahan.....	55
3.3 Prosedur Penelitian	55
3.4 Analisis Data	67
3.5 Penyajian Data	67
3.6 Alur Penelitian	68
BAB 4 HASIL DAN PEMBAHASAN	69
4.1 Isolasi Jamur Endofitik dari Tanaman Ciplukan.....	69
4.2 Keanekaragaman Jamur Endofitik Tanaman Ciplukan	89
4.3 Uji Aktivitas Antioksidan dan Antibakteri Ekstrak Etil Asetat Jamur Endofitik yang Diisolasi dari Tanaman Ciplukan.....	92
4.4 Aktivitas Antioksidan dan Antibakteri Ekstrak Jamur Endofit Terpilih yang Diisolasi dari Tanaman Ciplukan dengan Metode Kromatografi Lapis Tipis (KLT) dan Bioautografi	96
4.5 Isolasi dan Identifikasi Senyawa Metabolit Sekunder dari Isolat CA3.....	100

4.6 Isolasi dan Identifikasi Senyawa Metabolit Sekunder dari Isolat CA7	111
4.7 Isolasi dan Identifikasi Senyawa Metabolit Sekunder dari Isolat CB1	115
4.8 Isolasi dan Identifikasi Senyawa Metabolit Sekunder dari Isolat CB5	122
4.9 Isolasi dan Identifikasi Senyawa Metabolit Sekunder dari Isolat CD1	129
4.10 Isolasi dan Identifikasi Senyawa Metabolit Sekunder dari Isolat CH1	137
4.11 Analisis LC-MS/MS dari Isolat CT5	140
4.12 Analisis LC-MS/MS dari Isolat EP4.....	142
4.13 Perbandingan Senyawa yang Dihasilkan oleh Inang dan Jamur Endofitik	144
4.14 Identifikasi Molekuler Jamur Endofit Terpilih Sebagai Penghasil Senyawa Bioaktif dan Aktivitas Terbaik	147
4.15 Uji Aktivitas Antioksidan dan Antibakteri Senyawa Murni.....	167
4.16 Analisis Komponen Utama	180
BAB 5 SIMPULAN DAN SARAN.....	188
5.1 Simpulan	188
5.2 Saran	189
DAFTAR PUSTAKA.....	190
LAMPIRAN.....	244

DAFTAR GAMBAR

	Halaman
Gambar 2.1 Tanaman ciplukan	28
Gambar 2.2 Struktur kimia berbagai jenis physalin.....	31
Gambar 2.3 Skema biosintesis metabolit sekunder	33
Gambar 2.4 Struktur kimia terpen.....	35
Gambar 2.5 Struktur beberapa senyawa fenolik	37
Gambar 2.6 Struktur 3- hydroxy -4-(hydroxy (4-hydroxyphenyl) methyl) dihydrofuran-2-on	43
Gambar 2.7 Bakteri <i>S. aureus</i>	47
Gambar 2.8 Bakteri <i>B. subtilis</i>	49
Gambar 2.9 Bakteri <i>E. coli</i>	50
Gambar 2.10 Bakteri <i>S. typhi</i>	51
Gambar 3.1 Alur penelitian.....	68
Gambar 4.1 Jamur endofitik yang tumbuh dari berbagai organ tanaman ciplukan.....	69
Gambar 4.2 Penampakan koloni dan konidia jamur endofitik dari akar ciplukan.....	71
Gambar 4.3 Penampakan koloni dan konidia jamur endofitik dari batang ciplukan.....	74
Gambar 4.4 Penampakan koloni dan konidia jamur endofitik dari daun ciplukan.....	77
Gambar 4.5 Penampakan koloni dan konidia jamur endofitik dari tangkai buah ciplukan	80
Gambar 4.6 Penampakan koloni dan konidia jamur endofitik dari kelopak buah ciplukan	84
Gambar 4.7 Penampakan koloni dan konidia jamur endofitik dari perikarp buah ciplukan	87
Gambar 4.8 Kromatogram ekstrak jamur endofitik terpilih.....	97
Gambar 4.9 Bioautografi ekstrak jamur endofitik	99
Gambar 4.10 Kromatogram hasil kolom CA3	100
Gambar 4.11 Kromatogram senyawa murni dari CA3	101
Gambar 4.12 Senyawa 1 dan 2 dari isolat CA3	101
Gambar 4.13 Spektrum ¹ H-NMR (A) and ¹³ C-NMR (B) senyawa 1	102
Gambar 4.14 Spektrum HSQC senyawa 1	103
Gambar 4.15 Spektrum HMBC senyawa 1.....	103
Gambar 4.16 Struktur senyawa 1	106
Gambar 4.17 Spektrum ¹ H-NMR (A) and ¹³ C-NMR (B) senyawa 2	107
Gambar 4.18 Spektrum HSQC senyawa 2.....	108
Gambar 4.19 Spektrum HMBC senyawa 2.....	109
Gambar 4.20 Struktur senyawa 2	111
Gambar 4.21 Kromatogram hasil kolom CA7	112
Gambar 4.22 Kromatogram senyawa murni dari CA7	112
Gambar 4.23 Senyawa murni dari isolat CA7	112
Gambar 4.24 Spektrum ¹ H-NMR (A) and ¹³ C-NMR (B) senyawa 3	114
Gambar 4.25 Struktur senyawa 3	115
Gambar 4.26 Kromatogram hasil kolom CB1	116

Gambar 4.27 Kromatogram senyawa murni dari CB1.....	116
Gambar 4.28 Senyawa 4 dan 5 dari isolat CB1	117
Gambar 4.29 Spektrum ¹ H-NMR (A) dan ¹³ C-NMR (B) senyawa 4	118
Gambar 4.30 Struktur senyawa 4	119
Gambar 4.31 Spektrum ¹ H-NMR (A) dan ¹³ C-NMR (B) senyawa 5	120
Gambar 4.32 Struktur senyawa 5	121
Gambar 4.33 Kromatogram hasil kolom CB5	122
Gambar 4.34 Kromatogram senyawa murni dari CB5.....	123
Gambar 4.35 Senyawa 6 dari isolat CB5	123
Gambar 4.36 Spektrum ¹ H-NMR senyawa 6	123
Gambar 4.37 Spektrum ¹³ C-NMR senyawa 6	124
Gambar 4.38 Spektrum HSQC senyawa 6.....	126
Gambar 4.39 Spektrum HMBC senyawa 6.....	127
Gambar 4.40 Struktur molekul senyawa 6.....	129
Gambar 4.41 Kromatogram hasil kolom CD1	129
Gambar 4.42 Kromatogram senyawa murni dari CD1	130
Gambar 4.43 Senyawa 7 dari isolat CD1	130
Gambar 4.44 Spektrum ¹ H-NMR senyawa 7	130
Gambar 4.45 Spektrum ¹³ C-NMR senyawa 7	131
Gambar 4.46 Spektrum HSQC senyawa 7.....	133
Gambar 4.47 Spektrum HMBC senyawa 7.....	134
Gambar 4.48 Struktur kimia senyawa 7.....	135
Gambar 4.49 Kromatogram hasil kolom CH1	136
Gambar 4.50 Kromatogram senyawa murni dari CH1	137
Gambar 4.51 Senyawa 8 dari isolat CH1	137
Gambar 4.52 Spektrum ¹ H-NMR senyawa 8	137
Gambar 4.53 Spektrum ¹³ C-NMR senyawa 8	138
Gambar 4.54 Struktur molekul senyawa 8.....	139
Gambar 4.55 Kromatogram LC-MS/MS dari isolat CT5	141
Gambar 4.56 Kromatogram LC-MS/MS dari isolat EP4.....	142
Gambar 4.57 Pohon filogenetik jamur endofitik.....	148
Gambar 4.58 Struktur flavonoid	173
Gambar 4.59 Reaksi senyawa flavonoid.....	173
Gambar 4.60 Mekanisme khelasi logam oleh senyawa fenolik.....	175
Gambar 4.61 Keterkaitan organ/bagian tanaman inang dengan keberadaan jamur endofitik dan bioaktivitasnya.....	181

DAFTAR TABEL

	Halaman
Tabel 2.1 Persen aktivitas antibakteri jamur endofitik dari daun <i>P. canescens</i>	422
Tabel 2.2 Aktivitas antibakteri jamur endofitik dari daun <i>S. suffruticosa</i>	433
Tabel 4.1 Karakteristik makroskopik dan mikroskopik isolat jamur endofitik yang diisolasi dari akar ciplukan.....	71
Tabel 4.2 Karakter kunci genus jamur endofit yang diisolasi dari akar ciplukan	72
Tabel 4.3 Karakteristik makroskopik dan mikroskopik isolat jamur endofitik yang diisolasi dari batang ciplukan	75
Tabel 4.4 Karakter kunci genus jamur endofit yang diisolasi dari batang ciplukan	75
Tabel 4.5 Karakteristik makroskopik dan mikroskopik isolat jamur endofitik yang diisolasi dari daun ciplukan	78
Tabel 4.6 Karakter kunci genus jamur endofit yang diisolasi dari daun ciplukan	78
Tabel 4.7 Karakteristik makroskopik dan mikroskopik isolat jamur endofitik yang diisolasi dari tangkai buah ciplukan.....	81
Tabel 4.8 Karakter kunci genus jamur endofit yang diisolasi dari tangkai buah ciplukan.....	81
Tabel 4.9 Karakteristik makroskopik dan mikroskopik isolat jamur endofitik dari kelopak buah ciplukan	82
Tabel 4.10 Karakter kunci genus jamur endofit yang diisolasi dari kelopak buah ciplukan.....	84
Tabel 4.11 Karakteristik makroskopik dan mikroskopik isolat jamur endofitik yang diisolasi dari perikarp buah ciplukan.....	87
Tabel 4.12 Karakter kunci genus jamur endofit yang diisolasi dari perikarp buah ciplukan.....	88
Tabel 4.13 Variasi genus, jumlah individu dan indeks keanekaragaman jamur endofit tanaman ciplukan	900
Tabel 4.14 Aktivitas antibakteri dan antioksidan tanaman inang ciplukan	92
Tabel 4.15 Aktivitas antibakteri dan antioksidan jamur endofitik yang diisolasi dari tanaman ciplukan	93
Tabel 4.16 Uji aktivitas antioksidan dan identifikasi tentatif senyawa dari ekstrak jamur endofitik terpilih	97
Tabel 4.17 Data spektrum NMR 1D dan 2D senyawa 1.....	106
Tabel 4.18 Data spektrum NMR 1D dan 2D senyawa 2.....	111
Tabel 4.19 Data spektrum NMR 1D dan 2D senyawa 3.....	115
Tabel 4.20 Data spektrum NMR 1D senyawa 4	121
Tabel 4.21 Data spektrum NMR 1D senyawa 5	1213
Tabel 4.22 Data spektrum NMR 1D dan 2D senyawa 6.....	130
Tabel 4.23 Data spektroskopi NMR 1D dan 2D senyawa 7	137
Tabel 4.24 Data spektrum NMR 1D senyawa 8	141
Tabel 4.25 Senyawa yang diprediksi pada kromatogram LCMS/MS isolat CT5.....	140
Tabel 4.26 Senyawa yang diprediksi pada kromatogram LCMS/MS isolat	

EP4	143
Tabel 4.27 Perbandingan senyawa yang dihasilkan oleh inang dan jamur endofitik.....	144
Tabel 4.28 Persen kesamaan jamur endofitik	149
Tabel 4.29 Bioaktivitas senyawa murni	168

BAB 1

PENDAHULUAN

1.1 Latar Belakang

Penyakit degeneratif dan infeksi bakteri termasuk jenis penyakit yang umum diderita oleh masyarakat, baik di Indonesia maupun di dunia, serta sering menjadi fokus kajian ilmiah dalam bidang kedokteran dan farmasi. Penyakit degeneratif seperti *diabetes mellitus* (DM) dan infeksi bakteri seperti diare merupakan penyakit dengan prevalensi kematian tinggi di dunia. Data GBD (*Global Burden of Disease Study*) tahun 2021 menunjukkan, kematian akibat DM pada semua umur meningkat dari 238.100 kasus menjadi 723.700 kasus dari seluruh negara, atau naik 203,9% sejak tahun 1990. Kemudian, data jumlah DALY (*Disability Adjusted Life Years*) terkait DM pada semua umur juga meningkat dari 10,4 juta menjadi 39,3 juta, atau meningkat 276,7% dari tahun 1990. Kasus tersebut diperkirakan dapat mencapai 1,3 juta kasus kematian dan 85,5 juta untuk jumlah DALY pada semua umur di tahun 2045. Prevalensi global memperkirakan pada tahun 2030, kasus kematian mencapai 7.079 kasus dari setiap 100 ribu penduduk dunia (Huang *et al.*, 2025; Khan *et al.*, 2020). Data GBD tahun 2019 menunjukkan kematian akibat infeksi bakteri pathogen mencapai 7,7 juta dari total 13,7 juta kasus kematian dunia. Dari keseluruhan data, sebanyak 54,9% kasus infeksi disebabkan oleh bakteri jenis *Staphylococcus aureus*, *Escherichia coli*, *Streptococcus pneumoniae*, *Klebsiella pneumoniae* dan *Pseudomonas aeruginosa*. Selebihnya disebabkan oleh bakteri jenis lain (GBD, 2022; Vaart *et al.*, 2022). Penyakit DM, disebabkan oleh resistensi insulin (T2DM) atau kerusakan sel beta pankreas (T1DM). Resistensi insulin atau apoptosis sel β -pankreas dapat diinduksi oleh stress oksidatif akibat tingginya konsentrasi radikal bebas di dalam tubuh (Masenga *et al.*, 2023; Yavuz *et al.*, 2024). Radikal bebas dinetralisir dengan antioksidan untuk mengurangi stres oksidatif, menurunkan kerusakan sel dan menginisiasi regenerasi sel yang rusak (Chaudhary *et al.*, 2023). Kemudian, penyakit diare terjadi akibat infeksi bakteri di dalam tubuh melalui saluran cerna atau luka, ketika imun tubuh mengalami penurunan akibat jumlah atau fungsi neutrofil terlalu rendah (Nemeth & Pflieger, 2025). Bakteri penyebab diare seperti *Escherichia coli*, *Shigella*

dysenteriae atau *Salmonella typhi* dapat ditemukan pada air tercemar atau makanan mentah (Teshome *et al.*, 2019). Untuk mengobati penyakit akibat radikal bebas dan infeksi bakteri diperlukan antioksidan dan antibakteri.

Antioksidan meredam radikal bebas dengan cara mendonorkan atom H dari gugus hidroksilnya (Moussa & Al-Mamary, 2021). Antioksidan sangat diperlukan tubuh, baik untuk pencegahan penyakit maupun pengobatan. Semakin beragam senyawa antioksidan, maka semakin banyak jenis radikal bebas yang dapat dinetralkan (Zehiroglu & Ozturk Sarikaya, 2019). Selanjutnya, antibakteri membunuh bakteri dengan cara membentuk ikatan dengan membran maupun merusak sel bakteri dengan mengikat DNA polymerase. Senyawa antibakteri, yang dalam penggunaan sehari-hari dikenal sebagai antibiotik, suatu metabolit yang dihasilkan oleh mikroorganisme, yang dapat membunuh atau menghambat pertumbuhan mikroorganisme lain (Byrne *et al.*, 2019). Antioksidan sintetis dan antibiotik tersedia banyak dalam berbagai produk yang sudah terstandar.

Penggunaan antioksidan sintetis dan antibiotik memberikan efek yang cepat dan konsisten. Namun, penggunaan yang tidak sesuai dosis dan dalam jangka waktu yang lama, berpotensi menimbulkan efek samping yang membahayakan. Antioksidan *Butylated Hydroxytoluene* (BHT) memiliki efek toksik pada sel Leydig (penghasil testosterone) tikus melalui induksi disregulasi kalsium dan disfungsi retikulum endoplasma-mitokondria (Ham *et al.*, 2020). Selanjutnya penggunaan antibiotik secara terus menerus menyebabkan bakteri resisten, seperti bakteri *Staphylococcus aureus* yang resisten terhadap antibiotik penicillin dengan tingkat resistensi 93,1% (Gizaw *et al.*, 2023). Bakteri *E. coli* resisten terhadap antibiotik sulfonamide dan trimethoprim (Mulder *et al.*, 2019). Bakteri *S. typhi* resisten terhadap antibiotik *sulfamethoxazole* dengan daya resistensi diatas 50% (Rahman, 2019). Bakteri *B. subtilis* juga resisten terhadap chloramphenicol (95,2%), erythromycin (85,7%) dan gentamicin (42,9%) (X. Jin *et al.*, 2024). Beberapa fakta tersebut mengindikasikan pentingnya eksplorasi senyawa antioksidan dan antibakteri dari bahan alam yang lebih aman dengan efek samping yang relatif kecil. Bahan alam dapat berasal dari akar, rimpang, batang, kulit batang, daun, bunga, buah atau biji. Diantara bahan alam yang berpotensi, tanaman ciplukan telah dikenal sebagai obat alami.

Tanaman ciplukan dimanfaatkan sebagai obat tradisional untuk mengatasi diabetes, gusi berdarah, hipertensi, bisul, tukak lambung, demam, atau mengobati penyakit yang terkait dengan sistem saraf pusat dan sindrom metabolik (Vargas-Arana *et al.*, 2025; Fadhilla *et al.*, 2020). Metabolit sekunder yang terdapat pada tanaman ciplukan diantaranya alkaloid, physalin, withanolides, saponin, steroid, flavonoid dan triterpenoid (Wang *et al.*, 2023a; Odusina & Onocha, 2022). Berdasarkan penelitian, tanaman ciplukan memiliki efek farmakologi seperti antioksidan, antibakteri, antiradang, hepatoprotektor, antifibrotik dan antidiare (Pillai *et al.*, 2022; Lin *et al.*, 2020; Liu *et al.*, 2022). Tanaman obat lain yang masih satu familia (Solanaceae) memiliki potensi mirip dengan ciplukan diantaranya *Capsicum spp.*, *Solanum nigrum*, *Datura metel*, *Nicotiana tabacum*, *Withania somnifera* dan *Lycium barbarum*. Beberapa tanaman ini memiliki aktivitas antioksidan yang kuat terhadap DPPH dan antibakteri terhadap *S. aureus*, *E. coli*, *Salmonella*, *Helicobacter pylori* dan *P. aeruginosa* (Zhang *et al.*, 2025; Zhang *et al.*, 2024b; Islam *et al.*, 2023; Mikulska *et al.*, 2023; Shailabi *et al.*, 2025; Periferakis *et al.*, 2023; Romero-Luna *et al.*, 2023). Keunggulan tanaman ciplukan dibanding beberapa tanaman tersebut karena komposisi bioaktif yang kuat. Ciplukan mengandung *physalins*, *withanolides*, *polifenol* dan *flavonoid*, kombinasi yang memberikan mekanisme ganda: pemerangkap radikal bebas (antioksidan) dan gangguan dinding/virulensi bakteri (antibakteri) (Vargas-Arana *et al.*, 2025; Liang *et al.*, 2024). Fakta tersebut mendukung tanaman ciplukan untuk dipilih sebagai sampel pada penelitian ini.

Perkembangan ilmu pengetahuan dan teknologi menunjukkan metabolit sekunder yang dihasilkan tanaman ciplukan dapat diperoleh melalui jamur endofitik dalam tanaman. Jamur endofitik dapat ditemukan pada akar batang, daun, bunga dan buah yang menghabiskan setengah atau seluruh siklus hidupnya di dalam jaringan tumbuhan yang sehat, baik intraseluler maupun interseluler. Jamur endofitik memiliki kemampuan untuk meniru metabolit sekunder dari inang dan menunjukkan aktivitas farmakologis yang mirip (Wen *et al.*, 2022; Alam *et al.*, 2021). Jamur endofitik yang diisolasi dari tanaman ciplukan diantaranya *Penicillium sp.*, *Hyphomycetes sp.* dan *Fusarium sp.* Jamur *Penicillium sp.* memiliki aktivitas antibakteri dengan diameter hambat sebesar 7,6 mm pada *S.*

aureus dan 6,4 mm pada *E. coli*. Jamur *Hyphomycetes* sp., menunjukkan aktivitas antioksidan paling baik dengan nilai IC_{50} 52,43 $\mu\text{g/mL}$ (Mahardhika *et al.*, 2021). Aktivitas antibakteri dari kedua jamur tersebut terhadap bakteri *S. aureus* dan *E. coli*, memiliki nilai KHM 8 - 64 $\mu\text{g/mL}$ (Palupi *et al.*, 2021). Penelitian tersebut menunjukkan jamur endofitik pada tanaman ciplukan berkhasiat sebagai antioksidan dan antibakteri, mirip seperti inangnya.

Berdasarkan keadaan global tentang bahaya penyakit degeneratif dan infeksi bakteri yang dapat menyebabkan kematian serta efek samping obat sintetik, diperlukan solusi dari bahan alam seperti tanaman ciplukan dengan potensi kuat sebagai obat alami. Khasiat obat alami tanaman ini dapat diperoleh melalui jamur endofitik. Selanjutnya karena kemampuan jamur endofitik dalam meniru metabolit sekunder inang, waktu kultivasi yang singkat dan aktivitasnya yang kuat, perlu adanya eksplorasi lebih lanjut terhadap jamur endofit yang ada pada tanaman ciplukan. Eksplorasi ini bertujuan untuk mengidentifikasi jamur endofit dari akar, batang, daun dan buah tanaman ciplukan, menguji bioaktivitas ekstrak jamur endofitik dan mengisolasi senyawa murni dari jamur tersebut dengan aktivitas paling kuat.

Penelitian ini dimulai dengan mengisolasi jamur endofitik dari organ akar, batang, daun dan buah ciplukan. Isolat jamur dipisahkan dari jamur lain untuk memperoleh isolat jamur murni yang akan digunakan pada proses karakterisasi dan kultivasi. Hasil karakterisasi akan digunakan untuk proses identifikasi fenotip, sementara untuk identifikasi molekuler akan dilakukan terhadap jamur endofitik dengan bioaktivitas paling kuat. Jamur endofitik murni dikultivasi, dipartisi dengan etil asetat kemudian diuapkan hingga diperoleh ekstrak pekat. Ekstrak yang diperoleh akan digunakan untuk uji aktivitas antioksidan dan antibakteri secara invitro. Ekstrak jamur endofitik dengan bioaktivitas paling kuat akan diisolasi senyawa murninya menggunakan kromatografi kolom gravitasi. Senyawa murni yang diperoleh akan digunakan pada uji antioksidan untuk memperoleh IC_{50} dan uji antibakteri untuk memperoleh konsentrasi hambat minimum (KHM).

Eksplorasi jamur endofitik yang akan dilakukan pada penelitian ini bertujuan untuk mengisolasi senyawa murni berkhasiat yang dihasilkan oleh jamur endofitik, karena secara umum penelitian yang dilakukan terhadap jamur endofitik

dari tanaman ini terbatas pada bioaktivitas ekstrak jamur endofitik, bukan pada senyawa murni. Selanjutnya, senyawa murni tersebut akan dibandingkan dengan senyawa yang dihasilkan oleh inang, untuk mengetahui kemiripan struktur atau bioaktivitasnya. Senyawa murni yang dihasilkan sangat diperlukan dalam pembuatan antioksidan dan antibakteri untuk mengatasi penyakit akibat radikal bebas dan infeksi bakteri. Senyawa murni ini dapat memberikan pilihan bahan obat yang berkhasiat untuk dikembangkan pada industri farmasi.

Penelitian ini diharapkan dapat memberikan pengetahuan kepada masyarakat, akademisi, peneliti maupun praktisi kesehatan tentang jamur endofitik potensial dari tanaman obat ciplukan dan aktivitas farmakologinya. Diharapkan informasi ini mampu membangkitkan kreativitas masyarakat dalam memanfaatkan jamur endofitik secara lebih sederhana tanpa harus mengisolasi senyawa murninya, sehingga manfaat jamur endofit dari tanaman ciplukan dapat dirasakan oleh semua lapisan masyarakat.

1.2 Rumusan Masalah

Penyakit degeneratif dan infeksi bakteri memiliki tingkat bahaya tinggi hingga kematian. Untuk mengatasi kedua penyakit tersebut digunakan obat sintetik dengan efek yang cepat, terukur dan konsisten. Namun karena penggunaan yang tidak disiplin, timbul efek samping yang justru berbahaya. Selain itu, perubahan struktur sel pada mikroorganisme karena proses adaptasi menimbulkan resistensi obat. Keadaan ini mendorong para ilmuwan atau praktisi kesehatan untuk menemukan agen antioksidan atau antibakteri baru dari bahan alam yang sudah dikenal berkhasiat sebagai obat seperti tanaman ciplukan. Efek penyembuhan dari bahan alam relatif lebih lambat, namun efek samping atau potensi resistensi dari bahan alam relatif lebih kecil. Khasiat tanaman ciplukan tidak selalu menggunakan tanaman inang, melainkan dapat diperoleh melalui jamur endofitik. Diduga jamur endofitik memiliki bioaktivitas yang lebih kuat dibandingkan inang. Pemilihan jamur endofitik juga dikarenakan kultivasi yang singkat dan tidak memerlukan tempat yang luas. Pemanfaatan ekstrak jamur endofitik sebagai obat akan lebih kuat bioaktivitasnya menggunakan senyawa murni yang terbukti efektif sebagai antioksidan dan antibakteri. Senyawa murni diisolasi dari ekstrak jamur endofitik setelah kultivasi. Kultivasi jamur endofitik hanya menggunakan kultur murni jamur

yang sudah diidentifikasi fenotip. Karena itu, diperlukan adanya eksplorasi dan identifikasi fenotip jamur endofitik yang tumbuh dan berkembang di dalam organ tanaman ciplukan. Berdasarkan alasan tersebut, rumusan masalah dalam penelitian ini sebagai berikut.

1. Bagaimana keanekaragaman jamur endofitik yang diisolasi dari akar, batang, daun dan buah tanaman ciplukan yang dianalisis berdasarkan karakter fenotipiknya?
2. Apakah ekstrak jamur endofitik yang diisolasi dari tanaman ciplukan menunjukkan aktivitas antioksidan dan antibakteri?
3. Apakah ekstrak dan senyawa murni dari jamur endofitik yang diisolasi dari tanaman ciplukan menunjukkan aktivitas atau struktur yang mirip dengan inangnya?

1.3 Hipotesis

- H1 : Jamur endofitik yang ditemukan pada tanaman ciplukan menunjukkan keanekaragaman yang tinggi berdasarkan karakter fenotipnya
- H2 : Ekstrak jamur endofitik yang diisolasi dari tanaman ciplukan menunjukkan aktivitas antioksidan dan antibakteri
- H3 : Ekstrak dan senyawa murni dari jamur endofitik tanaman ciplukan menunjukkan aktivitas dan struktur yang mirip dengan inangnya

1.4 Tujuan Penelitian

Tujuan penelitian ini adalah:

1. Menganalisis karakter fenotip dan mengidentifikasi jenis jamur endofitik dari akar, batang, daun dan buah tanaman ciplukan untuk menentukan keanekaragaman jamur endofitik
2. Menganalisis aktivitas antioksidan dan antibakteri ekstrak jamur endofitik yang diisolasi dari tanaman ciplukan
3. Menganalisis karakterisasi dan menentukan struktur senyawa murni dari jamur endofitik, membandingkan struktur senyawa murni yang berasal dari jamur endofitik dengan inang, dan menganalisis aktivitas antioksidan dan antibakteri senyawa murni dari jamur endofitik

1.5 Kebaruan Penelitian

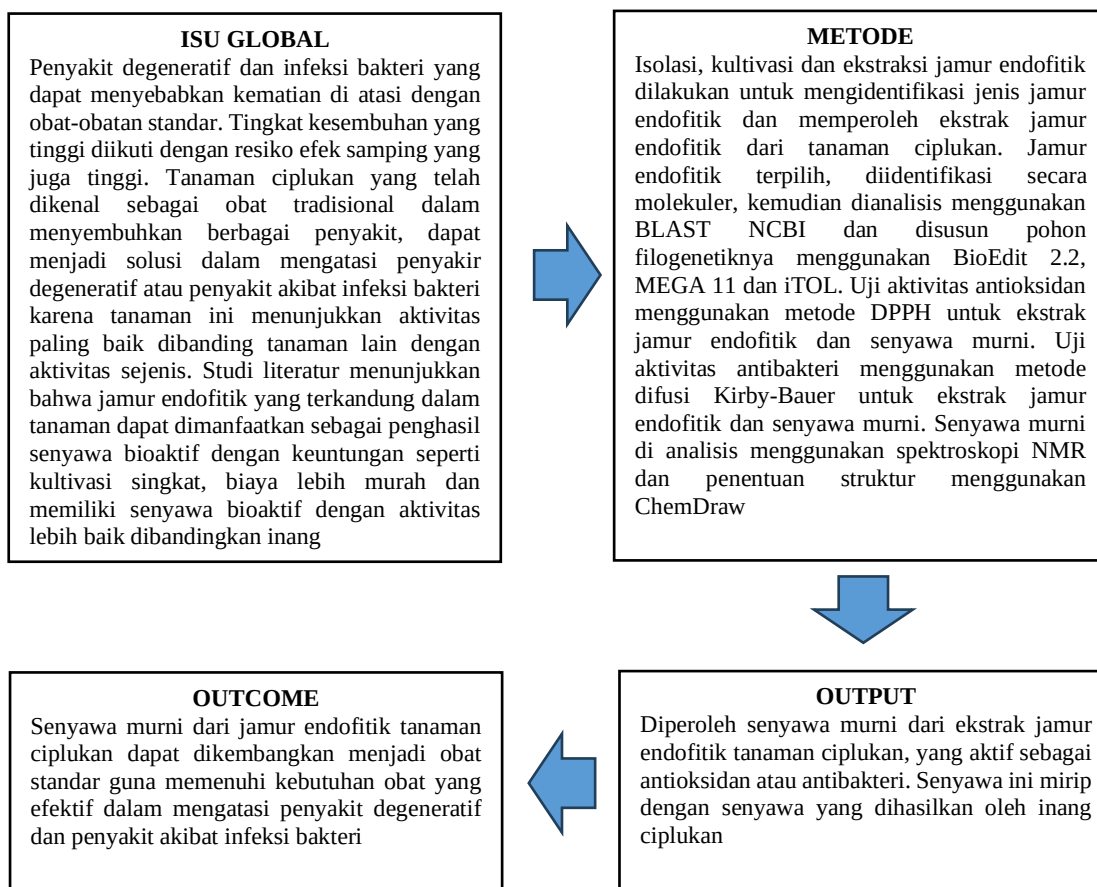
Jamur endofitik yang berhasil diisolasi dari akar, batang, daun dan buah (tangkai, kelopak, perikarp) tanaman ciplukan, diidentifikasi berdasarkan karakter fenotip. Genera jamur endofitik ini belum pernah ditemukan sebelumnya pada tanaman ciplukan yaitu *Bispora*, *Cladorrhinum*, *Cylindrocarpon*, *Diaporthe*, *Helicocephalum*, *Lasiodiplodia*, *Mortierella*, *Mucor*, *Nigrospora*, *Paecilomyces*, *Papulaspora*, *Periconia*, *Phialophora*, *Pythium*, *Trichocladium*, dan *Verticillium*. Analisis molekuler terhadap jamur endofitik terpilih menunjukkan spesies yang baru ditemukan pada ciplukan yaitu *Trichoderma virens* (CA3/akar), *Papulaspora equi* (CA9/akar), *Paecilomyces variotii* (CB5/batang), *Lasiodiplodia theobromae* (CD1/daun), *Fusarium equiseti* (CD8/daun), *Diaporthe eucalyptorum* (CT5/tangkai), *Penicillium citrinum* (EP6/kelopak) dan *Penicillium chermesinum* (BP2/perikarp). Jamur endofitik menunjukkan aktivitas antioksidan tertinggi pada isolat CT5 (tangkai buah) dengan nilai IC_{50} 14,11 $\mu\text{g/mL}$ dan antibakteri tertinggi pada isolat CA3 (akar) dengan presentase hambat 73,91%-96,15%. Aktivitas ini lebih kuat dari penelitian terdahulu (IC_{50} 52,43 $\mu\text{g/mL}$ dan diameter hambat 7,6 mm). **Kebaruan** pada penelitian ini adalah senyawa murni yang berhasil diisolasi menunjukkan struktur yang berbeda dari inang. Senyawa tersebut antara lain 10-hydroxy-benzoisochromen-1-one dihasilkan oleh isolat CA3 (*Trichoderma virens*) dan CB1 (*Papulaspora*). Senyawa 7-hydroxy-benzochromen-6-one dihasilkan oleh isolat CA3 (*Trichoderma virens*), isolat CA7 (*Verticillium*) dan CB1 (*Papulaspora*). Senyawa 4,5,6-trihydroxy- 2',3',5',6 -tetramethyl- [3,4'-bipyranlydene]-2-one dihasilkan oleh isolat CB5 (*Paecilomyces variotii*) dan CH1 (*Fusarium*). Senyawa 2-hydroxy-5- ((6-methylheptyl) oxy) phenyl) (phenyl) methanone dihasilkan oleh isolat CD1 (*Lasiodiplodia theobromae*). Uji aktivitas antioksidan senyawa murni ditunjukkan dengan nilai IC_{50} yaitu 10-hydroxy-benzoisochromen-1-one (IC_{50} 262,68 dan 264,09 $\mu\text{g/mL}$) 7-hydroxy-benzochromen-6-one (IC_{50} 259,11 $\mu\text{g/mL}$, 256,13 $\mu\text{g/mL}$ dan 258,20 $\mu\text{g/mL}$), 4,5,6-trihydroxy- 2',3',5',6-tetramethyl-[3,4'-bipyranlydene]-2-one (IC_{50} 23,82 $\mu\text{g/mL}$ dan 24,40 $\mu\text{g/mL}$), dan senyawa murni 2-hydroxy-5-((6-methylheptyl) oxy)phenyl) (phenyl) methanone (IC_{50} 137,07 $\mu\text{g/mL}$). Uji aktivitas antibakteri senyawa murni dengan nilai KHM ≤ 64 $\mu\text{g/mL}$ (kuat) pada senyawa 10-hydroxy-

benzoisochromen-1-one, 7-hydroxy-benzochromen -6-one dan 4,5,6- trihydroxy -2',3',5',6 -tetramethyl -[3,4'-bipyranylidene]-2-one. Selanjutnya nilai KHM ≥ 65 $\mu\text{g/mL}$ (sedang dan lemah) pada senyawa *2-hydroxy-5-((6-methylheptyl)oxy)phenyl)(phenyl) methanone*.

1.6 Manfaat Penelitian

Penelitian ini diharapkan dapat memberikan pengetahuan kepada masyarakat, akademisi, peneliti maupun praktisi kesehatan tentang keanekaragaman jamur endofitik dari tanaman ciplukan, bioaktivitas ekstrak jamur endofitik, spesies jamur endofitik potensial, dan senyawa murni dari ekstrak jamur endofitik potensial. Selain itu, senyawa murni jamur endofitik yang berhasil diisolasi dapat bermanfaat untuk dikembangkan menjadi obat standar dalam mengobati penyakit degeneratif atau penyakit akibat infeksi bakteri.

1.7 Kerangka Pikir Penelitian



1.8 State of The Art

Tanaman ciplukan telah digunakan secara tradisional untuk mengatasi demam, infeksi, gangguan liver, diabetes, dan peradangan kronis. Penelitian telah mengkonfirmasi bahwa ekstrak dari berbagai bagian tanaman ciplukan seperti buah, daun, batang dan akar, mengandung senyawa sekunder seperti *withanolides* dan *physalins* yang menunjukkan aktivitas antimikroba, anti-inflamasi, antidiabetik, dan antioksidan yang signifikan. Studi kimia-farmakologis pada ciplukan mengungkapkan lebih dari 350 senyawa withanolide, termasuk physangulidines A–C, yang menunjukkan efek antiproliferatif terhadap sel tumor. Beberapa physalin, seperti physalin F, telah dilaporkan memiliki aktivitas immunosupresif terhadap sel mononuklear pasien dengan HTLV-1-associated myelopathy. Selain itu, aktivitas inhibisi nitric oxide oleh ekstrak tanaman ini telah menunjukkan potensi sebagai agen anti-inflamasi melalui modulasi jalur NF- κ B. Secara keseluruhan, data kimia dan bioaktivitas mendukung penggunaan tradisional ciplukan sebagai sumber senyawa terapeutik alami (Vargas-Arana *et al.*, 2025; Wang *et al.*, 2021b).

Eksplorasi senyawa tidak berhenti pada inang tanaman ciplukan. Perhatian dunia juga tertuju pada mikroorganisme yang hidup simbiotik di dalam jaringan tanaman yaitu jamur endofitik. Jamur endofitik telah diakui sebagai reservoir metabolit sekunder bioaktif seperti alkaloid, polifenol, terpenoid, steroid, isokumarin, dan lain-lain yang memiliki potensi antikanker, antioksidan, antimikroba, dan antidiabetik (Ebadi & Ebadi, 2024). Studi umum menunjukkan jamur endofitik dari genus *Colletotrichum*, *Penicillium*, *Fusarium*, dan *Epicoccum* mampu menghasilkan senyawa dengan aktivitas farmakologis kuat, termasuk taxol dan emodin (Hashem *et al.*, 2023). Khusus tanaman ciplukan, terdapat 14 isolat jamur endofitik dari bunga, batang, daun, dan kelopak buah tanaman, menunjukkan aktivitas antioksidan dan antibakteri. Jamur endofitik *Hyphomycetes* (dari bunga) menunjukkan aktivitas antioksidan moderat dengan IC₅₀ 52,43 μ g/mL), dan enam isolat lainnya menunjukkan aktivitas antibakteri kuat terhadap *Escherichia coli* dan *Staphylococcus aureus* dengan MIC 8–64 μ g/mL. Isolat-isolat tersebut termasuk satu *Fusarium* spp. dari batang dan empat *Colletotrichum* spp. dari daun dan kelopak buah yang menunjukkan aktivitas antibakteri terbaik (Palupi *et al.*, 2021).

Penelitian lain berhasil mengidentifikasi berbagai jenis jamur endofitik dari tanaman ciplukan seperti *Penicillium verrucosum*, *Colletotrichum alienum*, *Fusarium subglutinans*, *Aspergillus nidulans*, *Mycelia sterilia*, dan *Rhizoctonia* spp. Metabolit yang dihasilkan oleh *Penicillium verrucosum* menunjukkan aktivitas antimikroba tertinggi terhadap *B. subtilis*, *E. coli*, dan *S. aureus* (Hastuti *et al.*, 2021).

Penelitian jamur endofitik pada ciplukan sampai saat ini sebagian besar hanya mencakup ekstrak kasar, profil KLT dan aktivitas bioaktif umum, tanpa isolasi dan karakterisasi sistematis senyawa bioaktif murni. Jamur endofitik *Colletotrichum* yang juga ada pada ciplukan mampu memproduksi senyawa antimikroba seperti emodin dan taxol pada inang lain (Yehia, 2023; Hashem *et al.*, 2023). Selanjutnya, banyak jamur endofitik menghasilkan berbagai jenis senyawa bioaktif seperti flavonoid, fenolik, terpenoid, isokumarin dan steroid yang belum dievaluasi dari tanaman ciplukan secara terstruktur (Anwar *et al.*, 2023; Bando *et al.*, 2024; Yu *et al.*, 2023).

Berdasarkan data hasil penelitian, terdapat gap penelitian proposional yaitu belum ada laporan isolasi dan karakterisasi lengkap senyawa murni dari jamur endofitik yang berasal dari tanaman ciplukan, sementara jamur endofitik *Colletotrichum*, *Fusarium*, dan *Penicillium* yang terbukti aktif sebagai antioksidan atau antibakteri belum dievaluasi lebih lanjut ke arah senyawa murni. Oleh karena itu, arah penelitian ke depan sangat penting untuk fokus pada isolasi fraksi aktif dari ekstrak endofitik ciplukan dengan pendekatan kromatografi (KLT, kolom) dan karakterisasi struktural senyawa murni menggunakan spektroskopi NMR. Setiap senyawa yang diisolasi harus diuji aktivitas antioksidan, antibakteri, sitotoksitas, dan menentukan nilai IC_{50} atau MIC. Kemudian, membandingkan sifat bioaktif senyawa murni endofit dengan senyawa yang diperoleh dari tanaman itu sendiri, seperti physalin atau withanolide, akan memberikan pemahaman lebih dalam tentang kontribusi mikroba endofitik terhadap khasiat farmakologis ciplukan tradisional. Lebih lanjut, penelitian ini dapat diarahkan untuk identifikasi genetik (DNA ITS sequencing) dari isolat jamur endofitik yang terbukti aktif sebagai antioksidan atau antibakteri.

DAFTAR PUSTAKA

- Abdul-Halim, A. M. A. A., Shivanand, P., Krishnamoorthy, S., & Taha, H. (2023). A review on the biological properties of *Trichoderma* spp. as a prospective biocontrol agent and biofertilizer. *Journal of Applied Biology and Biotechnology*, 11(5), 34–46. <https://doi.org/10.7324/JABB.2023.11504>
- Abdulahadi, S. Y., Hasan, G. Q., & Gergees, R. N. (2020). Molecular detection and antimicrobial activity of Endophytic fungi isolated from a medical plant *Rosmarinus officinalis*. *Annals of Tropical Medicine and Public Health*, 23(13B). <https://doi.org/10.36295/ASRO.2020.231384>
- Abdulkareem, H. M., Abood, A. I., & Dakheel, M. M. (2022). Antimicrobial Resistance of Tannin Extract against *E. coli* Isolates from Sheep. *Archives of Razi Institute*, 77(2), 697–701. <https://doi.org/10.22092/ARI.2022.356982.1955>
- Abramczyk, B., Marzec-Grzadziel, A., Grzadziel, J., Król, E., Gałazka, A., & Oleszek, W. (2022). Biocontrol Potential and Catabolic Profile of Endophytic *Diaporthe eres* Strain 1420S from *Prunus domestica* L. in Poland—A Preliminary Study. *Agronomy*, 12(1). <https://doi.org/10.3390/agronomy12010165>
- Abumaali, D. A., Al-Hadidi, S. H., Daly Yahia, M. N., Erfanian, M. B., Ahmed, T. A., & Alatalo, J. M. (2023). The date palm microbiome: A review. *Ecological Genetics and Genomics*, 29(May). <https://doi.org/10.1016/j.egg.2023.100212>
- Adione, N. M., Peter, I., Christiana, C., Nkasi, N., & Maduabuchi, P. (2022). Antimicrobial Activities of the Endophytic Fungus , *Fusarium equiseti* , Isolated from The leaves of *Ocimum gratissimum*. *Journal of Advances in Medical and Pharmaceutical Sciences*, 24(7), 11–23. <https://doi.org/10.9734/JAMPS/2022/v24i730312>
- Aini, K., Widjajanti, H., Setiawan, A., & Kurniawati, A. R. (2022). Antibacterial activity of endophytic fungi isolated from the stem bark of jambu mawar (*Syzygium jambos*). *Biodiversitas*, 23(1), 521–532. <https://doi.org/10.13057/biodiv/d230156>
- Aisyiyah, I. N., Rahmawati, H., Agustini, D. M., Purbaya, S., Aisyah, L. S., & Yun, Y. F. (2023). ANTIOXIDANT PROPERTIES OF THE ETHYL ACETATE EXTRACT OF ENDOPHYTIC FUNGUS *Penicillium Citrinum* FROM *Kalanchoe Millotii* STEM THROUGH SECONDARY METABOLITES. *Al-Kimiya*, 10(2), 123–132. <https://doi.org/10.15575/ak.v10i2.30323>
- Akram, S., Ahmed, A., He, P., He, P., Liu, Y., Wu, Y., Munir, S., & He, Y. (2023). Uniting the Role of Endophytic Fungi against Plant Pathogens and Their Interaction. *Journal of Fungi*, 9(1). <https://doi.org/10.3390/jof9010072>
- Al-Bader, S. M. (2023). Proteolytic Activity and Health Impact of Fungi Isolated

- from Chickens Feathers in House Breeding Cages. *International Journal of Agriculture and Biology*, 30(2), 154–160. <https://doi.org/10.17957/IJAB/15.2070>
- Al-Khayri, J. M., Rashmi, R., Toppo, V., Chole, P. B., Banadka, A., Sudheer, W. N., Nagella, P., Shehata, W. F., Al-Mssallem, M. Q., Alessa, F. M., Almaghasla, M. I., & Rezk, A. A. S. (2023). Plant Secondary Metabolites: The Weapons for Biotic Stress Management. *Metabolites*, 13(6), 1–37. <https://doi.org/10.3390/metabo13060716>
- Al-Obaidi, J. R., Jambari, N. N., & Ahmad-Kamil, E. I. (2021). Mycopharmaceuticals and nutraceuticals: Promising agents to improve human well-being and life quality. *Journal of Fungi*, 7(7). <https://doi.org/10.3390/jof7070503>
- Al Mamari, H. (2021). *Phenolic Compounds: Classification, Chemistry, and Updated Techniques of Analysis and Synthesis* (F. Badria (ed.)). IntechOpen. <https://doi.org/10.5772/intechopen.98958>
- Alam, B., Li, J., Gě, Q., Khan, M. A., Gōng, J., Mehmood, S., Yuán, Y., & Gōng, W. (2021). Endophytic Fungi: From Symbiosis to Secondary Metabolite Communications or Vice Versa? *Front Plant Sci*, 12(December), 1–24. <https://doi.org/10.3389/fpls.2021.791033>
- Alara, O. R., Abdurahman, N. H., & Ukaegbu, C. I. (2021). Extraction of phenolic compounds: A review. *Current Research in Food Science*, 4(March), 200–214. <https://doi.org/10.1016/j.crfs.2021.03.011>
- Alawode, T. T., Lajide, L., Olaleye, M., & Owolabi, B. (2021). Stigmasterol and β -Sitosterol: Antimicrobial Compounds in the Leaves of *Icacina trichantha* identified by GC–MS. *Beni-Suef University Journal of Basic and Applied Sciences*, 10(1). <https://doi.org/10.1186/s43088-021-00170-3>
- Ali, F., Akhtar, N., Shafique, S., & Shafique, S. (2021). Isolation and identification of *Aspergilli* causing Banana fruit rot. *Open Journal of Chemistry*, 4(1), 8–18. <https://doi.org/10.30538/psrp-ojc2021.0019>
- Ali, M. A., Ahmed, T., Ibrahim, E., Rizwan, M., Chong, K. P., & Yong, J. W. H. (2024). A review on mechanisms and prospects of endophytic bacteria in biocontrol of plant pathogenic fungi and their plant growth-promoting activities. *Heliyon*, 10(11), e31573. <https://doi.org/10.1016/j.heliyon.2024.e31573>
- Ali, W. M., Abdel-Mageed, M. A., Hegazy, M. G. A., Abou-Shlell, M. K., Sultan, S. M. E., Salama, E. A. A., & Yousef, A. F. (2023). Biocontrol agent of root-knot nematode *Meloidogyne javanica* and root-rot fungi, *Fusarium solani* in okra morphological, anatomical characteristics and productivity under greenhouse conditions. *Scientific Reports*, 13(1), 1–13.

<https://doi.org/10.1038/s41598-023-37837-z>

- Aljeldah, M. M., El-Sayyad, H., Elhadi, N., & Rabaan, A. A. (2019). Effect of Gamma-Rays on the Growth and Penicillin Production of *Penicillium chrysogenum*. *Journal of Pure and Applied Microbiology*, 13(2), 779–788. <https://doi.org/10.22207/JPAM.13.2.13>
- Alomran, M., Houbraken, J., & Newcombe, G. (2020). *Aspergillus tubingensis* is a pre-emergent pathogen of date palm seedlings. *Forests*, 11(12), 1–8. <https://doi.org/10.3390/f11121327>
- Alshaibani, D., Zhang, R., & Wu, V. C. H. (2017). Antibacterial characteristics and activity of *Vaccinium macrocarpon* proanthocyanidins against diarrheagenic *Escherichia coli*. *Journal of Functional Foods*, 39, 133–138. <https://doi.org/10.1016/j.jff.2017.10.003>
- Ameen, M., Mahmood, A., Sahkoo, A., Zia, M. A., & Ullah, M. S. (2024). The role of endophytes to combat abiotic stress in plants. *Plant Stress*, 12(September 2023), 100435. <https://doi.org/10.1016/j.stress.2024.100435>
- Ampomah-Wireko, M., Luo, C., Cao, Y., Wang, H., Nininahazwe, L., & Wu, C. (2021). Chemical probe of AHL modulators on quorum sensing in Gram-Negative Bacteria and as antiproliferative agents: A review. *European Journal of Medicinal Chemistry*, 226, 113864. <https://doi.org/10.1016/j.ejmech.2021.113864>
- Andrés, C. M. C., Lastra, J. M. P. de la, Juan, C. A., & Pérez-Lebeña, E. (2024). Michael Acceptors as Anti-Cancer Compounds: Coincidence or Causality? *Int. J. Mol. Sci.*, 25, 2–48.
- Angeles, D. M., & Scheffers, D.-J. (2021). The Cell Wall of *Bacillus subtilis*. *Curr. Issues Mol. Biol.*, 41, 539–596.
- Anh, N. M., Huyen, V. T. T., Quyen, V. T., Dao, P. T., Quynh, D. T., Huong, D. T. M., Van Cuong, P., Dat, T. T. H., & Minh, L. T. H. (2023). Antimicrobial and Cytotoxic Secondary Metabolites from a Marine-Derived Fungus *Penicillium Citrinum* VM6. *Current Microbiology*, 81(1), 32. <https://doi.org/10.1007/s00284-023-03568-7>
- Annadurai, P. (2021). Extraction and isolation of bioactive compounds from lantana camara leaves by column chromatographic techniques. *Research Journal of Pharmacy and Technology*, 14(3), 1607–1611. <https://doi.org/10.5958/0974-360X.2021.00285.7>
- Anwar, R., Pratama, G. B., Supratman, U., Harneti, D., Azhari, A., Fajriah, S., Azmi, M. N., & Shiono, Y. (2023). Steroids from *Toona sureni*-derived Endophytic Fungi *Stemphylium* sp. MAFF 241962 and Their Heme Polymerization Inhibition Activity. *Natural and Life Sciences Communications*, 22(3), 1–29. <https://doi.org/10.12982/NLSC.2023.055>

- Arruda, J. C. C., Rocha, N. C., Santos, E. G., Ferreira, L. G. B., Bello, M. L., Penido, C., Costa, T. E. M. M., Santos, J. A. A., Ribeiro, I. M., Tomassini, T. C. B., & Faria, R. X. (2021). Physalin pool from *Physalis angulata* L. leaves and physalin D inhibit P2X7 receptor function in vitro and acute lung injury in vivo. *Biomedicine and Pharmacotherapy*, 142(February), 112006. <https://doi.org/10.1016/j.biopha.2021.112006>
- Aryal, S. (2021). *McFarland Standards- Principle , Preparation , Uses , Limitations* P. <https://microbenotes.com/mcfarland-standards>
- Aryal, S. (2022). *Mueller Hinton Agar (MHA) – Composition , Principle , Uses and Preparation*. <https://microbiologyinfo.com/mueller-hinton-agar-mha-composition-principle-uses-and-preparation>
- Ashoka, G. B., & Shivanna, M. B. (2023). Antibacterial, antioxidant, and anticancer activities of *Penicillium citrinum* Thom. endophytic in *Jatropha heynei*. *Journal of Applied Pharmaceutical Science*, 13(3), 196–207. <https://doi.org/10.7324/JAPS.2023.95958>
- Asomadu, R. O., Ezeorba, T. P. C., Ezike, T. C., & Uzoechina, J. O. (2024). Exploring the antioxidant potential of endophytic fungi: a review on methods for extraction and quantification of total antioxidant capacity (TAC). 3 *Biotech*, 14(5), 1–18. <https://doi.org/10.1007/s13205-024-03970-3>
- Atkins, S. D., Clark, I. M., Pande, S., Hirsch, P. R., & Kerry, B. R. (2005). The use of real-time PCR and species-specific primers for the identification and monitoring of *Paecilomyces lilacinus*. *FEMS Microbiology Ecology*, 51(2), 257–264. <https://doi.org/10.1016/j.femsec.2004.09.002>
- Bahri, S., Setiawan, W. A., Setiawan, F., Lutfiah, R., Juliasih, N. L. G. R., Ambarwati, Y., Ahmadi, P., Arai, M., Hendri, J., Hadi, S., & Setiawan, A. (2024). Activity of Mangrove-Derived *Fusarium equiseti* 20CB07RF Extract Against Clinical, Antibacterial-Resistant *Pseudomonas aeruginosa*. *Science and Technology Indonesia*, 9(3).
- Bai, J., Zhang, Y., Tang, C., Hou, Y., Ai, X., Chen, X., Zhang, Y., Wang, X., & Meng, X. (2021). Gallic acid: Pharmacological activities and molecular mechanisms involved in inflammation-related diseases. *Biomedicine and Pharmacotherapy*, 133(November 2020), 110985. <https://doi.org/10.1016/j.biopha.2020.110985>
- Bakrim, S., Benkhaira, N., Bourais, I., Benali, T., Lee, L. H., El Omari, N., Sheikh, R. A., Goh, K. W., Ming, L. C., & Bouyahya, A. (2022). Health Benefits and Pharmacological Properties of Stigmasterol. *Antioxidants*, 11(10), 1–32. <https://doi.org/10.3390/antiox11101912>
- Baliyan, S., Mukherjee, R., Priyadarshini, A., Vibhuti, A., Gupta, A., Pandey, R. P., & Chang, C. M. (2022). Determination of Antioxidants by DPPH Radical Scavenging Activity and Quantitative Phytochemical Analysis of *Ficus*

- religiosa. *Molecules*, 27(4). <https://doi.org/10.3390/molecules27041326>
- Ball, J., Panerai, R. B., Williams, C. A. L., & Beishon, L. (2022). Principal component analysis to identify the major contributors to task-activated neurovascular responses. *Cerebral Circulation - Cognition and Behavior*, 3(October 2021), 100039. <https://doi.org/10.1016/j.cccb.2022.100039>
- Bando, K., Kushibe, R., Kitaoka, N., Tamai, Y., Narisawa, K., & Matsuura, H. (2024). Isolation, structural elucidation, and biological activity of a novel isocoumarin from the dark septate endophytic fungus *Phialocephala fortinii*. 79(3–4), 89–92. <https://doi.org/doi:10.1515/znc-2023-0139>
- Barker, D. (2019). Lignans. *Molecules*, 24(7), 2–5. <https://doi.org/10.3390/molecules24071424>
- Baron, N. C., & Rigobelo, E. C. (2022). Endophytic fungi: a tool for plant growth promotion and sustainable agriculture. *Mycology*, 13(1), 39–55. <https://doi.org/10.1080/21501203.2021.1945699>
- Bashabsheh, R. H. F., AL-Fawares, O., Natsheh, I., Bdeir, R., Al-Khreshieh, R. O., & Bashabsheh, H. H. F. (2024). Staphylococcus aureus epidemiology, pathophysiology, clinical manifestations and application of nano-therapeutics as a promising approach to combat methicillin resistant Staphylococcus aureus. *Pathogens and Global Health*, 118(3), 209–231. <https://doi.org/10.1080/20477724.2023.2285187>
- Bashir, A., Manzoor, M. M., Ahmad, T., Farooq, S., Sultan, P., Gupta, A. P., & Riyaz-Ul-Hassan, S. (2023). Endophytic fungal community of *Rosa damascena* Mill. as a promising source of indigenous biostimulants: Elucidating its spatial distribution, chemical diversity, and ecological functions. *Microbiological Research*, 276(July), 127479. <https://doi.org/10.1016/j.micres.2023.127479>
- Basso Valeria, G., Gabriela, D. B., & Gustavo, R. (2024). Trichoderma spp.: characteristics and applications. *Journal of Applied Biotechnology & Bioengineering*, 11(1), 18–22. <https://doi.org/10.15406/jabb.2024.11.00354>
- Benfield, A. H., & Henriques, S. T. (2020). Mode-of-Action of Antimicrobial Peptides: Membrane Disruption vs. Intracellular Mechanisms. *Frontiers in Medical Technology*, 2(December), 25–28. <https://doi.org/10.3389/fmedt.2020.610997>
- Bhaskar, R., Xavier, L. S. E., Udayakumaran, G., Kumar, D. S., Venkatesh, R., & Nagella, P. (2022). Biotic elicitors: a boon for the in-vitro production of plant secondary metabolites. *Plant Cell, Tissue and Organ Culture*, 149(1–2), 7–24. <https://doi.org/10.1007/s11240-021-02131-1>
- Biang, J. (2022). The Importance of Catechins. *Oxidants and Antioxidants in Medical Science*, 11(2), 1.

- Bibi, A., Mubeen, F., Rizwan, A., Ullah, I., Hammad, M., Waqas, M. A. B., Ikram, A., Abbas, Z., Halterman, D., & Saeed, N. A. (2024). Morpho-Molecular Identification of *Fusarium equiseti* and *Fusarium oxysporum* Associated with Symptomatic Wilting of Potato from Pakistan. *Journal of Fungi*, 10(10). <https://doi.org/10.3390/jof10100701>
- Bina, E. F. S., Irawan, B., Setiawan, W. A., & Ekowati, C. N. (2022). Artikel Review: Aplikasi Inokulum Fungi *Trichoderma* spp. Untuk Pertumbuhan dan Penekan Fitopatogen. *Jurnal Biologi Papua*, 14(2), 158–168. <https://doi.org/10.31957/jbp.2377>
- Boban, T., Nadar, S., & Tauro, S. (2023). Breaking down bacterial communication: a review of quorum quenching agents. *Future Journal of Pharmaceutical Sciences*, 9(1). <https://doi.org/10.1186/s43094-023-00526-9>
- Brereton, R. G. (2022). Graphical introduction to principal components analysis. *Journal of Chemometrics*, 36(6), 1–8. <https://doi.org/10.1002/cem.3404>
- Budiono, Elfita, Muharni, Yohandini, H., & Widjajanti, H. (2019). Antioxidant activity of *Syzygium samarangense* L. and their endophytic fungi. *Molekul*, 14(1), 48–55. <https://doi.org/10.20884/1.jm.2019.14.1.503>
- Byrne, M. K., Miellet, S., McGlinn, A., Fish, J., Meedya, S., Reynolds, N., & Van Oijen, A. M. (2019). The drivers of antibiotic use and misuse: The development and investigation of a theory driven community measure. *BMC Public Health*, 19(1), 1–11. <https://doi.org/10.1186/s12889-019-7796-8>
- Carneiro, R. da S., de Almeida Da Costa, M. H., Santos, J. Z. L. V., Uchôa, V. T., da Silva Lopes, L., & Mendes, A. N. (2025). Therapeutic actions of terpenes in neurodegenerative disorders and their correlations with the regulation and remodeling of the extracellular matrix. *Biocell*, 49(1), 109–125. <https://doi.org/10.32604/biocell.2024.058405>
- Carvalho, N., da Mata, L. R., Ribeiro, P. G., Moreira, A. L. D. C., Pinedo, A. S., Ribeiro, C. S. d. C., Souza, N. O. S., & Buso, G. S. C. (2021). Morphological and anatomical characterization of peduncle, flower and fruit related to easy fruit abscission of *Capsicum chinense* (Solanaceae) genotypes. *Genetics and Molecular Research*, 20(4). <https://doi.org/10.4238/GMR18887>
- Chang, Y., Che, Q., Xing, L., Ma, C., Han, Y., Zhu, T., Pfeifer, B. A., Peng, J., Zhang, G., & Li, D. (2021). Antibacterial p-Terphenyl with a Rare 2,2'-Bithiazole Substructure and Related Compounds Isolated from the Marine-Derived Actinomycete *Nocardiosis* sp. HDN154086. *Journal of Natural Products*, 84(4), 1226–1231. <https://doi.org/10.1021/acs.jnatprod.0c01296>
- Chaudhary, P., Janmeda, P., Docea, A. O., Yeskaliyeva, B., Abdull Razis, A. F., Modu, B., Calina, D., & Sharifi-Rad, J. (2023). Oxidative stress, free radicals and antioxidants: potential crosstalk in the pathophysiology of human diseases. *Frontiers in Chemistry*, 11(May), 1–24.

<https://doi.org/10.3389/fchem.2023.1158198>

- Chen, H., Singh, H., Bhardwaj, N., Bhardwaj, S. K., Khatri, M., Kim, K. H., & Peng, W. (2022). An exploration on the toxicity mechanisms of phytotoxins and their potential utilities. *Critical Reviews in Environmental Science and Technology*, 52(3), 395–435. <https://doi.org/10.1080/10643389.2020.1823172>
- Chen, Q., Li, Z., Qu, Z., Zhou, H., Qi, Y., Liu, Z., & Zhang, M. (2020). Maize yield and root morphological characteristics affected by controlled-release diammonium phosphate and *Paecilomyces variotii* extracts. *Field Crops Research*, 255(April), 107862. <https://doi.org/10.1016/j.fcr.2020.107862>
- Chen, X., Li, X., Zhang, G., Wang, C., & Li, C. (2024). Application of Synthetic Biology to the Biosynthesis of Polyketides. *Synthetic Biology and Engineering*, 2(3), 10012–10012. <https://doi.org/10.35534/sbe.2024.10012>
- Chhabra, S., Taksande, A. B., & Munjewar, P. (2024). The Penicillin Pioneer: Alexander Fleming's Journey to a Medical Breakthrough. *Cureus*, 16(7). <https://doi.org/10.7759/cureus.65179>
- Cho, H. U., Yoon, N. Y., Noh, H. J., Kim, Y. I., & Kim, S. H. (2024). Six Unrecorded Species of Sordariomycetes Isolated from Soil in the Republic of Korea. *Korean Journal of Mycology*, 52(4), 349–370. <https://doi.org/10.4489/kjm.520414>
- Christaki, E., Giannenas, I., Bonos, E., & Florou-Paneri, P. (2019). Innovative uses of aromatic plants as natural supplements in nutrition. *Feed Additives: Aromatic Plants and Herbs in Animal Nutrition and Health, October 2022*, 19–34. <https://doi.org/10.1016/B978-0-12-814700-9.00002-9>
- Collins, J. A., & Osherooff, N. (2024). Gyrase and Topoisomerase IV: Recycling Old Targets for New Antibacterials to Combat Fluoroquinolone Resistance. *ACS Infectious Diseases*, 10(4), 1097–1115. <https://doi.org/10.1021/acsinfecdis.4c00128>
- Corsini, P. M., Wang, S., Rehman, S., Fenn, K., Sagar, A., Sirovica, S., Cleaver, L., Edwards-Gayle, C. J. C., Mastroianni, G., Dorgan, B., Sewell, L. M., Lynham, S., Iuga, D., Franks, W. T., Jarvis, J., Carpenter, G. H., Curtis, M. A., Bernadó, P., Darbari, V. C., & Garnett, J. A. (2022). Molecular and cellular insight into *Escherichia coli* SslE and its role during biofilm maturation. *Npj Biofilms and Microbiomes*, 8(1), 1–16. <https://doi.org/10.1038/s41522-022-00272-5>
- Cortês, M., de Haas, A., Unterbusch, R., Fujimori, A., Schütze, T., Meyer, V., & Moeller, R. (2020). *Aspergillus niger* Spores Are Highly Resistant to Space Radiation. *Frontiers in Microbiology*, 11(April), 1–12. <https://doi.org/10.3389/fmicb.2020.00560>

- da Silva Santos, S. da S., da Silva, A. A. da, Polonio, J. C., Polli, A. D., Orlandelli, R. C., dos Santos Oliveira, J. A. D. S., Brandão Filho, J. U. T., Azevedo, J. L., & Pamphile, J. A. (2022). Influence of plant growth-promoting endophytes *Colletotrichum siamense* and *Diaporthe masirevici* on tomato plants (*Lycopersicon esculentum* Mill.). *Mycology*, 13(4), 257–270. <https://doi.org/10.1080/21501203.2022.2050825>
- Dai, Z. B., Wang, X., & Li, G. H. (2020). Secondary Metabolites and Their Bioactivities Produced by *Paecilomyces*. *Molecules*, 25(21), 1–18. <https://doi.org/10.3390/molecules25215077>
- Damen, F., Simo Mpetga, J. D., Demgne, O. M. F., Çelik, İ., Wamba, B. E. N., Tapondjou, L. A., Beng, V. P., Levent, S., Kuete, V., & Tene, M. (2022). Roeperone A, a new tetraoxygenated xanthone and other compounds from the leaves of *Hypericum roeperianum* Schimp. (Hypericaceae). *Natural Product Research*, 36(8), 2071–2077. <https://doi.org/10.1080/14786419.2020.1845674>
- Danişman, B., Kelek, S. E., & Aslan, M. (2023). Resveratrol in Neurodegeneration, in Neurodegenerative Diseases, and in the Redox Biology of the Mitochondria. *Psychiatry and Clinical Psychopharmacology*, 33(2), 147–155. <https://doi.org/10.5152/pcp.2023.23633>
- Das, A. K., Islam, M. N., Faruk, M. O., Ashaduzzaman, M., & Dungani, R. (2020). Review on tannins: Extraction processes, applications and possibilities. *South African Journal of Botany*, 135, 58–70. <https://doi.org/10.1016/j.sajb.2020.08.008>
- Datta, C., Jha, R. K., Ahmed, W., Ganguly, S., Ghosh, S., & Nagaraja, V. (2019). Physical and functional interaction between nucleoid-associated proteins HU and Lsr2 of *Mycobacterium tuberculosis*: altered DNA binding and gene regulation. *Molecular Microbiology*, 111(4), 981–994. <https://doi.org/10.1111/mmi.14202>
- De Almeida, A. J. P. O., De Oliveira, J. C. P. L., Da Silva Pontes, L. V., De Souza Júnior, J. F., Gonçalves, T. A. F., Dantas, S. H., De Almeida Feitosa, M. S., Silva, A. O., & De Medeiros, I. A. (2022). ROS: Basic Concepts, Sources, Cellular Signaling, and its Implications in Aging Pathways. *Oxidative Medicine and Cellular Longevity*, 2022. <https://doi.org/10.1155/2022/1225578>
- de Oliveira, A. M., Malunga, L. N., Perussello, C. A., Beta, T., & Ribani, R. H. (2020). Phenolic acids from fruits of *Physalis angulata* L. in two stages of maturation. *South African Journal of Botany*, 131(May), 448–453. <https://doi.org/10.1016/j.sajb.2020.02.029>
- Dędek, K., Rosicka-Kaczmarek, J., Nebesny, E., & Kowalska, G. (2019). Biotechnology and Food Science Characteristics and biological properties of ferulic acid. *Biotechnol Food Sci*, 83(1), 71–85. <http://www.bfs.p.lodz.pl>

- Degani, O., Khatib, S., Becher, P., Gordani, A., & Harris, R. (2021). Trichoderma asperellum secreted 6-pentyl- α -pyrone to control magnaporthiopsis maydis, the maize late wilt disease agent. *Biology*, 10(9). <https://doi.org/10.3390/biology10090897>
- Deng, S., Liu, Q., Chang, W., Liu, J., & Wang, H. (2023). First specific detection and validation of tomato wilt caused by Fusarium brachygibbosum using a PCR assay. *PeerJ*, 11, 1–15. <https://doi.org/10.7717/peerj.16473>
- Dhayanithy, G., Subban, K., & Chelliah, J. (2019). Diversity and biological activities of endophytic fungi associated with Catharanthus roseus. *BMC Microbiology*, 19(1), 1–14. <https://doi.org/10.1186/s12866-019-1386-x>
- Dias, M. C., Pinto, D. C. G. A., & Silva, A. M. S. (2021). Plant flavonoids: Chemical characteristics and biological activity. *Molecules*, 26(17), 1–16. <https://doi.org/10.3390/molecules26175377>
- Díaz-García, E., Valenzuela-Quintanar, A. I., Sánchez-Estrada, A., González-Mendoza, D., Tiznado-Hernández, M. E., Islas-Rubio, A. R., & Troncoso-Rojas, R. (2024). Phenolic Compounds Synthesized by Trichoderma longibrachiatum Native to Semi-Arid Areas Show Antifungal Activity against Phytopathogenic Fungi of Horticultural Interest. *Microbiology Research*, 15(3), 1425–1440. <https://doi.org/10.3390/microbiolres15030096>
- Ding, Z., Tao, T., Wang, L., Zhao, Y., Huang, H., Zhang, D., Liu, M., Wang, Z., & Han, J. (2019). Bioprospecting of novel and bioactive metabolites from endophytic fungi isolated from rubber tree ficus elastica leavess. *Journal of Microbiology and Biotechnology*, 29(5), 731–738. <https://doi.org/10.4014/jmb.1901.01015>
- Divekar, P. A., Narayana, S., Divekar, B. A., Kumar, R., Gadratagi, B. G., Ray, A., Singh, A. K., Rani, V., Singh, V., Singh, A. K., Kumar, A., Singh, R. P., Meena, R. S., & Behera, T. K. (2022). Plant Secondary Metabolites as Defense Tools against Herbivores for Sustainable Crop Protection. *International Journal of Molecular Sciences*, 23(5). <https://doi.org/10.3390/ijms23052690>
- Dong, Z., Manawasinghe, I. S., Huang, Y., Shu, Y., Phillips, A. J. L., Dissanayake, A. J., Hyde, K. D., Xiang, M., & Luo, M. (2021). Endophytic Diaporthe Associated With Citrus grandis cv. Tomentosa in China. *Frontiers in Microbiology*, 11(February), 1–20. <https://doi.org/10.3389/fmicb.2020.609387>
- Du, W., Yao, Z., Li, J., Id, C. S., Xia, J., Wang, B., Shi, D., & Ren, L. (2020). Diversity and antimicrobial activity of endophytic fungi isolated from Securinega suffruticosa in the Yellow River Delta. 1–18.
- Du, W., Yao, Z., Li, J., Sun, C., Xia, J., Wang, B., Shi, D., & Ren, L. (2020). Diversity and antimicrobial activity of endophytic fungi isolated from Securinega suffruticosa in the Yellow River Delta. *PLoS ONE*, 15(3), 1–18.

<https://doi.org/10.1371/journal.pone.0229589>

- Dudaš, T., Todić, A., Budakov, D., Grahovac, M., Loc, M., & Stojšin, V. (2022). *Penicillium*: Species causing blue mold on stored apple fruits. *Biljni Lekar*, 50, 92–100. <https://doi.org/10.5937/BiljLek2202092D>
- Duta-Bratu, C. G., Nitulescu, G. M., Mihai, D. P., & Olaru, O. T. (2023). Resveratrol and Other Natural Oligomeric Stilbenoid Compounds and Their Therapeutic Applications. *Plants*, 12(16). <https://doi.org/10.3390/plants12162935>
- Dutra, R., Nogueira, A., Rossi, S., Chacon Dória, L., Buttò, V., & Marcatti, C. R. (2023). The Role of Organ and Leaf Habit on the Secondary Xylem Anatomy Variation across 15 Species from Brazilian Cerrado. *Forests*, 14(2), 1–13. <https://doi.org/10.3390/f14020269>
- Ebadi, M., & Ebadi, A. (2024). *Importance and Applications of Endophytic Fungi* (J. Putzke (ed.)). IntechOpen. <https://doi.org/10.5772/intechopen.1006383>
- Echeverria, M., Izzi, Y. S., Criado, M. V., & Caputo, C. (2024). Isolation and characterization of dematiaceous endophytic fungi isolated from barley (*Hordeum vulgare* L.) roots and their potential use as phosphate solubilizers. *The Microbe*, 3(June 2023), 100058. <https://doi.org/10.1016/j.microb.2024.100058>
- Ekeke, C., Obute, G. C., & Ogazie, C. A. (2019). ISSN 1995-6673 Pages 1-4 Jordan Journal of Biological Sciences A Comparative Anatomical and Epidermal Analysis of *Physalis angulata* L. and *Physalis micrantha* L. *Jjbs*, 12(1), 1–4.
- Ekwomadu, T. I., & Mwanza, M. (2023). *Fusarium Fungi Pathogens, Identification, Adverse Effects, Disease Management, and Global Food Security: A Review of the Latest Research. Agriculture (Switzerland)*, 13(9). <https://doi.org/10.3390/agriculture13091810>
- El-Zehery, H. R. A., Ashry, N. M., Faiesal, A. A., Attia, M. S., Abdel-Maksoud, M. A., El-Tayeb, M. A., Aufy, M., & El-DougDoug, N. K. (2024). Antibacterial and anticancer potential of bioactive compounds and secondary metabolites of endophytic fungi isolated from *Anethum graveolens*. *Frontiers in Microbiology*, 15(October), 1–10. <https://doi.org/10.3389/fmicb.2024.1448191>
- El Obeid Sewaikit, A. M. (2025). Identification and Antimicrobial Activity of Endophytic Fungi Associated with *Ocimum basilicum* (L.) from Sudan. *International Journal of Current Microbiology and Applied Sciences*, 14(5), 152–169. <https://doi.org/10.20546/ijcmas.2025.1405.016>
- ELFITA, MARDIYANTO, FITRYA, LARASATI, J. E., JULINAR, WIDJAJANTI, H., & MUHARNI. (2019). Antibacterial activity of *Cordyline fruticosa* leaf extracts and its endophytic fungi extracts. *Biodiversitas*, 20(12),

3804–3812. <https://doi.org/10.13057/biodiv/d201245>

- Elfita, Oktiansyah, R., Mardiyanto, Setiawan, A., & Widjajanti, H. (2024). Combination Effect of Extracts and Pure Compounds of Endophytic Fungi Isolated from Sungkai (*Peronema canescens*) Leaves on Antioxidant Activity. *Science and Technology Indonesia*, 9(1), 69–76. <https://doi.org/10.26554/sti.2024.9.1.69-76>
- Elfita, Oktiansyah, R., Mardiyanto, Widjajanti, H., & Setiawan, A. (2022). Antibacterial and antioxidant activity of endophytic fungi isolated from *Peronema canescens* leaves. *Biodiversitas*, 23(9), 4783–4792. <https://doi.org/10.13057/biodiv/d230946>
- Elfita, Oktiansyah, R., Mardiyanto, Widjajanti, H., Setiawan, A., & Nasution, S. S. A. (2023). Bioactive Compounds of Endophytic Fungi *Lasiodiplodia theobromae* Isolated From The Leaves of Sungkai (*Peronema canescens*). *Biointerface Research in Applied Chemistry*, 13(6). <https://doi.org/10.33263/BRIAC136.530>
- Elkhateeb, W. A., Daba, G. M., Elnahas, M., & Thomas, P. (2019). Anticoagulant Capacities of Some Medicinal Mushrooms. *ARC Journal of Pharmaceutical Sciences*, 5(4). <https://doi.org/10.20431/2455-1538.0504001>
- Elkhateeb, W., & Daba, G. (2023). Fungal Pigments: Their Diversity, Chemistry, Food and Non-Food Applications. *Applied Microbiology*, 3(3), 735–751. <https://doi.org/10.3390/applmicrobiol3030051>
- Elshafie, H. S., Camele, I., & Mohamed, A. A. (2023). A Comprehensive Review on the Biological, Agricultural and Pharmaceutical Properties of Secondary Metabolites Based-Plant Origin. *International Journal of Molecular Sciences*, 24(4). <https://doi.org/10.3390/ijms24043266>
- Elumalai, S., Soundararajan, S., Kanagaraj, P., & Sadhasivam, D. R. (2022). In vitro antioxidant and antibacterial activity of Quercetin isolated from *Indigofera aspalathoides* and Quercetin-Zinc metal complex. *International Journal of Health Sciences*, 6(March), 4314–4326. <https://doi.org/10.53730/ijhs.v6ns3.6859>
- Engel, A., Simler-Williamson, A., Ravenscraft, A., Bittleston, L., & de Graaff, M. A. (2024). Interactive effects of fungal community structure and soil moisture on Wyoming big sagebrush performance. *Plant and Soil*, 0123456789. <https://doi.org/10.1007/s11104-024-06809-1>
- Erdoğan, M., Polat Köse, L., Eşsiz, S., & Gülçin, İ. (2021). Synthesis and biological evaluation of some 1-naphthol derivatives as antioxidants, acetylcholinesterase, and carbonic anhydrase inhibitors. *Archiv Der Pharmazie*, 354(8). <https://doi.org/10.1002/ardp.202100113>
- Eze, P. M., Abonyi, D. O., Abba, C. C., Proksch, P., Okoye, F. B. C., & Esimone,

- C. O. (2019). *Toxic , but beneficial compounds from endophytic fungi of Carica papaya*. 3(2), 105–111. <https://doi.org/10.2478/ebtj-2019-0012>
- Fadhilla, G., Adnyana, I. K., & Chaniago, R. (2020). AKTIVITAS ANALGETIK EKSTRAK ETANOL DAUN CIPLUKAN (*Physalis peruviana* L.) PADA MENCIT SWISS WEBSTER JANTAN DENGAN METODE GELIAT (Sigmund). *Jurnal Ilmiah Farmako Bahari*, 11(1), 75. <https://doi.org/10.52434/jfb.v11i1.716>
- Fadhillah, Elfita, Muharni, Yohandini, H., & Widjajanti, H. (2019). Chemical compound isolated from antioxidant active extract of endophytic fungus *Cladosporium tenuissimum* in *Swietenia mahagoni* leaf stalks. *Biodiversitas*, 20(9), 2645–2650. <https://doi.org/10.13057/biodiv/d200929>
- Falahiya, M. T., Bahri, S., Marnita, Y., & Dalimunthe, C. I. (2023). Uji EFEKTIVITAS BEBERAPA ISOLAT *Trichoderma* sp . TERHADAP. *J. Agroplantae*, 21(1), 204–216.
- Fan, M., Chen, X., Luo, X., Zhang, H., Liu, Y., Zhang, Y., Wu, J., Zhao, C., & Zhao, P. (2020). Diversity of endophytic fungi from the leaves of *Vaccinium dunalianum*. *Letters in Applied Microbiology*, 71(5), 479–489. <https://doi.org/10.1111/lam.13345>
- Farha, A. K., Yang, Q. Q., Kim, G., Li, H. Bin, Zhu, F., Liu, H. Y., Gan, R. Y., & Corke, H. (2020). Tannins as an alternative to antibiotics. *Food Bioscience*, 38(August), 100751. <https://doi.org/10.1016/j.fbio.2020.100751>
- Fathallah, N., Elkady, W. M., Zahran, S. A., Darwish, K. M., Elhady, S. S., & Elkhawas, Y. A. (2024). Unveiling the Multifaceted Capabilities of Endophytic *Aspergillus flavus* Isolated from *Annona squamosa* Fruit Peels against *Staphylococcus* Isolates and HCoV 229E—In Vitro and In Silico Investigations. *Pharmaceuticals*, 17(5). <https://doi.org/10.3390/ph17050656>
- Fathoni, A., Ilyas, M., Praptiwi, Wulansari, D., & Agusta, A. (2022). Antibacterial and Antioxidant Activities of Fungal Endophytes Isolated from Medicinal Plants in Simeulue Island, Aceh. *HAYATI Journal of Biosciences*, 29(6), 720–732. <https://doi.org/10.4308/hjb.29.6.720-732>
- Fernandez, M. R., Abdellatif, L., Lokuruge, P., Schellenberg, M. P., & Lupwayi, N. Z. (2022). Root disease and fungal populations in organic crops under different tillage–cropping systems. *Crop Science*, 62(3), 1288–1304. <https://doi.org/10.1002/csc2.20663>
- Ferraz, C. R., Franciosi, A., Emidio, N. B., Rasquel-Oliveira, F. S., Manchope, M. F., Carvalho, T. T., Artero, N. A., Fattori, V., Vicentini, F. T. M. C., Casagrande, R., & Verri, W. A. (2021). *Chapter 15 - Quercetin as an antiinflammatory analgesic* (M. Mushtaq & F. B. T.-A. C. of V. P. B. Anwar (eds.); pp. 319–347). Academic Press. <https://doi.org/https://doi.org/10.1016/B978-0-12-822923-1.00023-6>

- Ferraz, M. P. (2024). Antimicrobial Resistance: The Impact from and on Society According to One Health Approach. *Societies*, 14(9), 1–22. <https://doi.org/10.3390/soc14090187>
- Ferreira, L., Vale, A. do, Souza, A. de, & Leite, K. (2019). Anatomical and phytochemical characterization of *Physalis angulata* L.: A plant with therapeutic potential. *Pharmacognosy Research*, 11, 171–17. <https://doi.org/10.4103/pr.pr>
- Figueiredo, M. C. C., Passos, A. R., Hughes, F. M., Santos, K. S. dos, Silva, A. L. da, & Soares, T. L. (2020). Reproductive biology of *Physalis angulata* L. (Solanaceae). *Scientia Horticulturae*, 267(February), 109307. <https://doi.org/10.1016/j.scienta.2020.109307>
- Foti, M. C. (2015). Use and Abuse of the DPPH• Radical. *Journal of Agricultural and Food Chemistry*, 63(40), 8765–8776. <https://doi.org/10.1021/acs.jafc.5b03839>
- Gabrekiristos, E., & Demiyio, T. (2020). Hot Pepper Fusarium Wilt (*Fusarium oxysporum* f. sp. *capsici*): Epidemics, Characteristic Features and Management Options. *Journal of Agricultural Science*, 12(10), 347. <https://doi.org/10.5539/jas.v12n10p347>
- Gao, Y.-Q., Du, S.-T., Xiao, J., Wang, D.-C., Han, W.-B., Zhang, Q., & Gao, J.-M. (2020). Isolation and Characterization of Antifungal Metabolites from the *Melia azedarach*-Associated Fungus *Diaporthe eucalyptorum*. *Journal of Agricultural and Food Chemistry*, 68(8), 2418–2425. <https://doi.org/10.1021/acs.jafc.9b07825>
- Gao, Y., Xu, Y., Dong, Z., Guo, Y., Luo, J., Wang, F., Yan, L., & Zou, X. (2025). Endophytic Fungal Diversity and Its Interaction Mechanism with Medicinal Plants. *Molecules*, 30(5), 1–22. <https://doi.org/10.3390/molecules30051028>
- Garde, S., Chodisetti, P. K., & Reddy, M. (2021). Peptidoglycan: Structure, Synthesis, and Regulation. *EcoSal Plus*, 9(2). <https://doi.org/10.1128/ecosalplus.esp-0010-2020>
- Gärditz, K. F., & Czesnick, H. (2024). Paclitaxel - a Product of Fungal Secondary Metabolism or an Artefact? *Planta Medica*, 726–735. <https://doi.org/10.1055/a-2309-6298>
- Garrett, A. T. de A., Dos Santos, Á. F., Filho, A. F., Tambarussi, E. V., Gonçalves, A. B., & Garcia, F. A. de O. (2019). Mycelial growth and sporulation of *apoharknessia eucalyptorum* on different culture media. *Summa Phytopathologica*, 45(3), 272–278. <https://doi.org/10.1590/0100-5405/207327>
- GBD. (2022). Global mortality associated with 33 bacterial pathogens in 2019: a systematic analysis for the Global Burden of Disease Study 2019. In *The*

- Lancet* (Vol. 400, Issue 10369, pp. 2221–2248). [https://doi.org/10.1016/S0140-6736\(22\)02185-7](https://doi.org/10.1016/S0140-6736(22)02185-7)
- Ghareeb, R. Y., Jaremko, M., Abdelsalam, N. R., Abdelhamid, M. M. A., EL-Argawy, E., & Ghozlan, M. H. (2024). Biocontrol potential of endophytic fungi against phytopathogenic nematodes on potato (*Solanum tuberosum* L.). *Scientific Reports*, 14(1), 1–14. <https://doi.org/10.1038/s41598-024-64056-x>
- Gizaw, F., Kekeba, T., Teshome, F., Kebede, M., Abreham, T., Berhe, H. H., Ayana, D., Edao, B. M., Waktole, H., Tufa, T. B., Abunna, F., Beyi, A. F., & Abdi, R. D. (2023). Multidrug-Resistant *Staphylococcus aureus* Strains Thrive in Dairy and Beef Production, Processing, and Supply Lines in Five Geographical Areas in Ethiopia. *Veterinary Sciences*, 10(12). <https://doi.org/10.3390/vetsci10120663>
- Gorai, P. S., Barman, S., Gond, S. K., & Mandal, N. C. (2020). Chapter 28 - *Trichoderma* (N. Amaesan, M. Senthil Kumar, K. Annapurna, K. Kumar, & A. B. T.-B. M. in A.-E. Sankaranarayanan (eds.); pp. 571–591). Academic Press. <https://doi.org/https://doi.org/10.1016/B978-0-12-823414-3.00028-9>
- Górniak, I., Bartoszewski, R., & Króliczewski, J. (2019). Comprehensive review of antimicrobial activities of plant flavonoids. In *Phytochemistry Reviews* (Vol. 18, Issue 1). <https://doi.org/10.1007/s11101-018-9591-z>
- Gowtham, H. G., Hema, P., Murali, M., Shilpa, N., Nataraj, K., Basavaraj, G. L., Singh, S. B., Aiyaz, M., Udayashankar, A. C., & Amruthesh, K. N. (2024). Fungal Endophytes as Mitigators against Biotic and Abiotic Stresses in Crop Plants. *Journal of Fungi*, 10(2), 1–34. <https://doi.org/10.3390/jof10020116>
- Grigor'eva, A., Bardasheva, A., Tupitsyna, A., Amirkhanov, N., Tikunova, N., Pyshnyi, D., & Ryabchikova, E. (2020). Changes in the ultrastructure of *staphylococcus aureus* treated with cationic peptides and chlorhexidine. *Microorganisms*, 8(12), 1–19. <https://doi.org/10.3390/microorganisms8121991>
- Gryganskyi, A. P., Golan, J., Muszewska, A., Idnurm, A., Dolatabadi, S., Mondo, S. J., Kutovenko, V. B., Kutovenko, V. O., Gajdeczka, M. T., Anishchenko, I. M., Pawlowska, J., Tran, N. V., Ebersberger, I., Voigt, K., Wang, Y., Chang, Y., Pawlowska, T. E., Heitman, J., Vilgalys, R., ... Stajich, J. E. (2023). Sequencing the Genomes of the First Terrestrial Fungal Lineages: What Have We Learned? *Microorganisms*, 11(7), 1–24. <https://doi.org/10.3390/microorganisms11071830>
- Gu, H., Zhang, S., Liu, L., Yang, Z., Zhao, F., & Tian, Y. (2022). Antimicrobial Potential of Endophytic Fungi From *Artemisia argyi* and Bioactive Metabolites From *Diaporthe* sp. AC1. *Frontiers in Microbiology*, 13(June), 1–13. <https://doi.org/10.3389/fmicb.2022.908836>
- Gu, K., Chen, C. Y., Selvaraj, P., Pavagadhi, S., Yeap, Y. T., Swarup, S., Zheng,

- W., & Naqvi, N. I. (2023). *Penicillium citrinum* Provides Transkingdom Growth Benefits in Choy Sum (*Brassica rapa* var. *parachinensis*). *Journal of Fungi*, 9(4). <https://doi.org/10.3390/jof9040420>
- Gulcin, İ. (2020). Antioxidants and antioxidant methods: an updated overview. *Archives of Toxicology*, 94(3), 651–715. <https://doi.org/10.1007/s00204-020-02689-3>
- Gulcin, İ. (2025). Antioxidants: a comprehensive review. In *Archives of Toxicology*. <https://doi.org/10.1007/s00204-025-03997-2>
- Gulcin, İ., & Alwasel, S. H. (2023). DPPH Radical Scavenging Assay. *Processes*, 11(8). <https://doi.org/10.3390/pr11082248>
- Gultom, E. D., Rambe, R., Paramitha, R., & Ginting, O. S. B. (2021). Uji AKTIVITAS ANTIDIARE EKSTRAK ETANOL DAUN CIPLUKAN (*Physalis minima* L.) TERHADAP MENICT JANTAN (*Mus musculus*). *Forte Journal*, 1(1), 26–44. <https://doi.org/10.51771/fj.v1i1.37>
- Guo, D., Wang, H., Zhang, S., & Lan, T. (2022). The type III polyketide synthase supergene family in plants: complex evolutionary history and functional divergence. *Plant Journal*, 112(2), 414–428. <https://doi.org/10.1111/tpj.15953>
- Gupta, S., Schillaci, M., Walker, R., Smith, P. M. C., Watt, M., & Roessner, U. (2021). Alleviation of salinity stress in plants by endophytic plant-fungal symbiosis: Current knowledge, perspectives and future directions. *Plant and Soil*, 461(1–2), 219–244. <https://doi.org/10.1007/s11104-020-04618-w>
- Hădărugă, N.-G., & Hădărugă, D.-I. (2022). *Stilbenes and Its Derivatives and Glycosides BT - Handbook of Food Bioactive Ingredients: Properties and Applications* (S. M. Jafari, A. Rashidinejad, & J. Simal-Gandara (eds.); pp. 1–58). Springer International Publishing. https://doi.org/10.1007/978-3-030-81404-5_14-1
- Ham, J., Lim, W., Whang, K. Y., & Song, G. (2020). Butylated hydroxytoluene induces dysregulation of calcium homeostasis and endoplasmic reticulum stress resulting in mouse Leydig cell death. *Environmental Pollution*, 256(xxxx), 113421. <https://doi.org/10.1016/j.envpol.2019.113421>
- Hami, A., Rasool, R. S., Khan, N. A., Mansoor, S., & Mir, M. A. (2021). Morpho - molecular identification and first report of *Fusarium equiseti* in causing chilli wilt from Kashmir (Northern Himalayas). *Scientific Reports*, 0123456789, 1–14. <https://doi.org/10.1038/s41598-021-82854-5>
- Hammoumi, D., Al-Aizari, H. S., Alaraidh, I. A., Okla, M. K., Assal, M. E., Al-Aizari, A. R., Moshab, M. S., Chakiri, S., & Bejjaji, Z. (2024). Seasonal Variations and Assessment of Surface Water Quality Using Water Quality Index (WQI) and Principal Component Analysis (PCA): A Case Study. *Sustainability (Switzerland)*, 16(13), 1–22.

<https://doi.org/10.3390/su16135644>

- Han, Y.-K., Dumin, W., Park, M.-J., Bae, Y.-S., Park, J.-H., & Back, C.-G. (2022). *Disease Note Diseases Caused by Fungi and Fungus-Like Organisms First Report of Fusarium Wilt Disease Caused by Fusarium equiseti on Grafted Watermelon in Korea. November, 2989*. <https://doi.org/10.1094/PDIS->
- Haque, S., Sengupta, S., Khan, A., Mukhopadhyay, A. K., Bhan, M. K., Kumar, R., & Jaiikhani, B. (2023). Immune response of S. Typhi-derived Vi polysaccharide and outer membrane protein a conjugate in mice. *Pediatrics and Neonatology*, 64(5), 518–527. <https://doi.org/10.1016/j.pedneo.2022.12.011>
- Hashem, A. H., Attia, M. S., Kandil, E. K., Fawzi, M. M., Abdelrahman, A. S., Khader, M. S., Khodaira, M. A., Emam, A. E., Goma, M. A., & Abdelaziz, A. M. (2023). Bioactive compounds and biomedical applications of endophytic fungi: a recent review. *Microbial Cell Factories*, 22(1), 1–23. <https://doi.org/10.1186/s12934-023-02118-x>
- Hassanpour, S. H., & Doroudi, A. (2023). Review of the antioxidant potential of flavonoids as a subgroup of polyphenols and partial substitute for synthetic antioxidants. *Avicenna Journal of Phytomedicine*, 13(4), 354–376. <https://doi.org/10.22038/AJP.2023.21774>
- Hastuti, U. S., Sulisetijono, Rahmawati, D., Sari, R. Y., Hartono, S., Thoyibah, C., Maulita, F., Ningsih, F. N. I., & Pratiwi, S. H. (2021). Identification and histological observation of endophytic fungus isolated from medicinal plant, physalis angulata L. *HAYATI J Biosci*, 28(2), 130–135. <https://doi.org/10.4308/hjb.28.2.130>
- Hawar, S. N., Taha, Z. K., Hamied, A. S., Al-Shmgani, H. S., Sulaiman, G. M., & Elsilk, S. E. (2023). Antifungal Activity of Bioactive Compounds Produced by the Endophytic Fungus Paecilomyces sp. (JN227071.1) against Rhizoctonia solani. *International Journal of Biomaterials*, 2023. <https://doi.org/10.1155/2023/2411555>
- He, T., Li, X., Iacovelli, R., Hackl, T., & Haslinger, K. (2023). Genomic and Metabolomic Analysis of the Endophytic Fungus Fusarium sp. VM-40 Isolated from the Medicinal Plant Vinca minor. *Journal of Fungi*, 9(7). <https://doi.org/10.3390/jof9070704>
- Hengge, R. (2020). Linking bacterial growth, survival, and multicellularity – small signaling molecules as triggers and drivers. *Current Opinion in Microbiology*, 55, 57–66. <https://doi.org/10.1016/j.mib.2020.02.007>
- Herve, M., & Ndomou, S. C. (2023). *The Use of Plants as Phytobiotics: A New Challenge*. <https://doi.org/10.5772/intechopen.110731>
- Hilje-Rodríguez, I., & Molina-Bravo, R. (2020). Inoculation and co-inoculation of

- two monosporic fungi onto surface-sterile blackberry fruits for quantification experiments. *MethodsX*, 7, 10–13. <https://doi.org/10.1016/j.mex.2020.101092>
- Himawan, C. H., Inawati, & Lubis, A. (2020). AKTIVITAS ANTIOKSIDAN EKSTRAK AIR DAUN JAMBU MAWAR (*Syzygium Jambos* Alston) METODE PERENDAMAN RADIKAL BEBAS DENGAN DPPH. *Jurnal Farmamedika (Pharmamedica Journal)*, 5(2), 52–59. <https://doi.org/10.47219/ath.v5i2.105>
- Holzknicht, J., Kühbacher, A., Papp, C., Farkas, A., Váradi, G., Marcos, J. F., Manzanares, P., Tóth, G. K., Galgóczy, L., & Marx, F. (2020). The penicillium chrysogenum Q176 antimicrobial protein pafc effectively inhibits the growth of the opportunistic human pathogen *Candida albicans*. *Journal of Fungi*, 6(3), 1–21. <https://doi.org/10.3390/jof6030141>
- Hoque, N., Khan, Z. R., Rashid, P. T., Begum, M. N., Sharmin, S., Hossain, M. J., Rana, M. S., & Sohrab, M. H. (2023). Antimicrobial, antioxidant, and cytotoxic properties of endophytic fungi isolated from *Thysanolaena maxima* Roxb., *Dracaena spicata* Roxb. and *Aglaonema hookerianum* Schott. *BMC Complementary Medicine and Therapies*, 23(1), 1–11. <https://doi.org/10.1186/s12906-023-04185-4>
- Hossain, M., Rahman, W., Ali, M., Sultana, T., & Hossain, K. (2021). Identification and Antibioqram Assay of *Escherichia Coli* Isolated From Chicken Eggs. *Journal of Bio-Science*, 29(1), 123–133. <https://doi.org/10.3329/jbs.v29i0.54828>
- Hu, X., Li, X., Yang, S., Liu, H., & Meng, L. (2020). Three New Sesquiterpenoids from the Algal-Derived. *Mar. Drugs*, 18(194), 1–10.
- Huang, J., Xu, W., Zhai, J., Hu, Y., Guo, J., Zhang, C., Zhao, Y., Zhang, L., Martine, C., Ma, H., & Huang, C. H. (2023). Nuclear phylogeny and insights into whole-genome duplications and reproductive development of Solanaceae plants. *Plant Communications*, 4(4), 1–22. <https://doi.org/10.1016/j.xplc.2023.100595>
- Huang, W., Wang, Y., Tian, W., Cui, X., Tu, P., Li, J., Shi, S., & Liu, X. (2022). Biosynthesis Investigations of Terpenoid, Alkaloid, and Flavonoid Antimicrobial Agents Derived from Medicinal Plants. *Antibiotics*, 11(10). <https://doi.org/10.3390/antibiotics11101380>
- Huang, X., Wu, Y., Ni, Y., Xu, H., & He, Y. (2025). Global, regional, and national burden of type 2 diabetes mellitus caused by high BMI from 1990 to 2021, and forecasts to 2045: analysis from the global burden of disease study 2021. *Frontiers in Public Health*, 13(January), 1–12. <https://doi.org/10.3389/fpubh.2025.1515797>
- Huda-Shakirah, A. R., Kee, Y. J., Wong, K. L., Zakaria, L., & Mohd, M. H. (2021). *Diaporthe* species causing stem gray blight of red-fleshed dragon fruit

- (*Hylocereus polyrhizus*) in Malaysia. *Scientific Reports*, 11(1), 1–12. <https://doi.org/10.1038/s41598-021-83551-z>
- Hurtado-Barroso, S., Quifer-Rada, P., Marhuenda-Muñoz, M., Rinaldi de Alvarenga, J. F., Tresserra-Rimbau, A., & Lamuela-Raventós, R. M. (2019). Increase of 4-hydroxybenzoic, a bioactive phenolic compound, after an organic intervention diet. *Antioxidants*, 8(9), 1–12. <https://doi.org/10.3390/antiox8090340>
- Hussein, Z. M., Abedali, A. H., & Ahmead, A. S. (2019). Improvement Properties of Self-Healing Concrete by Using Bacteria. *IOP Conference Series: Materials Science and Engineering*, 584(1). <https://doi.org/10.1088/1757-899X/584/1/012034>
- Hymøller, K. M., Christiansen, S. H., Schlosser, A. G., Skov Sørensen, U. B., Lee, J. C., & Thiel, S. (2024). Recognition of *Staphylococcus aureus* by the pattern recognition molecules langerin, mannan-binding lectin, and surfactant protein D: the influence of capsular polysaccharides and wall teichoic acid. *Frontiers in Immunology*, 15(January), 1–14. <https://doi.org/10.3389/fimmu.2024.1504886>
- Ibrahim, M., Oyebanji, E., Fowora, M., Aiyeolemi, A., Orabuchi, C., Akinshaw, B., & Adekunle, A. A. (2021). Extracts of endophytic fungi from leaves of selected Nigerian ethnomedicinal plants exhibited antioxidant activity. *BMC Complementary Medicine and Therapies*, 21(1), 1–13. <https://doi.org/10.1186/s12906-021-03269-3>
- Igiehon, N. O., Babalola, O. O., Cheseto, X., & Torto, B. (2021). Effects of rhizobia and arbuscular mycorrhizal fungi on yield, size distribution and fatty acid of soybean seeds grown under drought stress. *Microbiological Research*, 242, 126640. <https://doi.org/10.1016/j.micres.2020.126640>
- Ikram, M., Ali, N., Jan, G., Hamayun, M., Jan, F. G., & Iqbal, A. (2019). Novel antimicrobial and antioxidative activity by endophytic *Penicillium roqueforti* and *Trichoderma reesei* isolated from *Solanum surattense*. *Acta Physiologiae Plantarum*, 41(9), 164. <https://doi.org/10.1007/s11738-019-2957-z>
- Inayati, A., Setyowati, L., Aini, L. Q., & Yusnawan, E. (2021). Plant growth promoter produced by *Trichoderma virens* and its effect on mungbean (*Vigna radiata* (L.) Wilczek) seedling. *IOP Conference Series: Earth and Environmental Science*, 803(1). <https://doi.org/10.1088/1755-1315/803/1/012013>
- Indradi, R. B., Muhaimin, M., Barliana, M. I., & Khatib, A. (2023). Potential Plant-Based New Antiplasmodial Agent Used in Papua Island, Indonesia. *Plants*, 12(9). <https://doi.org/10.3390/plants12091813>
- Islam, T., Ara, I., Islam, T., Sah, P. K., Almeida, R. S. de, Matias, E. F. F., Ramalho, C. L. G., Coutinho, H. D. M., & Islam, M. T. (2023). Ethnobotanical uses and

- phytochemical, biological, and toxicological profiles of *Datura metel* L.: A review. *Current Research in Toxicology*, 4(January), 100106. <https://doi.org/10.1016/j.crttox.2023.100106>
- ITIS. (2023). *Physalis angulata* L. In *ITIS - Report: Physalis angulata*. <https://doi.org/10.5860/choice.49-6871>
- Iwansyah, A. C., Luthfiyanti, R., Ardiansyah, R. C. E., Rahman, N., Andriana, Y., & Hamid, H. A. (2022). Antidiabetic activity of *Physalis angulata* L. fruit juice on streptozotocin-induced diabetic rats. *South African Journal of Botany*, 145, 313–319. <https://doi.org/10.1016/j.sajb.2021.08.045>
- Jalab, A. Z., & Al-Rufaye, Z. T. A. (2023). Chemical composition and antioxidant activity of *Physalis angulata* AH-ZE. *World Journal of Advanced Research and Reviews*, 19(2), 1072–1077. <https://doi.org/10.30574/wjarr.2023.19.2.1610>
- Jamali, S. (2015). First report of *Helicocephalum sarcophilum* (Zoopagomycotina) in Iran. *Mycol. Iran*, 2(June 2014), 78.
- Jayalakshmi, R., Oviya, R., Premalatha, K., Mehetre, S. T., Paramasivam, M., Kannan, R., Theradimani, M., Pallavi, M. S., Mukherjee, P. K., & Ramamoorthy, V. (2021). Production, stability and degradation of *Trichoderma* gliotoxin in growth medium, irrigation water and agricultural soil. *Scientific Reports*, 11(1), 1–14. <https://doi.org/10.1038/s41598-021-95907-6>
- Jayatilake, P. L., & Munasinghe, H. (2020). Antimicrobial Activity of Cultivable Endophytic and Rhizosphere Fungi Associated with “mile-a-Minute,” *Mikania cordata* (Asteraceae). *BioMed Research International*, 2020. <https://doi.org/10.1155/2020/5292571>
- Jha, P., Kaur, T., Chhabra, I., Panja, A., Paul, S., Kumar, V., & Malik, T. (2023). Endophytic fungi: hidden treasure chest of antimicrobial metabolites interrelationship of endophytes and metabolites. *Frontiers in Microbiology*, 14(July), 1–16. <https://doi.org/10.3389/fmicb.2023.1227830>
- Jian, Z., Zeng, L., Xu, T., Sun, S., Yan, S., Yang, L., Huang, Y., Jia, J., & Dou, T. (2021). Antibiotic resistance genes in bacteria: Occurrence, spread, and control. *Journal of Basic Microbiology*, 61(12), 1049–1070. <https://doi.org/10.1002/jobm.202100201>
- Jiang, Y., Chen, J., Wang, Z., Zhou, J., & Hu, G. (2025). Mechanical Properties of Korla Fragrant Pear Fruiting Branches and Pedicels: Implications for Non-Destructive Harvesting. *Horticulturae*, 11(8), 880. <https://doi.org/10.3390/horticulturae11080880>
- Jiang, Y., Jin, Y., Shan, Y., Zhong, Q., Wang, H., Shen, C., & Feng, S. (2024). Advances in *Physalis* molecular research: applications in authentication,

- genetic diversity, phylogenetics, functional genes, and omics. *Frontiers in Plant Science*, 15(June), 1–11. <https://doi.org/10.3389/fpls.2024.1407625>
- Jildeh, Z. B., Wagner, P. H., & Schöning, M. J. (2021). Sterilization of Objects, Products, and Packaging Surfaces and Their Characterization in Different Fields of Industry: The Status in 2020. *Physica Status Solidi (A) Applications and Materials Science*, 218(13). <https://doi.org/10.1002/pssa.202000732>
- Jin, L., Yang, L., Li, W., Xu, D., Yang, N., Li, G., & Wan, P. (2021). Diversity and Biocontrol Potential of Culturable Endophytic Fungi in Cotton. *Frontiers in Microbiology*, 12(August), 1–15. <https://doi.org/10.3389/fmicb.2021.698930>
- Jin, X., Zhang, L., Cao, Y., Dai, Z., Ge, X., Cai, R., Wang, R., & Hu, Z. (2024). Antibiotic resistance characterization, virulence factors and molecular characteristics of *Bacillus* species isolated from probiotic preparations in China. *Journal of Global Antimicrobial Resistance*, 43, 35–39. <https://doi.org/10.1016/j.jgar.2024.06.015>
- Jomova, K., Alomar, S. Y., Alwasel, S. H., Nepovimova, E., Kuca, K., & Valko, M. (2024). Several lines of antioxidant defense against oxidative stress: antioxidant enzymes, nanomaterials with multiple enzyme-mimicking activities, and low-molecular-weight antioxidants. In *Archives of Toxicology* (Vol. 98, Issue 5). Springer Berlin Heidelberg. <https://doi.org/10.1007/s00204-024-03696-4>
- Ju, M., Zhang, Q., Wang, R., Yan, S., Zhang, Q., Li, P., Hao, F., & Gu, P. (2024). Community ecological succession of endophytic fungi associates with medicinal compound accumulation in *Sophora alopecuroides*. *Microbiology Spectrum*, 12(2), 1–18. <https://doi.org/10.1128/spectrum.03076-23>
- Kalinga, J., Samarakoon, K., Siriwardhane, U., Yakandawala, D., Adikaram, N., Jayasinghe, L., Araya, H., & Fujimoto, Y. (2025). α -Glucosidase inhibitor and phytotoxic metabolite produced by an endophytic fungus *Diaporthe melonis* isolated from *Acalypha indica* L. *Vegetos*. <https://doi.org/10.1007/s42535-025-01313-y>
- Kalogerakou, T., & Antoniadou, M. (2024). The Role of Dietary Antioxidants, Food Supplements and Functional Foods for Energy Enhancement in Healthcare Professionals. *Antioxidants*, 13(12), 1–37. <https://doi.org/10.3390/antiox13121508>
- Kamran, F., Phillips, M., Harman, D. G., & Reddy, N. (2023). Antioxidant activities of lupin (*Lupinus angustifolius*) protein hydrolysates and their potential for nutraceutical and functional foods. *Food Chem. Adv*, 2(May), 100297. <https://doi.org/10.1016/j.focha.2023.100297>
- Kapoor, M. P., Sugita, M., Fukuzawa, Y., Timm, D., & Ozeki, M. (2021). Green Tea Catechin Association with Ultraviolet. *Molecules*, 26(3702), 1–20. <https://doi.org/10.3390/molecules26123702>

- Kastanis, G. J., Santana-Quintero, L. V., Sanchez-Leon, M., Lomonaco, S., Brown, E. W., & Allard, M. W. (2019). In-depth comparative analysis of Illumina® MiSeq run metrics: Development of a wet-lab quality assessment tool. *Molecular Ecology Resources*, 19(2), 377–387. <https://doi.org/10.1111/1755-0998.12973>
- Kauffmann, A. C., & Castro, V. S. (2023). Phenolic Compounds in Bacterial Inactivation: A Perspective from Brazil. *Antibiotics*, 12(4), 1–24. <https://doi.org/10.3390/antibiotics12040645>
- Kaur, H., Jamir, I., Yangzes, S., Ahmad, H., Agnihotri, S., Gupta, S., Ghosh, A., & Rudramurthy, S. (2024). Papulaspora equi keratitis in an infant. *Indian Journal of Medical Microbiology*, 52, 100748. <https://doi.org/10.1016/j.ijmmb.2024.100748>
- Khadiri, M., Boubaker, H., Lahmamsi, H., Taoussi, M., Ezzougari, R., Askarne, L., Farhaoui, A., Barka, E. A., & Lahlali, R. (2024). Challenges in apple preservation: Fungicide resistance and emerging biocontrols. *Physiological and Molecular Plant Pathology*, 129(October 2023), 102205. <https://doi.org/10.1016/j.pmpp.2023.102205>
- Khan, M. A. (2022). Pathogenic Bacteria and Medical Treatment Abstract. *Archives of Clinical Microbiology*, 13(8), 7–9. <https://doi.org/10.36648/1989-8436X.22.16.8.200>
- Khan, M. A. B., Hashim, M. J., King, J. K., Govender, R. D., Mustafa, H., & Kaabi, J. Al. (2020). Epidemiology of Type 2 Diabetes – Global Burden of Disease and Forecasted Trends. *Journal of Epidemiology and Global Health*, 10(1), 107–111. <https://doi.org/10.3109/9780849379581-6>
- Khan, M., & Shamim, S. (2022). Understanding the Mechanism of Antimicrobial Resistance and Pathogenesis of Salmonella enterica Serovar Typhi. *Microorganisms*, 10(10). <https://doi.org/10.3390/microorganisms10102006>
- Khan, R., Anwar, F., & Ghazali, F. M. (2024). A comprehensive review of mycotoxins: Toxicology, detection, and effective mitigation approaches. *Heliyon*, 10(8), e28361. <https://doi.org/10.1016/j.heliyon.2024.e28361>
- Kheir, E. A. M., Elhussein, A. A., & Yagi, M. (2023). endophytic fungi isolates inhabited *Acacia nilotica*. 4–9.
- Kim, Y., & Kim, Y. (2024). Synthesis of Penta- and Tetrasubstituted Quercetins and Their Fluorescent Properties. *European Journal of Organic Chemistry*, 27(2), e202300807. <https://doi.org/https://doi.org/10.1002/ejoc.202300807>
- Kirit, H. A., Bollback, J. P., & Lagator, M. (2022). The Role of the Environment in Horizontal Gene Transfer. *Molecular Biology and Evolution*, 39(11), 1–11. <https://doi.org/10.1093/molbev/msac220>
- Ko, Y. C., Choi, H. S., Liu, R., & Lee, D. S. (2021). Physalin a, 13,14-seco-16, 24-

- cyclo-steroid, inhibits stemness of breast cancer cells by regulation of hedgehog signaling pathway and yes-associated protein 1 (Yap1). *International Journal of Molecular Sciences*, 22(16). <https://doi.org/10.3390/ijms22168718>
- Kozlov, A. V., Javadov, S., & Sommer, N. (2024). Cellular ROS and Antioxidants: Physiological and Pathological Role. *Antioxidants*, 13(5), 1–11. <https://doi.org/10.3390/antiox13050602>
- Krawczyk, S. J., Leśniczak-Staszak, M., Gowin, E., & Szaflarski, W. (2024). Mechanistic Insights into Clinically Relevant Ribosome-Targeting Antibiotics. *Biomolecules*, 14(10), 1–13. <https://doi.org/10.3390/biom14101263>
- Kremer, M. L. (2023). Initial Steps in the Reaction of H₂O₂ with Fe²⁺ and Fe³⁺ Ions: Inconsistency in the Free Radical Theory. *Reactions*, 4(1), 171–175. <https://doi.org/10.3390/reactions4010010>
- Kruk, J., Aboul-Enein, H. Y., Kładna, A., & Bowser, J. E. (2019). Oxidative stress in biological systems and its relation with pathophysiological functions: the effect of physical activity on cellular redox homeostasis. *Free Radical Research*, 53(5), 497–521. <https://doi.org/10.1080/10715762.2019.1612059>
- Krzyżek, P. (2019). Challenges and limitations of anti-quorum sensing therapies. *Frontiers in Microbiology*, 10(OCT). <https://doi.org/10.3389/fmicb.2019.02473>
- Kubayi, S., Makola, R. T., & Dithebe, K. (2025). Exploring the Antimicrobial, Antioxidant and Extracellular Enzymatic Activities of Culturable Endophytic Fungi Isolated from the Leaves of *Kirkia acuminata* Oliv. *Microorganisms*, 13(3), 1–17. <https://doi.org/10.3390/microorganisms13030692>
- Kumar, N., & Goel, N. (2019). Phenolic acids: Natural versatile molecules with promising therapeutic applications. *Biotechnology Reports*, 24, e00370. <https://doi.org/10.1016/j.btre.2019.e00370>
- Kumar, S., Abedin, M. M., Singh, A. K., & Das, S. (2020). Role of Phenolic Compounds in Plant-Defensive Mechanisms. *Plant Phenolics in Sustainable Agriculture*, 1, 1–594. <https://doi.org/10.1007/978-981-15-4890-1>
- Kumar, V., Kumar, H., Vishal, V., & Lal, S. (2023). Studies on the morphology, phylogeny, and bioremediation potential of *Penicillium citrinum* and *Paecilomyces variotii* (Eurotiales) from oil-contaminated areas. *Archives of Microbiology*, 205(1), 50. <https://doi.org/10.1007/s00203-022-03383-x>
- Kumara, P., K, S., & Kumar B, A. (2018). Determination of DPPH Free Radical Scavenging Activity by RP-HPLC, Rapid Sensitive Method for the Screening of Berry Fruit Juice Freeze Dried Extract. *Natural Products Chemistry & Research*, 06(05). <https://doi.org/10.4172/2329-6836.1000341>

- Kumari, P., Deepa, N., Trivedi, P. K., Singh, B. K., Srivastava, V., & Singh, A. (2023). Plants and endophytes interaction: a “secret wedlock” for sustainable biosynthesis of pharmaceutically important secondary metabolites. *Microbial Cell Factories*, 22(1), 1–19. <https://doi.org/10.1186/s12934-023-02234-8>
- Kuru, E., Radkov, A., Meng, X., Egan, A., Alvarez, L., Dowson, A., Booher, G., Breukink, E., Roper, D. I., Cava, F., Vollmer, W., Brun, Y., & Vannieuwenhze, M. S. (2019). Mechanisms of Incorporation for D -Amino Acid Probes That Target Peptidoglycan Biosynthesis. *ACS Chemical Biology*, 14(12), 2745–2756. <https://doi.org/10.1021/acscchembio.9b00664>
- Kusumah, D., Wakui, M., Murakami, M., Xie, X., Yukihiro, K., & Maeda, I. (2020). Linoleic acid, α -linolenic acid, and monolinolenins as antibacterial substances in the heat-processed soybean fermented with *Rhizopus oligosporus*. *Bioscience, Biotechnology and Biochemistry*, 84(6), 1285–1290. <https://doi.org/10.1080/09168451.2020.1731299>
- Kuźniar, A., Kruczyńska, A., Włodarczyk, K., Vangronsveld, J., & Wolińska, A. (2025). Endophytes as Permanent or Temporal Inhabitants of Different Ecological Niches in Sustainable Agriculture. *Applied Sciences (Switzerland)*, 15(3), 1–29. <https://doi.org/10.3390/app15031253>
- Lakey-Beitia, J., Burillo, A. M., La Penna, G., Hegde, M. L., & Rao, K. S. (2021). Polyphenols as Potential Metal Chelation Compounds against Alzheimer's Disease. *Journal of Alzheimer's Disease*, 82(s1), S335–S357. <https://doi.org/10.3233/JAD-200185>
- Langer, P. (2021). Domino Reactions of Chromones with Heterocyclic Enamines. *Synlett*, 33(03), 207–223. <https://doi.org/10.1055/s-0040-1719830>
- Lautenschläger, T., Rüggeberg, M., Noack, N., Bunk, K., Mawunu, M., Speck, T., & Neinhuis, C. (2020). Functional principles of baobab fruit pedicels - Anatomy and biomechanics. *Annals of Botany*, 126(7), 1215–1223. <https://doi.org/10.1093/aob/mcaa149>
- Li, Q. xi, Lin, F. cheng, & Su, Z. zhu. (2025). Endophytic fungi—Big player in plant-microbe symbiosis. *Current Plant Biology*, 42(November 2024), 100481. <https://doi.org/10.1016/j.cpb.2025.100481>
- Li, W., Chen, H., Xu, B., Wang, Y., Zhang, C., Cao, Y., & Xing, X. (2023). Research progress on classification, sources and functions of dietary polyphenols for prevention and treatment of chronic diseases. *Journal of Future Foods*, 3(4), 289–305. <https://doi.org/10.1016/j.jfutfo.2023.03.001>
- Li, W., Koshkarov, A., & Tahiri, N. (2024). Comparison of phylogenetic trees defined on different but mutually overlapping sets of taxa: A review. *Ecology and Evolution*, 14(8), 1–16. <https://doi.org/10.1002/ece3.70054>
- Li, X. Q., Xu, K., Liu, X. M., & Zhang, P. (2020). A Systematic Review on

- Secondary Metabolites of *Paecilomyces* Species: Chemical Diversity and Biological Activity. *Planta Medica*, 86(12), 805–821. <https://doi.org/10.1055/a-1196-1906>
- Li, Z., Wen, W., Qin, M., He, Y., Xu, D., & Li, L. (2022). Biosynthetic Mechanisms of Secondary Metabolites Promoted by the Interaction Between Endophytes and Plant Hosts. *Frontiers in Microbiology*, 13(July). <https://doi.org/10.3389/fmicb.2022.928967>
- Li, Z., Xiong, K., Wen, W., Li, L., & Xu, D. (2023). Functional Endophytes Regulating Plant Secondary Metabolism: Current Status, Prospects and Applications. *International Journal of Molecular Sciences*, 24(2). <https://doi.org/10.3390/ijms24021153>
- Liang, Y., Liang, L., Shi, R., Luo, R., Yue, Y., Yu, J., Wang, X., Lin, J., Zhou, T., Yang, M., Zhong, L., Wang, Y., & Shu, Z. (2024). Genus *Physalis* L.: A review of resources and cultivation, chemical composition, pharmacological effects and applications. *Journal of Ethnopharmacology*, 324(August 2023), 117736. <https://doi.org/10.1016/j.jep.2024.117736>
- Lin, Y.-H., Hsiao, Y.-H., Ng, K.-L., Kuo, Y.-H., Lim, Y.-P., & Hsieh, W.-T. (2020). Physalin A attenuates inflammation through down-regulating c-Jun NH2 kinase phosphorylation/Activator Protein 1 activation and up-regulating the antioxidant activity. *Toxicology and Applied Pharmacology*, 402, 115115. <https://doi.org/https://doi.org/10.1016/j.taap.2020.115115>
- Liu, H. Y., Luo, D., Huang, H. L., & Yang, Q. (2024). Two new species of *Diaporthe* (Diaporthaceae, Diaporthales) associated with *Camellia oleifera* leaf spot disease in Hainan Province, China. *MycoKeys*, 102(2024), 225–243. <https://doi.org/10.3897/mycokeys.102.113412>
- Liu, W., Zhang, H., Wan, H., Hou, J., Lee, D., Xu, J., & Guo, Y. (2022). Anti-inflammatory withanolides from the aerial parts of *Physalis minima*. *Phytochemistry*, 202, 113301. <https://doi.org/https://doi.org/10.1016/j.phytochem.2022.113301>
- Liu, X., Zhou, Z. Y., Cui, J. L., Wang, M. L., & Wang, J. H. (2021). Biotransformation ability of endophytic fungi: from species evolution to industrial applications. *Applied Microbiology and Biotechnology*, 105(19), 7095–7113. <https://doi.org/10.1007/s00253-021-11554-x>
- Lobatto, V. L., García, M. E., Nicotra, V. E., Orozco, C. I., & Casero, C. N. (2023). Antibacterial activity of withanolides and their structure-activity relationship. *Steroids*, 199(June), 109297. <https://doi.org/10.1016/j.steroids.2023.109297>
- Lobiuc, A., Pavăl, N. E., Mangalagiu, I. I., Gheorghită, R., Teliban, G. C., Amăriucăi-Mantu, D., & Stoleru, V. (2023). Future Antimicrobials: Natural and Functionalized Phenolics. *Molecules*, 28(3). <https://doi.org/10.3390/molecules28031114>

- Lombardi, N., Caira, S., Troise, A. D., Scaloni, A., Vitaglione, P., Vinale, F., Marra, R., Salzano, A. M., Lorito, M., & Woo, S. L. (2020). Trichoderma Applications on Strawberry Plants Modulate the Physiological Processes Positively Affecting Fruit Production and Quality. *Frontiers in Microbiology*, 11(July), 1–17. <https://doi.org/10.3389/fmicb.2020.01364>
- Lončarić, A., Šarkanj, B., Gotal, A. M., Kovač, M., Nevistić, A., Fruk, G., Babojelić, M. S., Babić, J., Miličević, B., & Kovač, T. (2021). Penicillium expansum impact and patulin accumulation on conventional and traditional apple cultivars. *Toxins*, 13(10). <https://doi.org/10.3390/toxins13100703>
- Lu, L., Karunarathna, S. C., Hyde, K. D., Suwannarach, N., Elgorban, A. M., Stephenson, S. L., Al-Rejaie, S., Jayawardena, R. S., & Tibpromma, S. (2022). Endophytic Fungi Associated with Coffee Leaves in China Exhibited In Vitro Antagonism against Fungal and Bacterial Pathogens. *Journal of Fungi*, 8(7), 1–21. <https://doi.org/10.3390/jof8070698>
- Lü, Z. W., Liu, H. Y., Wang, C. L., Chen, X., Huang, Y. X., Zhang, M. M., Huang, Q. L., & Zhang, G. F. (2023). Isolation of endophytic fungi from Cotoneaster multiflorus and screening of drought-tolerant fungi and evaluation of their growth-promoting effects. *Frontiers in Microbiology*, 14. <https://doi.org/10.3389/fmicb.2023.1267404>
- Lucini, F., de Andrade, G. A. K., Victoria, F. de C., & de Albuquerque, M. P. (2022). Impact of the Temperature in Endophytic Ascomycota Isolated from Antarctic Hair-Grass. *Life*, 12(10). <https://doi.org/10.3390/life12101501>
- Luo, M., Guo, W., Zhao, M., Manawasinghe, I. S., Guarnaccia, V., Liu, J., Hyde, K. D., Dong, Z., & You, C. (2022). Endophytic Diaporthe Associated with Morinda officinalis in China. *Journal of Fungi*, 8(8). <https://doi.org/10.3390/jof8080806>
- MacDonald, B. L., Stalla, D., He, X., Rahemtulla, F., Emerson, D., Dube, P. A., Maschmann, M. R., Klesner, C. E., & White, T. A. (2019). Hunter-Gatherers Harvested and Heated Microbial Biogenic Iron Oxides to Produce Rock Art Pigment. *Scientific Reports*, 9(1), 1–13. <https://doi.org/10.1038/s41598-019-53564-w>
- Magagula, M., Motaung, T. E., Mbita, Z., & Dithebe, K. (2025). Investigating the antimicrobial and anticancer potential of culturable fungal endophytes isolated from the stems of Kirkia acuminata Oliv. *BMC Microbiology*, 25(1). <https://doi.org/10.1186/s12866-025-03964-y>
- Mahadevakumar, S., Joy, J., Bhanu, L. S. M., Sharvani, K. A., Sowmya, R., & Raj, S. N. (2023). *Disease Note*. January, 2023.
- Mahardhika, W. A., Isworo Rukmi, M. G., & Pujiyanto, S. (2021). Isolasi kapang endofit dari tanaman ciplukan (Physalis angulata L.) dan potensi antibakteri terhadap Escherichia coli dan Staphylococcus aureus. *NICHE J Trop*

- Bio*, 4(1), 33–39. <https://ejournal2.undip.ac.id/index.php/niche>
- Mahfouz, M. (2023). Cold Plasma in The Control of *Paecilomyces Variotii* Isolated from Fruit Juice Processing Lines. *Anais Do 15° SLACAN - Simpósio Latino Americano de Ciência de Alimentos e Nutrição*.
- Mahklouf, M. H. (2019). The first record of *Physalis angulata* L. (Solanaceae) for the flora of Libya . *Biodiversity Research and Conservation*, 53(1), 67–71. <https://doi.org/10.2478/biorc-2019-0004>
- MAHMUD, Y., ROMANTIS, C., & ZAM, S. I. (2020). EFEKTIVITAS *Trichoderma virens* DALAM MENGENDALIKAN *Ganoderma boninense* DI PRE NURSERY KELAPA SAWIT PADA MEDIUM GAMBUT. *Jurnal Agroteknologi*, 11(1), 11. <https://doi.org/10.24014/ja.v11i1.8751>
- Makarewicz, M., Drożdż, I., Tarko, T., & Duda-Chodak, A. (2021). The interactions between polyphenols and microorganisms, especially gut microbiota. *Antioxidants*, 10(2), 1–70. <https://doi.org/10.3390/antiox10020188>
- Manichart, N., Laosinwattana, C., Somala, N., Teerarak, M., & Chotsaeng, N. (2023). Physiological mechanism of action and partial separation of herbicide–active compounds from the *Diaporthe* sp. extract on *Amaranthus tricolor* L. *Scientific Reports*, 13(1), 1–12. <https://doi.org/10.1038/s41598-023-46201-0>
- Manieri, T. M., Sensi, S. L., Squitti, R., & Cerchiaro, G. (2021). Structural effects of stabilization and complexation of a zinc-deficient superoxide dismutase. *Heliyon*, 7(1), e06100. <https://doi.org/10.1016/j.heliyon.2021.e06100>
- Manzar, N., Kashyap, A. S., Goutam, R. S., Rajawat, M. V. S., Sharma, P. K., Sharma, S. K., & Singh, H. V. (2022). *Trichoderma*: Advent of Versatile Biocontrol Agent, Its Secrets and Insights into Mechanism of Biocontrol Potential. *Sustainability (Switzerland)*, 14(19), 1–32. <https://doi.org/10.3390/su141912786>
- Mao, Z., Zhang, W., Wu, C., Feng, H., Peng, Y., Shahid, H., Cui, Z., Ding, P., & Shan, T. (2021). Diversity and antibacterial activity of fungal endophytes from *Eucalyptus exserta*. *BMC Microbiology*, 21(1), 1–12. <https://doi.org/10.1186/s12866-021-02229-8>
- Martin, M., Saparrat, M., Barrera, V., & Gasoni, L. (2019). Mini-review: *Cladorrhinum* species, their diversity and application in biotechnology. *Nova Hedwigia*, 108(3–4), 489–503. https://doi.org/10.1127/nova_hedwigia/2018/0516
- Martinelli, L., Redou, V., Cochereau, B., Delage, L., Hymery, N., Poirier, E., Le Meur, C., Le Foch, G., Cladiere, L., Mehiri, M., Demont-Caulet, N., & Meslet-Cladiere, L. (2020). Identification and Characterization of a New Type III Polyketide Synthase from a Marine Yeast, *Naganishia uzbekistanensis*.

- Marine Drugs*, 18(12). <https://doi.org/10.3390/md18120637>
- Martinvalet, D., & Walch, M. (2022). Editorial: The Role of Reactive Oxygen Species in Protective Immunity. *Frontiers in Immunology*, 12(January), 10–13. <https://doi.org/10.3389/fimmu.2021.832946>
- Masenga, S. K., Kabwe, L. S., Chakulya, M., & Kirabo, A. (2023). Mechanisms of Oxidative Stress in Metabolic Syndrome. *International Journal of Molecular Sciences*, 24(9). <https://doi.org/10.3390/ijms24097898>
- Matevosyan, L., Bazukyan, I., & Trchounian, A. (2019). Comparative analysis of the effect of Ca and Mg ions on antibacterial activity of lactic acid bacteria isolates and their associations depending on cultivation conditions. *AMB Express*, 9(1). <https://doi.org/10.1186/s13568-019-0758-9>
- Matsue, M., Mori, Y., Nagase, S., Sugiyama, Y., Hirano, R., Ogai, K., Ogura, K., Kurihara, S., & Okamoto, S. (2019). Measuring the Antimicrobial Activity of Lauric Acid against Various Bacteria in Human Gut Microbiota Using a New Method. *Cell Transplantation*, 28(12), 1528–1541. <https://doi.org/10.1177/0963689719881366>
- Matušinsky, P., Florová, V., Sedláková, B., Mlčoch, P., & Bleša, D. (2024). Colonization dynamic and distribution of the endophytic fungus *Microdochium bolleyi* in plants measured by qPCR. *PLoS ONE*, 19(1 January), 1–14. <https://doi.org/10.1371/journal.pone.0297633>
- Mazlan, N. W., Tate, R., Yusoff, Y. M., Clements, C., & Edrada-Ebel, R. (2020). Metabolomics-Guided Isolation of Anti-Trypanosomal Compounds from Endophytic Fungi of the Mangrove plant *Avicennia Lanata*. In *Current Medicinal Chemistry* (Vol. 27, Issue 11, pp. 1815–1835). <https://doi.org/http://dx.doi.org/10.2174/0929867326666190704130105>
- Mcmurry, J. (2023). *Organic Chemistry: 10th Edition*. OpenStax.
- Medina, S., Torres, M., Durán, Y., Ramírez, R., Herrera, J., & Ramírez, P. (2012). Degradación de tiocianato por hongos aislados de ambientes mineros y evaluación de su capacidad degradativa. *Revista Peruana de Biología*, 19(1), 81–88. <https://doi.org/10.15381/rpb.v19i1.791>
- Meira, C. S., Soares, J. W. C., dos Reis, B. P. Z. C., Pacheco, L. V., Santos, I. P., Silva, D. K. C., de Lacerda, J. C., Daltro, S. R. T., Guimarães, E. T., & Soares, M. B. P. (2022). Therapeutic Applications of Physalins: Powerful Natural Weapons. *Frontiers in Pharmacology*, 13(April), 1–14. <https://doi.org/10.3389/fphar.2022.864714>
- Mejía-Bautista, M. A., Cristóbal-Alejo, J., Tun-Suárez, J. M., & Reyes-Ramírez, A. (2016). Actividad in vitro de *Bacillus* spp. en la inhibición de crecimiento micelial de *Fusarium equiseti* y *Fusarium solani* aislado de chile habanero (*Capsicum chinense* Jacq.). *Agrociencia*, 50(8), 1123–1135.

- Meressa, E. W., & Tseha, B. A. (2024). Horizontal gene transfer - The hidden hazards of genetic engineering. *Journal of Life Science and Biomedicine*, 14(4), 99–108.
- Metwally, R. A., Abdelhameed, R. E., Soliman, S. A., & Al-Badwy, A. H. (2022). Potential use of beneficial fungal microorganisms and C-phycocyanin extract for enhancing seed germination, seedling growth and biochemical traits of *Solanum lycopersicum* L. *BMC Microbiology*, 22(1), 1–17. <https://doi.org/10.1186/s12866-022-02509-x>
- Mickymaray, S., Alfaiz, F. A., & Paramasivam, A. (2020). Efficacy and mechanisms of flavonoids against the emerging opportunistic nontuberculous mycobacteria. *Antibiotics*, 9(8), 1–34. <https://doi.org/10.3390/antibiotics9080450>
- Mikulska, P., Malinowska, M., Ignacyk, M., Szustowski, P., Nowak, J., Pesta, K., Szela, M., Szklanny, D., Judasz, E., Kaczmarek, G., Ejiohuo, O. P., Paczkowska-Walendowska, M., Gościński, A., & Cielecka-Piontek, J. (2023). Ashwagandha (*Withania somnifera*)—Current Research on the Health-Promoting Activities: A Narrative Review. *Pharmaceutics*, 15(4). <https://doi.org/10.3390/pharmaceutics15041057>
- Miyano, R., Matsuo, H., Mokudai, T., Noguchi, Y., Higo, M., Nonaka, K., Niwano, Y., Sunazuka, T., Shiomi, K., Takahashi, Y., Omura, S., & Nakashima, T. (2020). Trichothioneic acid, a new antioxidant compound produced by the fungal strain *Trichoderma virens* FKI-7573. *Journal of Bioscience and Bioengineering*, 129(4), 508–513. <https://doi.org/10.1016/j.jbiosc.2019.11.007>
- Mokoroane, K. T., Pillai, M. K., & Magama, S. (2020). 2,2-diphenyl-1-picrylhydrazyl (Dpph) radical scavenging activity of extracts from *aloiampelos striatula*. *Food Research*, 4(6), 2062–2066. [https://doi.org/10.26656/fr.2017.4\(6\).241](https://doi.org/10.26656/fr.2017.4(6).241)
- Monteiro Espíndola, K. M., Ferreira, R. G., Mosquera Narvaez, L. E., Rocha Silva Rosario, A. C., Machado Da Silva, A. H., Bispo Silva, A. G., Oliveira Vieira, A. P., & Chagas Monteiro, M. (2019). Chemical and pharmacological aspects of caffeic acid and its activity in hepatocarcinoma. *Frontiers in Oncology*, 9(JUN), 3–5. <https://doi.org/10.3389/fonc.2019.00541>
- Moraes, M. R. de, Ryan, S. M., Godoy, H. T., Thomas, A. L., Maia, J. G. S., Richards, K. M., Tran, K., & Smith, R. E. (2020). Phenolic Compounds and Metals in Some Edible Annonaceae Fruits. *Biological Trace Element Research*, 197(2), 676–682. <https://doi.org/10.1007/s12011-019-02005-w>
- Morales-Vargas, A. T., López-Ramírez, V., Álvarez-Mejía, C., & Vázquez-Martínez, J. (2024). Endophytic Fungi for Crops Adaptation to Abiotic Stresses. *Microorganisms*, 12(7). <https://doi.org/10.3390/microorganisms12071357>

- Moreno-Gavira, A., Diáñez, F., Sánchez-Montesinos, B., & Santos, M. (2020). *Paecilomyces variotii* as a plant-growth promoter in horticulture. *Agronomy*, 10(4). <https://doi.org/10.3390/AGRONOMY10040597>
- Moreno-Gavira, A., Huertas, V., Diáñez, F., Santos, M., & Sánchez-Montesinos, B. (2020). *Paecilomyces* and its importance in the biological control of agricultural pests and diseases. *Plants*, 9(12), 1–28. <https://doi.org/10.3390/plants9121746>
- Morsy, M., Ahmed, A., Awad, M. F., El-shenawy, F. S., Mohamed, A., & El-bondkly, A. (2021). Saudi Journal of Biological Sciences antiproliferative activities of extracellular L-asparaginase produced by *Fusarium equiseti* AHMF4. *Saudi Journal of Biological Sciences*, 28(4), 2540–2548. <https://doi.org/10.1016/j.sjbs.2021.01.058>
- Moubareck, C. A. (2020). Polymyxins and bacterial membranes: A review of antibacterial activity and mechanisms of resistance. *Membranes*, 10(8), 1–30. <https://doi.org/10.3390/membranes10080181>
- Moussa, Z., & Al-Mamary, M. A. (2021). *Antioxidant Activity: The Presence and Impact of Hydroxyl Groups in Small Molecules of Natural and Synthetic Origin* (V. Y. Waisundara (ed.)). IntechOpen. <https://doi.org/10.5772/intechopen.95616>
- Muhammad Abdul Kadar, N. N., Ahmad, F., Teoh, S. L., & Yahaya, M. F. (2021). Caffeic Acid on Metabolic Syndrome: A Review. *Molecules (Basel, Switzerland)*, 26(18), 1–14. <https://doi.org/10.3390/molecules26185490>
- Muhibuddin, A., Salsabila, S., & Sektiono, A. W. (2021). KEMAMPUAN ANTAGONIS *Tricoderma harzianum* TERHADAP BEBERAPA JAMUR PATOGEN PENYAKIT TANAMAN. *Agrosaintifika*, 4(1), 225–233. <https://doi.org/10.32764/agrosaintifika.v4i1.2371>
- Mulder, M., Verbon, A., Lous, J., Goessens, W., & Stricker, B. H. (2019). Use of other antimicrobial drugs is associated with trimethoprim resistance in patients with urinary tract infections caused by *E. coli*. *European Journal of Clinical Microbiology and Infectious Diseases*, 38(12), 2283–2290. <https://doi.org/10.1007/s10096-019-03672-2>
- Munawir, A. M., & Basir, A. A. (2023). The Activity of *Salmonella Typhii* in Milkfish (*Chanos chanos*) with Invitro. *International Journal of Applied Biology*, 7(1), 26–34.
- Munteanu, I. G., & Apetrei, C. (2021). Analytical methods used in determining antioxidant activity: A review. *International Journal of Molecular Sciences*, 22(7). <https://doi.org/10.3390/ijms22073380>
- Muronetz, V. I., Barinova, K., Kudryavtseva, S., Medvedeva, M., Melnikova, A., Sevostyanova, I., Semenyuk, P., Stroylova, Y., & Sova, M. (2020). Natural

- and synthetic derivatives of hydroxycinnamic acid modulating the pathological transformation of amyloidogenic proteins. *Molecules*, 25(20). <https://doi.org/10.3390/molecules25204647>
- Musa, M., Jan, F. G., Hamayun, M., Jan, G., Khan, S. A., Rehman, G., Ali, S., & Lee, I. J. (2023). An Endophytic Fungal Isolate *Paecilomyces lilacinus* Produces Bioactive Secondary Metabolites and Promotes Growth of *Solanum lycopersicum* under Heavy Metal Stress. *Agronomy*, 13(3). <https://doi.org/10.3390/agronomy13030883>
- Muteeb, G., Rehman, M. T., Shahwan, M., & Aatif, M. (2023). Origin of Antibiotics and Antibiotic Resistance, and Their Impacts on Drug Development: A Narrative Review. *Pharmaceuticals*, 16(11), 1–54. <https://doi.org/10.3390/ph16111615>
- Nabavi, S. M., Šamec, D., Tomczyk, M., Milella, L., Russo, D., Habtemariam, S., Suntar, I., Rastrelli, L., Daglia, M., Xiao, J., Giampieri, F., Battino, M., Sobarzo-Sanchez, E., Nabavi, S. F., Yousefi, B., Jeandet, P., Xu, S., & Shirooie, S. (2020). Flavonoid biosynthetic pathways in plants: Versatile targets for metabolic engineering. *Biotechnology Advances*, 38(June 2018), 107316. <https://doi.org/10.1016/j.biotechadv.2018.11.005>
- Nagel, R., Schmidt, A., & Peters, R. J. (2019). Isoprenyl diphosphate synthases: the chain length determining step in terpene biosynthesis. *Planta*, 249(1), 9–20. <https://doi.org/10.1007/s00425-018-3052-1>
- Naha, A., Kumar Miryala, S., Debroy, R., Ramaiah, S., & Anbarasu, A. (2020). Elucidating the multi-drug resistance mechanism of *Enterococcus faecalis* V583: A gene interaction network analysis. *Gene*, 748(April), 144704. <https://doi.org/10.1016/j.gene.2020.144704>
- Naik, B. S. (2019). Developments in taxol production through endophytic fungal biotechnology: a review. *Oriental Pharmacy and Experimental Medicine*, 19(1), 1–13. <https://doi.org/10.1007/s13596-018-0352-8>
- Najjar, A. A. (2025). Therapeutic Potential of Endophytic Microbes: Emphasizing Both Fungal and Bacterial Endophytes. *Applied Microbiology (Switzerland)*, 5(1). <https://doi.org/10.3390/applmicrobiol5010005>
- Nandika, D., Karlinasari, L., Arinana, A., Batubara, I., Sitanggang, P. S., Santoso, D., Witasari, L. D., Rachmayanti, Y., Firmansyah, D., Sudiana, I. K., & Hertanto, D. M. (2021). Chemical components of fungus comb from Indo-Malayan termite macrotermes gilvus hagen mound and its bioactivity against wood-staining fungi. *Forests*, 12(11), 1–17. <https://doi.org/10.3390/f12111591>
- Nasution, R., Muhabbah, N., Helwati, H., Bahi, M., Marianne, M., & Amna, U. (2020). Isolation of lupeol acetate from fruit peels of *Artocarpus camansi*. *Journal of Advanced Pharmaceutical Technology and Research*, 11(3), 148–

156. https://doi.org/10.4103/japtr.JAPTR_6_20

- Nazir, A., Puthuveetil, A. R., Hussain, F. H. N., Hamed, K. E., & Munawar, N. (2024). Endophytic fungi: nature's solution for antimicrobial resistance and sustainable agriculture. *Frontiers in Microbiology*, 15(December), 1–8. <https://doi.org/10.3389/fmicb.2024.1461504>
- Nemeth, V., & Pfliegerhaa, N. (2025). *Diarrhea*. 1–11.
- Ng uni, T., Thato, M., Raymond, D., Jeremy, K., & Burtram, C. F. (2015). Additive antibacterial activity of naringenin and antibiotic combinations against multidrug resistant *Staphylococcus aureus*. *African Journal of Microbiology Research*, 9(23), 1513–1518. <https://doi.org/10.5897/ajmr2015.7514>
- Nguyen, K.-N. H., Nguyen, N.-V. T., & Kim, K. H. (2021). Determination of phenolic acids and flavonoids in leaves, calyces, and fruits of *Physalis angulata* L. in Viet Nam. *Pharmacia*, 68(2), 501–509. <https://doi.org/https://doi.org/10.3897/pharmacia.68.e66044>
- Nguyen, T. H., Helm, B., Hettiarachchi, H., Caucci, S., & Krebs, P. (2020). Quantifying the information content of a water quality monitoring network using principal component analysis: A case study of the freiberger mulde river basin, Germany. *Water (Switzerland)*, 12(2). <https://doi.org/10.3390/w12020420>
- Nikolic, P., & Mudgil, P. (2023). The Cell Wall, Cell Membrane and Virulence Factors of *Staphylococcus aureus* and Their Role in Antibiotic Resistance. *Microorganisms*, 11(2). <https://doi.org/10.3390/microorganisms11020259>
- Nischitha, R., & Shivanna, M. B. (2022). Screening of secondary metabolites and antioxidant potential of endophytic fungus *Penicillium citrinum* and host *Digitaria bicornis* by spectrophotometric and electrochemical methods. *Archives of Microbiology*, 204(4), 206. <https://doi.org/10.1007/s00203-022-02795-z>
- Nitbani, F. O., Tjitda, P. J. P., Nitti, F., Jumina, J., & Detha, A. I. R. (2022). Antimicrobial Properties of Lauric Acid and Monolaurin in Virgin Coconut Oil: A Review. *ChemBioEng Reviews*, 9(5), 442–461. <https://doi.org/https://doi.org/10.1002/cben.202100050>
- Nolfi-Donagan, D., Braganza, A., & Shiva, S. (2020). Mitochondrial electron transport chain: Oxidative phosphorylation, oxidant production, and methods of measurement. *Redox Biology*, 37, 101674. <https://doi.org/10.1016/j.redox.2020.101674>
- Nongthombam, K. S., Mutum, S. S., & Pandey, R. R. (2024). Distribution, Diversity and Biochemical Analysis of Endophytic Fungi Associated with *Chromolaena odorata*. *Journal of Applied Life Sciences International*, 27(1), 17–29. <https://doi.org/10.9734/jalsi/2024/v27i1634>

- Nouri, A., Heibati, F., & Heidarian, E. (2021). Gallic acid exerts anti-inflammatory, anti-oxidative stress, and nephroprotective effects against paraquat-induced renal injury in male rats. *Naunyn-Schmiedeberg's Archives of Pharmacology*, 394(1), 1–9. <https://doi.org/10.1007/s00210-020-01931-0>
- Novitasari, A., Rohmawaty, E., & Rosdianto, A. M. (2024). *Physalis angulata* Linn. as a medicinal plant (Review). *Biomed Rep*, 20(3). <https://doi.org/10.3892/br.2024.1735>
- Nuaimy, M. A. A. A., & Hawar, S. N. (2024). Antagonistic Activity of Endophytic Fungi Isolated From Aloe Vera Leaves Against Some Plant Pathogenic Fungi. *Iraqi Journal of Agricultural Sciences*, 55(Special Issue), 63–79. <https://doi.org/10.36103/ijas.v55iSpecial.1886>
- Nuez, L., Durand, S., Melelli, A., Berrin, J. G., Haon, M., Drula, E., Beaugrand, J., D'Arras, P., Bourmaud, A., & Baley, C. (2022). Exploring the impact of Verticillium wilt disease on the mechanical properties of elementary flax (*Linum usitatissimum* L.) fibres. *Industrial Crops and Products*, 182(March), 114900. <https://doi.org/10.1016/j.indcrop.2022.114900>
- Nuraini, F. R., Setyaningsih, R., & Susilowati, A. (2019). Antioxidant activity of bioactive compound produced by endophytic fungi isolated from endemic plant of South Kalimantan *Mangifera casturi* Kosterm. *AIP Conference Proceedings*, 2120. <https://doi.org/10.1063/1.5115751>
- Obidiegwu, O. C., Adione, N. M., Onyeka, I. P., & Eze, P. M. (2024). *Antioxidant and cytotoxicity evaluation of the Endophytic Fungus , Fusarium equiseti , isolated from the leaves of Ocimum gratissimum* *Antioxidant and cytotoxicity evaluation of the Endophytic Fungus , Fusarium equiseti , isolated from the leaves of Ocimum*. May. <https://doi.org/10.30574/msarr.2024.10.2.0009>
- Odelade, K. A., & Babalola, O. O. (2019). Bacteria, fungi and archaea domains in rhizospheric soil and their effects in enhancing agricultural productivity. *International Journal of Environmental Research and Public Health*, 16(20). <https://doi.org/10.3390/ijerph16203873>
- Oduşina, B. O., & Onocha, P. A. (2022). A new squalene derivative from *Physalis angulata* L. (Solanaceae). *Natural Product Research*, 36(8), 2154–2157. <https://doi.org/10.1080/14786419.2020.1844691>
- Oktiansyah, R., Elfita, E., Widjajanti, H., Setiawan, A., Mardiyanto, M., & Nasution, S. S. A. (2023). Antioxidant and Antibacterial Activity of Endophytic Fungi Isolated from The Leaves of Sungkai (*Peronema canescens*). *Tropical Journal of Natural Product Research*, 7(3), 2596–2604. <https://doi.org/10.26538/tjnpr/v7i3.20>
- Oktiansyah, R., Widjajanti, H., Setiawan, A., Nasution, S. S. A., Mardiyanto, & Elfita. (2023). Antibacterial and Antioxidant Activity of Endophytic Fungi Extract Isolated from Leaves of Sungkai (*Peronema canescens*). *Science and*

- Technology Indonesia*, 8(2), 170–177.
<https://doi.org/10.26554/sti.2023.8.2.170-177>
- Olchowik-Grabarek, E., Sękowski, S., Kwiatek, A., Płaczekiewicz, J., Abdulladjanova, N., Shlyonsky, V., Swiecicka, I., & Zamaraeva, M. (2022). The Structural Changes in the Membranes of *Staphylococcus aureus* Caused by Hydrolysable Tannins Witness Their Antibacterial Activity. *Membranes*, 12(11). <https://doi.org/10.3390/membranes12111124>
- Omeike, S. O., Kareem, S. O., & Lasisi, A. A. (2019). Potential antibiotic-producing fungal strains isolated from pharmaceutical waste sludge. *Beni-Suef University Journal of Basic and Applied Sciences*, 8(1). <https://doi.org/10.1186/s43088-019-0026-8>
- Ortega, H. E., Torres-Mendoza, D., & Cubilla-Ríos, L. (2025). Antibacterial Compounds Isolated from Endophytic Fungi Reported from 2021 to 2024. *Antibiotics*, 14(7), 644. <https://doi.org/10.3390/antibiotics14070644>
- Owusu, E. O., Kwoseh, C. K., Osekere, E., & Akromah, R. (2023). Incidence, diversity and distribution of *Fusarium wilt* pathogens of eggplant in some major growing areas of Ashanti, Eastern and Volta Regions of Ghana. *Ghana Journal of Agricultural Science*, 58(1), 118–127. <https://doi.org/10.4314/gjas.v58i1.11>
- Pachauri, S., Zaid, R., Sherkhane, P. D., Easa, J., Viterbo, A., & Chet, I. (2023). Comparative Phenotypic, Genomic, and Transcriptomic Analyses of Two Contrasting Strains of the Plant Beneficial Fungus *Trichoderma virens*. *American Society For Microbiology*, 11.
- Palupi, K. D., Ilyas, M., & Agusta, A. (2021). Endophytic fungi inhabiting *Physalis angulata* L. plant: Diversity, antioxidant, and antibacterial activities of their ethyl acetate extracts. *J Basic Clin Physiol Pharmacol*, 32(4), 823–829. <https://doi.org/10.1515/jbcpp-2020-0479>
- Pan, F., Yang, N., Zhu, X., Yu, C., Jiang, M., Jiang, Y., Liu, S., Wu, W., & Liu, Y. (2023). Discovery of a Natural Hybrid Polyketide Produced by Endophytic *Cladosporium sphaerospermum* for Biocontrol of Phytopathogenic Fungus *Botrytis cinerea*. *Journal of Agricultural and Food Chemistry*, 71(32), 12190–12202. <https://doi.org/10.1021/acs.jafc.3c02408>
- Pancu, D. F., Scurtu, A., Macasoi, I. G., Marti, D., Mioc, M., Soica, C., Coricovac, D., Horhat, D., Poenaru, M., & Dehelean, C. (2021). Antibiotics: Conventional therapy and natural compounds with antibacterial activity-a pharmacotoxicological screening. *Antibiotics*, 10(4). <https://doi.org/10.3390/antibiotics10040401>
- Pandey, S., Doo, H., Keum, G. B., Kim, E. S., Kwak, J., Ryu, S., Choi, Y., Kang, J., Kim, S., Lee, N. R., Oh, K. K., Lee, J. H., & Kim, H. B. (2024). Antibiotic resistance in livestock, environment and humans: One Health perspective.

- Journal of Animal Science and Technology*, 62(2), 266–278.
<https://doi.org/10.5187/JAST.2023.E129>
- Papilaya, F. B., Wahyuningsih, M. S. H., & Syarif, R. A. (2025). Active fraction of Ciplukan (*Physalis angulata* L.) increases phosphorylation on IRS-1 Tyr-612 and Akt Ser-473 in 24-hour incubation time. *Journal of Applied Pharmaceutical Science*, 15(5), 121–128.
<https://doi.org/10.7324/JAPS.2025.225043>
- Pasin, T. M., Moreira, E. A., Benassi, V. M., Spencer, P. V.D., Peres, N. T. A., Cereia, M., & Polizeli, M. de L. T. M. (2022). Effects of Ultraviolet Exposure on the Tropical Fungi *Aspergillus carbonarius* and *Aspergillus japonicus*: Survival, Amylase Production, and Thermostability. *Tropical Conservation Science*, 15, 1–7. <https://doi.org/10.1177/19400829221092638>
- Patangia, D. V., Anthony Ryan, C., Dempsey, E., Paul Ross, R., & Stanton, C. (2022). Impact of antibiotics on the human microbiome and consequences for host health. *MicrobiologyOpen*, 11(1), 1–23.
<https://doi.org/10.1002/mbo3.1260>
- Patel, K., Bunachita, S., Agarwal, A. A., Bhamidipati, A., & Patel, U. K. (2021). A Comprehensive Overview of Antibiotic Selection and the Factors Affecting It. *Cureus*, 13(3). <https://doi.org/10.7759/cureus.13925>
- Patel, P., Wermuth, H. R., Calhoun, C., & Hall, G. A. (2023). Antibiotics. In *NCBI Bookshelf. A service of the National Library of Medicine, National Institute of Health*. (pp. 3–5).
- Pazo-Cepeda, M. V., Aspromonte, S. G., & Alonso, E. (2021). Extraction of ferulic acid and feruloylated arabinoxylo-oligosaccharides from wheat bran using pressurized hot water. *Food Bioscience*, 44(PA), 101374.
<https://doi.org/10.1016/j.fbio.2021.101374>
- Pereira-Dias, L., Oliveira-Pinto, P. R., Fernandes, J. O., Regalado, L., Mendes, R., Teixeira, C., Mariz-Ponte, N., Gomes, P., & Santos, C. (2023). Peptaibiotics: Harnessing the potential of microbial secondary metabolites for mitigation of plant pathogens. *Biotechnology Advances*, 68(July).
<https://doi.org/10.1016/j.biotechadv.2023.108223>
- Periferakis, A. T., Periferakis, A., Periferakis, K., Caruntu, A., Badarau, I. A., Savulescu-Fiedler, I., Scheau, C., & Caruntu, C. (2023). Antimicrobial Properties of Capsaicin: Available Data and Future Research Perspectives. *Nutrients*, 15(19), 1–27. <https://doi.org/10.3390/nu15194097>
- Permana, R. C. E., Habibi, M., & Gunawan, E. (2021). Jamur *Paecilomyces* dari Leang Pettae di kawasan karst Maros dan saran pelestarian gambar cadasnya. *Berkala Arkeologi*, 41(1), 1–16. <https://doi.org/10.30883/jba.v41i1.602>
- Phillips, A. J. L., Alves, A., Abdollahzadeh, J., Slippers, B., Wingfield, M. J.,

- Groenewald, J. Z., & Crous, P. W. (2013). The Botryosphaeriaceae: Genera and species known from culture. *Studies in Mycology*, 76, 51–167. <https://doi.org/10.3114/sim0021>
- Pillai, J. R., Wali, A. F., Menezes, G. A., Rehman, M. U., Wani, T. A., Arafah, A., Zargar, S., & Mir, T. M. (2022). Chemical Composition Analysis, Cytotoxic, Antimicrobial and Antioxidant Activities of *Physalis angulata* L.: A Comparative Study of Leaves and Fruit. *Molecules*, 27(5). <https://doi.org/10.3390/molecules27051480>
- Plunkert, M. L., Martínez-Gómez, J., Madrigal, Y., Hernández, A. I., & Tribble, C. M. (2024). Tuber, or not tuber: Molecular and morphological basis of underground storage organ development. *Current Opinion in Plant Biology*, 80, 102544. <https://doi.org/10.1016/j.pbi.2024.102544>
- Podbielska, M., Kus-Li Skiewicz, M., Jagusztyn, B., Piechowicz, B., Sadło, S., Słowik-Borowiec, M., Twaru zek, M., & Szpyrka, E. (2020). Influence of *Bacillus subtilis* and *Trichoderma harzianum* on penthiopyrad degradation under laboratory and field studies. *Molecules*, 25(6), 1–16. <https://doi.org/10.3390/molecules25061421>
- Prachya, S., Wattanasiri, C., & Pansanit, A. (2020). Cytotoxic endophyte from *Camellia oleifera* Abel - *Penicillium chermesinum* NR121310. *Mycology*, 11(4), 316–321. <https://doi.org/10.1080/21501203.2019.1705928>
- Prajapati, C., Rai, S. N., Singh, A. K., Chopade, B. A., Singh, Y., Singh, S. K., Haque, S., Prieto, M. A., & Ashraf, G. M. (2025). An Update of Fungal Endophyte Diversity and Strategies for Augmenting Therapeutic Potential of their Potent Metabolites: Recent Advancement. In *Applied Biochemistry and Biotechnology* (Vol. 197, Issue 5). Springer US. <https://doi.org/10.1007/s12010-024-05098-9>
- Purbaya, S. (2023). Aktivitas Antioksidan Senyawa 6-propil-5,6-dihidro-2h-piran-2-on dari Jamur Endofit *Fusarium oxysporum* Akar *Kalanchoe millotii*. *Jurnal Kartika Kimia*, 6(2), 109–115. <https://doi.org/10.26874/jkk.v6i2.211>
- Purbaya, S., Sinaga, S. E., Safriansyah, W., Wulandari, A. P., Harneti, D., Mulyani, Y., Azhari, A., Farabi, K., Abdullah, F. F., & Supratman, U. (2024). Chemical constituents of *Lasioidiplodia theobromae* BPPCA 144, an endophytic fungus, isolated from *Aglaia argentea* Blume. *Journal of Biologically Active Products from Nature*, 14(4), 443–454. <https://doi.org/10.1080/22311866.2024.2399718>
- Pyrzynska, K. (2024). Ferulic Acid—A Brief Review of Its Extraction, Bioavailability and Biological Activity. *Separations*, 11(7). <https://doi.org/10.3390/separations11070204>
- Qin, J., Lyu, A., Zhang, Q. hua, Yang, L., Zhang, J., Wu, M. de, & Li, G. qing. (2019). Strain identification and metabolites isolation of *Aspergillus capensis*

- CanS-34A from *Brassica napus*. *Molecular Biology Reports*, 46(3), 3451–3460. <https://doi.org/10.1007/s11033-019-04808-5>
- Rachmy, S., Anna, J., & Tambunan, M. (2021). *Pengaruh Pemberian Ekstrak Fusarium equiseti terhadap Hewan Mus musculus dengan Pendedahan Akut Melalui Subkutan*. 45(283), 31–37.
- Radzi, N. C., Kamaruzaman, M. F. M., Mustapa, A. N., & Yunus, M. A. C. (2020). Extraction of Essential Oil from Indigenous Herb *Physalis angulata* Linn using Microwave Assisted Extraction. *Journal of Advanced Research in Fluid Mechanics and Thermal Sciences*, 72(1), 138–147. <https://doi.org/10.37934/arfmts.72.1.138147>
- Rahaman, M. M., Hossain, R., Herrera-Bravo, J., Islam, M. T., Atolani, O., Adeyemi, O. S., Owolodun, O. A., Kambizi, L., Daştan, S. D., Calina, D., & Sharifi-Rad, J. (2023). Natural antioxidants from some fruits, seeds, foods, natural products, and associated health benefits: An update. *Food Science and Nutrition*, 11(4), 1657–1670. <https://doi.org/10.1002/fsn3.3217>
- Rahman, I. (2019). Resistensi Antibiotik terhadap *Salmonella typhi* pada Penyakit Demam Tifoid di Kota Makassar. *Kieraha Medical Journal*, 1(2). <https://doi.org/10.33387/kmj.v1i2.1699>
- Rajesh, U., & Dhanaraj, S. (2023). A critical review on quercetin bioflavonoid and its derivatives: Scope, synthesis, and biological applications with future prospects. *Arabian Journal of Chemistry*, 16(8), 104881. <https://doi.org/10.1016/j.arabjc.2023.104881>
- Ramadhani, S., Khairullah, A. R., Effendi, M. H., Sukmanadi, M., Tyasningsih, W., Millannia, S. K., Afnani, D. A., Moses, I. B., Thoriq Ihza Farizqi, M., & Kaben, S. E. (2024). Detection of the methicillin-resistant *Staphylococcus aureus* gene encoding *mecA* in nasal swabs of cats in Surabaya, Indonesia. *Open Veterinary Journal*, 14(11), 3026–3036. <https://doi.org/10.5455/OVJ.2024.v14.i11.31>
- Ramírez-Mendoza, A. A., Ramírez-Herrera, M. A., Cortez-Álvarez, C. R., Nery-Flores, S. D., Tejeda-Martínez, A. R., Romero-Prado, M. M. de J., & Mendoza-Magaña, M. L. (2022). Curcumin Modifies the Activity of Plasmatic Antioxidant Enzymes and the Hippocampal Oxidative Profile in Rats upon Acute and Chronic Exposure to Ozone. *Molecules*, 27(14). <https://doi.org/10.3390/molecules27144531>
- Rasmi, A. H., Ahmed, E. F., Darwish, A. M. A., & Gad, G. F. M. (2022). Virulence genes distributed among *Staphylococcus aureus* causing wound infections and their correlation to antibiotic resistance. *BMC Infectious Diseases*, 22(1), 1–12. <https://doi.org/10.1186/s12879-022-07624-8>
- Rasooly, R., Choi, H. Y., Do, P., Morroni, G., Brescini, L., Cirioni, O., Giacometti, A., & Apostolidis, E. (2020). whISOBAX™ inhibits bacterial pathogenesis

- and enhances the effect of antibiotics. *Antibiotics*, 9(5). <https://doi.org/10.3390/antibiotics9050264>
- Rastogi, A. K., Taterh, S., & Kumar, B. S. (2023). Dimensionality Reduction Algorithms in Machine Learning: A Theoretical and Experimental Comparison†. *Engineering Proceedings*, 59(1), 1–9. <https://doi.org/10.3390/engproc2023059082>
- Rauf, A., Imran, M., Abu-Izneid, T., Iahtisham-Ul-Haq, Patel, S., Pan, X., Naz, S., Sanches Silva, A., Saeed, F., & Rasul Suleria, H. A. (2019). Proanthocyanidins: A comprehensive review. *Biomedicine and Pharmacotherapy*, 116(May). <https://doi.org/10.1016/j.biopha.2019.108999>
- Reis, J. B. dos A., Lorenzi, A. S., & do Vale, H. M. M. (2022). Methods used for the study of endophytic fungi: a review on methodologies and challenges, and associated tips. *Arch Microbiol*, 204(11), 1–30. <https://doi.org/10.1007/s00203-022-03283-0>
- Ribeiro, B. (2023). Publicación Oficial de Aepromo (Asociación Española de Profesionales Médicos en Ozonoterapia). Madrid, España Médico especialista em Medicina Desportiva. Mestrado em Medicina Desportiva. *Official Journal of Aepromo (Spanish Association of Medical Professionals in Ozone Therapy)*, 13(1), 175–197.
- Riga, R., Happyana, N., Quentmeier, A., Zammarelli, C., Kayser, O., & Hakim, E. H. (2021). Secondary metabolites from *Diaporthe lithocarpus* isolated from *Artocarpus heterophyllus*. *Natural Product Research*, 35(14), 2324–2328. <https://doi.org/10.1080/14786419.2019.1672685>
- Risdian, C., Mozef, T., & Wink, J. (2019). Biosynthesis of polyketides in *Streptomyces*. *Microorganisms*, 7(5), 1–18. <https://doi.org/10.3390/microorganisms7050124>
- Ritawati, A., Zuhud, E. A. M., & Hidayati, S. (2023). Analysis of the utilization of groundcherry (*Physalis angulata* L.) by the community around the Cibodas Resort Gunung Gede Pangrango National Park. *Jurnal Pengelolaan Sumberdaya Alam Dan Lingkungan*, 13(4), 613–623. <https://doi.org/10.29244/jpsl.13.4.613-623>
- Ritchie, A. M., Hua, X., Cardillo, M., Yaxley, K. J., Dinnage, R., & Bromham, L. (2021). Phylogenetic diversity metrics from molecular phylogenies: modelling expected degree of error under realistic rate variation. *Diversity and Distributions*, 27(1), 164–178. <https://doi.org/10.1111/ddi.13179>
- Roberts, J. M., Carvalhais, L. C., O'Dwyer, C., Rincón-Flórez, V. A., & Drenth, A. (2024). Diagnostics of *Fusarium* wilt in banana: Current status and challenges. *Plant Pathology*, 73(4), 760–776. <https://doi.org/10.1111/ppa.13863>
- Rohilla, P., Jain, H., Chhikara, A., Singh, L., & Dahiya, P. (2022). Anticancer

- potential of Solanaceae plants: A review. *South African Journal of Botany*, 149, 269–289. <https://doi.org/10.1016/j.sajb.2022.04.032>
- Rohmawaty, E., Rosdianto, A. M., Usman, H. A., Saragih, W. A. M., Zuhrotun, A., Hendriani, R., Wardhana, Y. W., Ekawardhani, S., Wiraswati, H. L., Agustanti, N., Bestari, M. B., & Dewi, S. (2021). Antifibrotic effect of the ethyl acetate fraction of ciplukan (*Physalis angulata* Linn.) in rat liver fibrosis induced by CCl₄. *Journal of Applied Pharmaceutical Science*, 11(12), 175–182. <https://doi.org/10.7324/JAPS.2021.1101217>
- Romero-Luna, H. E., Colina, J., Guzmán-Rodríguez, L., Sierra-Carmona, C. G., Farías-Campomanes, Á. M., García-Pinilla, S., González-Tijera, M. M., Malagón-Alvira, K. O., & Peredo-Lovillo, A. (2023). C aphysalis fruits as functional ingredients with antimicrobial activity: an emphasis on mechanisms of action. *Journal of Food Science and Technology*, 60(11), 2725–2735. <https://doi.org/10.1007/s13197-022-05578-y>
- Rosdee, S., Wisessombat, S., Tayeh, M., Malakul, R., Phanaksri, T., & Sianglum, W. (2025). Antibacterial activity of the endophytic fungal extracts and synergistic effects of combinations of ethylenediaminetetraacetic acid (EDTA) against *Pseudomonas aeruginosa* and *Escherichia coli*. *PeerJ*, 13(3). <https://doi.org/10.7717/peerj.19074>
- Rosic, N., Climstein, M., Boyle, G. M., Thanh Nguyen, D., & Feng, Y. (2023). Exploring Mycosporine-like Amino Acid UV-Absorbing Natural Products for a New Generation of Environmentally Friendly Sunscreens. *Marine Drugs*, 21(4). <https://doi.org/10.3390/md21040253>
- Rosmana, A., Sjam, S., Dewi, V. S., Asman, A., & Fhiqrah, M. (2022). *Root and collar rot disease: a new threat to young cacao (Theobroma cacao L.) plants in Sulawesi, Indonesia*. Springer.
- Rosmini, R., Hayati, N., Nasir, B., Pasar, F., & Lasmini, S. A. (2020). Pengaruh dekomposisi *Trichoderma virens* pada berbagai jenis kompos kotoran ternak untuk menekan penyakit busuk pangkal batang bawang merah. *Agromix*, 11(2), 177–188. <https://doi.org/10.35891/agx.v11i2.2081>
- Sadgrove, N. J., Oliveira, T. B., Khumalo, G. P., van Vuuren, S. F., & van Wyk, B. E. (2020). Antimicrobial isoflavones and derivatives from *erythrina* (Fabaceae): Structure activity perspective (SAR & QSAR) on experimental and mined values against *staphylococcus aureus*. *Antibiotics*, 9(5), 1–18. <https://doi.org/10.3390/antibiotics9050223>
- Sadiyah, H., Soegianto, A., Waluyo, B., & Ashari, S. (2020). Short communication: Preliminary characterization of groundcherry (*physalis angulata*) from east Java Province, Indonesia based on morpho-agronomic traits. *Biodiversitas*, 21(2), 759–769. <https://doi.org/10.13057/biodiv/d210244>
- Sadowska-Bartos, I., & Bartosz, G. (2024). Antioxidant Activity of Anthocyanins

- and Anthocyanidins: A Critical Review. *International Journal of Molecular Sciences*, 25(22). <https://doi.org/10.3390/ijms252212001>
- Safaie, N., Salehi, M., Felegari, M., Farhadi, S., Karimzadeh, S., Asadi, S., Yang, J. L., & Naghavi, M. R. (2024). Culture-based diversity of endophytic fungi of three species of *Ferula* grown in Iran. *Frontiers in Microbiology*, 15(May), 1–13. <https://doi.org/10.3389/fmicb.2024.1363158>
- Sahu, P., Tilgam, J., Mishra, S., Baba, S., Gupta, A., Krishnappa, J., Verma, S. K., & Kharwar, R. (2022). Surface sterilization for isolation of endophytes: Ensuring what (not) to grow. *Journal of Basic Microbiology*, 62. <https://doi.org/10.1002/jobm.202100462>
- Sahu, R., Mandal, S., Das, P., Ashraf, G. J., Dua, T. K., Paul, P., Nandi, G., & Khanra, R. (2023). The bioavailability, health advantages, extraction method, and distribution of free and bound phenolics of rice, wheat, and maize: A review. *Food Chemistry Advances*, 3(October), 100484. <https://doi.org/10.1016/j.focha.2023.100484>
- Saif, F. A., Yaseen, S. A., Alameen, A. S., Mane, S. B., & Undre, P. B. (2021). Identification and characterization of *Aspergillus* species of fruit rot fungi using microscopy, FT-IR, Raman and UV–Vis spectroscopy. *Spectrochimica Acta - Part A: Molecular and Biomolecular Spectroscopy*, 246. <https://doi.org/10.1016/j.saa.2020.119010>
- Salehi, B., Azzini, E., Zucca, P., Varoni, E. M., Kumar, N. V. A., Dini, L., Panzarini, E., Rajkovic, J., Fokou, P. V. T., Peluso, I., Mishra, A. P., Nigam, M., Rayess, Y. El, Beyrouthy, M. El, Setzer, W. N., Polito, L., Iriti, M., Sureda, A., Quetglas-Llabrés, M. M., ... Sharifi-Rad, J. (2020). Plant-derived bioactives and oxidative stress-related disorders: A key trend towards healthy aging and longevity promotion. *Applied Sciences (Switzerland)*, 10(3). <https://doi.org/10.3390/app10030947>
- Salem, S. H., El-Maraghy, S. S., Abdel-Mallek, A. Y., Abdel-Rahman, M. A. A., Hassanein, E. H. M., Al-Bedak, O. A., & El-Aziz, F. E. Z. A. A. (2022). The antimicrobial, antibiofilm, and wound healing properties of ethyl acetate crude extract of an endophytic fungus *Paecilomyces* sp. (AUMC 15510) in earthworm model. *Scientific Reports*, 12(1), 1–17. <https://doi.org/10.1038/s41598-022-23831-4>
- Salgado, E. R., & Arana, G. V. (2013). *Physalis angulata* L. (Bolsa Mullaca): A Review of its Traditional Uses, Chemistry and Pharmacology. *Boletín Latinoamericano y Del Caribe de Plantas Medicinales y Aromáticas*, 12(5), 431–445. <http://www.redalyc.org/articulo.oa?id=85628390001>
- Salvatore, M. M., Alves, A., & Andolfi, A. (2020). Secondary metabolites of *lasiodiplodia theobromae*: Distribution, chemical diversity, bioactivity, and implications of their occurrence. *Toxins*, 12(7). <https://doi.org/10.3390/toxins12070457>

- Samet, J. M., Chen, H., Pennington, E. R., & Bromberg, P. A. (2020). Non-redox cycling mechanisms of oxidative stress induced by PM metals. *Free Radical Biology and Medicine*, 151(December 2019), 26–37. <https://doi.org/10.1016/j.freeradbiomed.2019.12.027>
- Sandeep, A., Selvaraj, K., Kalleshwaraswamy, C. M., Hanumanthaswamy, B. C., & Mallikarjuna, H. B. (2022). Field efficacy of *Isaria fumosorosea* alone and in combination with insecticides against *Aleurodicus rugioperculatus* on coconut. *Egyptian Journal of Biological Pest Control*, 32(1). <https://doi.org/10.1186/s41938-022-00600-z>
- Sanders, E. R. (2012). Aseptic laboratory techniques: Plating methods. *Journal of Visualized Experiments*, 63, 1–18. <https://doi.org/10.3791/3064>
- Sandra, F. K., Nurhasanah, Y. S., MUTAQIN, K., Wiyono, S., & Tondok, E. T. (2021). Keragaman Morfologi dan Molekuler *Lasiodiplodia theobromae* dari Tanaman Jeruk, Kakao, Karet, Manggis, dan Pisang. *Jurnal Fitopatologi Indonesia*, 17(2), 58–66. <https://doi.org/10.14692/jfi.17.2.58-66>
- Saraswathi, M., Meshram, S. H., Siva, B., Misra, S., & Suresh Babu, K. (2022). Isolation, purification and structural elucidation of Mellein from endophytic fungus *Lasiodiplodia theobromae* strain (SJF-1) and its broad-spectrum antimicrobial and pharmacological properties. *Letters in Applied Microbiology*, 75(6), 1475–1485. <https://doi.org/10.1111/lam.13813>
- Saraswati, E., & Wijaya, A. S. (2019). Antibacterial activities of *Physalis angulata* herb extract on white feces diseases (WFD) in *Litopenaeus shrimp vannamei*. *IOP Conference Series: Earth and Environmental Science*, 236(1). <https://doi.org/10.1088/1755-1315/236/1/012103>
- Sarli, S., Kalani, M. R., & Moradi, A. (2020). A potent and safer anticancer and antibacterial taxus-based green synthesized silver nanoparticle. *International Journal of Nanomedicine*, 15, 3791–3801. <https://doi.org/10.2147/IJN.S251174>
- Sato, S., Sofian, F. F., Suehiro, W., Harneti, D., Maharani, R., Supratman, U., Abdullah, F. F., Salam, S., Koseki, T., & Shiono, Y. (2021). β -Resorcylic Acid Derivatives, with Their Phytotoxic Activities, from the Endophytic Fungus *Lasiodiplodia theobromae* in the Mangrove Plant *Xylocarpus granatum*. *Chemistry & Biodiversity*, 18(3), e2000928. <https://doi.org/https://doi.org/10.1002/cbdv.202000928>
- Satriawan, H., Zaimi, N. A., Eriadi, A., Efdi, M., Tallei, T. E., Andayani, R., & Handayani, D. (2025). Isolation and evaluation of the antimicrobial activity of endophytic fungi from *Orthosiphon aristatus*. *Biodiversitas*, 26(2), 963–970. <https://doi.org/10.13057/biodiv/d260245>
- Saxena, S., Dufossé, L., Deshmukh, S. K., Chhipa, H., & Gupta, M. K. (2024). Endophytic Fungi: A Treasure Trove of Antifungal Metabolites. In

- Microorganisms* (Vol. 12, Issue 9).
<https://doi.org/10.3390/microorganisms12091903>
- Schneewind, O., & Missiakas, D. M. (2019). Staphylococcal protein secretion and envelope assembly. *Gram-Positive Pathogens*, 7(4), 592–598.
<https://doi.org/10.1128/9781683670131.ch37>
- Shahlal, M., Mahmud, A., Bairagi, R. D., Debnath, D., & Nipa, B. S. (2024). Evaluation of Therapeutic Activity of *Physalis angulata* (In Vitro Studies). *Journal of Medicinal Natural Products*, 1(1).
- Shailabi, T. I., Kabbashi, A. S., Aldeeb, O. H., Basheer, E. S., Gumaa, R. A., Younes, E. J., Mohammed, R. F., & Ismail, A. M. (2025). Antioxidant and protective effects of methanolic *Solanum nigrum* leaf extract against CCl₄-induced hepatotoxicity and renotoxicity in albino rats. *Open Veterinary Journal*, 15(4), 1585–1592. <https://doi.org/10.5455/OVJ.2025.v15.i4.9>
- Shalapy, N. M., & Kang, W. (2022). *Fusarium oxysporum* & *Fusarium solani*: Identification, Characterization, and Differentiation the Fungal Phenolic Profiles by HPLC and the Fungal Lipid Profiles by GC-MS. *Journal of Food Quality*, 2022. <https://doi.org/10.1155/2022/4141480>
- Shamsudin, N. F., Ahmed, Q. U., Mahmood, S., Shah, S. A. A., Khatib, A., Mukhtar, S., Alsharif, M. A., Parveen, H., & Zakaria, Z. A. (2022). Antibacterial Effects of Flavonoids and Their Structure-Activity Relationship Study: A Comparative Interpretation. *Molecules*, 27(4). <https://doi.org/10.3390/molecules27041149>
- Sharifi-Rad, M., Anil Kumar, N. V., Zucca, P., Varoni, E. M., Dini, L., Panzarini, E., Rajkovic, J., Tsouh Fokou, P. V., Azzini, E., Peluso, I., Prakash Mishra, A., Nigam, M., El Rayess, Y., Beyrouthy, M. El, Polito, L., Iriti, M., Martins, N., Martorell, M., Docea, A. O., ... Sharifi-Rad, J. (2020). Lifestyle, Oxidative Stress, and Antioxidants: Back and Forth in the Pathophysiology of Chronic Diseases. *Frontiers in Physiology*, 11(July), 1–21. <https://doi.org/10.3389/fphys.2020.00694>
- Sharma, S., Mohler, J., Mahajan, S. D., Schwartz, S. A., Bruggemann, L., & Aalinker, R. (2023). Microbial Biofilm: A Review on Formation, Infection, Antibiotic Resistance, Control Measures, and Innovative Treatment. In *Microorganisms* (Vol. 11, Issue 6). <https://doi.org/10.3390/microorganisms11061614>
- Shen, N., Chen, Z., Cheng, G. X., Lin, W., Qin, Y., Xiao, Y., Chen, H., Tang, Z., Li, Q., Yuan, M., & Bu, T. (2023). Diversity, chemical constituents and biological activities of endophytic fungi from *Alisma orientale* (Sam.) Juzep. *Frontiers in Microbiology*, 14(June), 1–16. <https://doi.org/10.3389/fmicb.2023.1190624>
- Shen, Y., Gao, Y., Yang, G., Zhao, Z., Zhao, Y., Gao, L., Zhao, L., & Li, S. (2024).

- Transformation of Ginsenosides by *Lactiplantibacillus plantarum* MB11 Fermentation: Minor Ginsenosides Conversion and Enhancement of Anti-Colorectal Cancer Activity. *Molecules*, 29(1). <https://doi.org/10.3390/molecules29010027>
- Shi, X. S., Song, Y. P., Meng, L. H., Yang, S. Q., Wang, D. J., Zhou, X. W., Ji, N. Y., Wang, B. G., & Li, X. M. (2021). Isolation and characterization of antibacterial carotane sesquiterpenes from *artemisia argyi* associated endophytic *trichoderma virens* qa-8. *Antibiotics*, 10(2), 1–11. <https://doi.org/10.3390/antibiotics10020213>
- Shiono, Y., Sato, S., Sofian, F. F., Koseki, T., Abdullah, F. F., Salam, S., Harneti, D., Maharani, R., & Supratman, U. (2021). Phytotoxic β -resorcylic acid derivatives from the endophytic fungus *Lasiodiplodia theobromae* in the mangrove plant. *Phytochemistry Letters*, 44(February), 1–6. <https://doi.org/10.1016/j.phytol.2021.05.003>
- Shu, Y., Wang, J. P., Li, B. X., Gan, J. L., Ding, H., Liu, R., Cai, L., & Ding, Z. T. (2022). Bioactive cytochalasans from the fungus *Arthrinium arundinis* DJ-13. *Phytochemistry*, 194(November 2021), 113009. <https://doi.org/10.1016/j.phytochem.2021.113009>
- Simon, J., Baptiste, C., Lartaud, M., Verdeil, J. L., Brunel, B., Vercambre, G., Génard, M., Cardoso, M., Alibert, E., Goze-Bac, C., & Bertin, N. (2022). Pedicel anatomy and histology in tomato vary according to genotype and water-deficit environment, affecting fruit mass. *Plant Science*, 321(May). <https://doi.org/10.1016/j.plantsci.2022.111313>
- Singh, A., Kukreti, R., Saso, L., & Kukreti, S. (2019). Singh, A., Kukreti, R., Saso, L., & Kukreti, S. (2019). Oxidative Stress: A Key Modulator in Neurodegenerative Diseases. *Molecules* (Basel, Switzerland), 24(8), 1583. <https://doi.org/10.3390/molecules24081583>. *Molecules*, 24(8), 1–20.
- Singh, P., Arif, Y., Bajguz, A., & Hayat, S. (2021). *Posted on Authorea 11 Feb.* 1–24. <https://doi.org/10.22541/au.161306959.99455794/v1>
- Singla, R. K., Dubey, A. K., Garg, A., Sharma, R. K., Fiorino, M., Ameen, S. M., Haddad, M. A., & Al-Hiary, M. (2019). Natural polyphenols: Chemical classification, definition of classes, subcategories, and structures. *Journal of AOAC International*, 102(5), 1397–1400. <https://doi.org/10.5740/jaoacint.19-0133>
- Slika, H., Mansour, H., Wehbe, N., Nasser, S. A., Iratni, R., Nasrallah, G., Shaito, A., Ghaddar, T., Kobeissy, F., & Eid, A. H. (2022). Therapeutic potential of flavonoids in cancer: ROS-mediated mechanisms. *Biomedicine and Pharmacotherapy*, 146, 112442. <https://doi.org/10.1016/j.biopha.2021.112442>
- Soliman, H. M., & Abdel-Wahhab, M. A. (2024). Synthesis of Antibacterial

- Bioactive Compounds Using Linoleic Acid Extracted from Melon Seeds Oil and Evaluation of Its Waste Meal Ash for Fried Oil Regeneration. *Waste and Biomass Valorization*, 15(1), 487–499. <https://doi.org/10.1007/s12649-023-02161-0>
- Sottorff, I., Wiese, J., Lipfert, M., Preußke, N., Sönnichsen, F. D., & Imhoff, J. F. (2019). Different secondary metabolite profiles of phylogenetically almost identical streptomyces griseus strains originating from geographically remote locations. *Microorganisms*, 7(6). <https://doi.org/10.3390/microorganisms7060166>
- Stompor-Gorący, M., & Machaczka, M. (2021). Recent advances in biological activity, new formulations and prodrugs of ferulic acid. *International Journal of Molecular Sciences*, 22(23). <https://doi.org/10.3390/ijms222312889>
- Stosic, S., Ristic, D., Savkovic, Z., Grbic, M. L., Vukojevic, J., & Zivkovic, S. (2021). Penicillium and Talaromyces Species as Postharvest Pathogens of Pear Fruit (Pyrus communis) in Serbia. *Plant Disease*, 105(11), 3510–3521. <https://doi.org/10.1094/PDIS-01-21-0037-RE>
- Stypułkowska, A., Kolenda, R., Carolak, E., Czajkowska, J., Dutkiewicz, A., Waszczuk, W., Bińczyk, W., Thurston, T., & Grzymajlo, K. (2025). SanA is an inner membrane protein mediating Salmonella Typhimurium infection. *Microbiology Spectrum*, e0283324. <https://doi.org/10.1128/spectrum.02833-24>
- Su, P., Lu, Z., Tian, W., Chen, Y., & Maharachchikumbura, S. S. N. (2023). Six Additions to the Genus Periconia (Dothideomycetes: Periconiaceae) from Graminaceous Plants in China. *Journal of Fungi*, 9(3). <https://doi.org/10.3390/jof9030300>
- Subramani, R., Kumar, R., Prasad, P., & Aalbersberg, W. (2013). Cytotoxic and antibacterial substances against multi-drug resistant pathogens from marine sponge symbiont: Citrinin, a secondary metabolite of Penicillium sp. *Asian Pacific Journal of Tropical Biomedicine*, 3(4), 291–296. [https://doi.org/10.1016/S2221-1691\(13\)60065-9](https://doi.org/10.1016/S2221-1691(13)60065-9)
- Sun, Y., Ren, G., Shi, Q., Zhu, H., Zhou, N., Kong, X., Jiang, D., & Liu, C. (2023). Identification of a Novel Coumarins Biosynthetic Pathway in the Endophytic Fungus Fusarium oxysporum GU-7 with Antioxidant Activity Yifan. *Applied and Environmental Microbiology*, 89(1).
- Surya Hakim, S., & Yuwati, T. W. (2020). The Use of Fungal Endophyte Penicillium citrinum on Tree Seedling: Applicability and Limitation . *BIO Web of Conferences*, 20, 03005. <https://doi.org/10.1051/bioconf/20202003005>
- Sutar, N. G. (2021). Assessment and Quantification of Pharmacologically Active Marker Gallic Acid and Ellagic Acid from Leaf and Stem of Pergularia daemia Forsk. by HPTLC Method. *Technological Innovation in Pharmaceutical*

- Research Vol. 1, January*, 148–157. <https://doi.org/10.9734/bpi/tipr/v1/7532d>
- Svetlov, M. S., Koller, T. O., Meydan, S., Shankar, V., Klepacki, D., Polacek, N., Gydosh, N. R., Vázquez-Laslop, N., Wilson, D. N., & Mankin, A. S. (2021). Context-specific action of macrolide antibiotics on the eukaryotic ribosome. *Nature Communications*, 12(1), 1–14. <https://doi.org/10.1038/s41467-021-23068-1>
- Swaminathan, C., & Pandian, K. (2021). Latest Trends in Soil Science (Volume - 1). In *Latest Trends in Soil Science (Volume - 1)* (Issue January). <https://doi.org/10.22271/int.book.33>
- Swibawa, I. G., Fitriana, Y., Solikhin, Suharjo, R., Susilo, F. X., Rani, E. K. A., Haryani, M. S., & Wardana, R. A. (2020). Morpho-molecular identification and pathogenicity test on fungal parasites of guava root-knot nematode eggs in lampung, indonesia. *Biodiversitas*, 21(3), 1108–1115. <https://doi.org/10.13057/biodiv/d210334>
- Syarifah, Elfita, Widjajanti, H., Setiawan, A., & Kurniawati, A. R. (2021). Diversity of endophytic fungi from the root bark of *Syzygium zeylanicum*, and the antibacterial activity of fungal extracts, and secondary metabolite. *Biodiversitas*, 22(10), 4572–4582. <https://doi.org/10.13057/biodiv/d221051>
- Taheri, P. (2025). Endophytic fungi as regulators of phytohormones production: Cytomolecular effects on plant growth, stress protection and importance in sustainable agriculture. *Plant Stress*, 17(February), 100978. <https://doi.org/10.1016/j.stress.2025.100978>
- Tahmasebi, P., Kamrava, S., Bai, T., & Sahimi, M. (2020). Machine learning in geo- and environmental sciences: From small to large scale. *Advances in Water Resources*, 142(May), 103619. <https://doi.org/10.1016/j.advwatres.2020.103619>
- Takó, M., Kerekes, E. B., Zambrano, C., Kotogán, A., Papp, T., Krisch, J., & Vágvolgyi, C. (2020). Plant phenolics and phenolic-enriched extracts as antimicrobial agents against food-contaminating microorganisms. *Antioxidants*, 9(2). <https://doi.org/10.3390/antiox9020165>
- Tamura, K., Stecher, G., & Kumar, S. (2021). MEGA11: Molecular Evolutionary Genetics Analysis Version 11. *Mol Biol Evol*, 38(7), 3022–3027. <https://doi.org/10.1093/molbev/msab120>
- Tan, G. H., Ali, A., & Siddiqui, Y. (2023). Major fungal postharvest diseases of papaya: Current and prospective diagnosis methods. *Crop Protection*, 174(March).
- Tan, Y.-H., Cai, Y.-X., Kong, M., Li, Y.-L., Yu, Z.-P., Kong, L.-Y., & Luo, J. (2023). Withanolides from *Physalin angulata* and Their Inhibitory Effects on Nitric Oxide Production. *Chemistry & Biodiversity*, 20.

<https://doi.org/10.1002/cbdv.202300195>

- Tang, M., Fan, J., Liu, X., Liu, Z., Pei, Y., & Sun, Y. (2019). *Secondary Metabolites of Endophytic Fungus Trichoderma harzianum Isolated from Physalis angulata* / *中国实验方剂学杂志* *Objective : Method : Result : Conclusion :* 24, 802319.
- Tang, Z., Qin, Y., Chen, W., Zhao, Z., Lin, W., Xiao, Y., Chen, H., Liu, Y., Chen, H., Bu, T., Li, Q., Cai, Y., Yao, H., & Wan, Y. (2021). Diversity, Chemical Constituents, and Biological Activities of Endophytic Fungi Isolated From *Ligusticum chuanxiong* Hort. *Frontiers in Microbiology*, 12(November), 1–14. <https://doi.org/10.3389/fmicb.2021.771000>
- Tariq, H., Asif, S., Andleeb, A., Hano, C., & Abbasi, B. H. (2023). Flavonoid Production: Current Trends in Plant Metabolic Engineering and De Novo Microbial Production. *Metabolites*, 13(1). <https://doi.org/10.3390/metabo13010124>
- Taufiq, M. M. J., & Darah, I. (2019). Antibacterial activity of an endophytic fungus *Lasiodiplodia pseudotheobromae* IBRL OS-64 residing in leaves of a medicinal herb, *Ocimum sanctum* linn. *Journal of Applied Biology and Biotechnology*, 7(2), 35–41. <https://doi.org/10.7324/JABB.2019.70207>
- Taylor, J. T., Mukherjee, P. K., Puckhaber, L. S., Dixit, K., Igumenova, T. I., Suh, C., Horwitz, B. A., & Kenerley, C. M. (2020). Deletion of the *Trichoderma virens* NRPS, *Tex7*, induces accumulation of the anti-cancer compound heptelidic acid. *Biochemical and Biophysical Research Communications*, 529(3), 672–677. <https://doi.org/10.1016/j.bbrc.2020.06.040>
- Teshome, B., Teklemariam, Z., Admassu Ayana, D., Marami, D., & Asaminew, N. (2019). *Salmonella* and *Shigella* among patients with diarrhea at public health facilities in Adama, Ethiopia: Prevalence, antimicrobial susceptibility pattern, and associated factors. *SAGE Open Medicine*, 7. <https://doi.org/10.1177/2050312119846041>
- Thi Duy Ngoc, N., Yurchenko, E. A., Thi Hoai Trinh, P., Menchinskaya, E. S., Thi Dieu, T. V., Savagina, A. D., Minin, A., Thinh, P. D., Khanh, H. H. N., Thi Thanh Van, T., & Yurchenko, A. N. (2025). Secondary metabolites of Vietnamese marine fungus *Penicillium chermesinum* 2104NT-1.3 and their cardioprotective activity. *Regional Studies in Marine Science*, 81(December 2024), 104003. <https://doi.org/10.1016/j.rsma.2024.104003>
- Thomidis, T., & Kazantzis, K. (2023). Occurrence of shoot blight disease, caused by *Cylindrocarpon destructans* (Zinss) Scholten, on cherry trees (*Prunus avium*) in Greece. *European Journal of Plant Pathology*, 166(3), 359–367. <https://doi.org/10.1007/s10658-023-02669-0>
- Tian, Y., & Li, Y. (2022). A Review on Bioactive Compounds from Marine-Derived *Chaetomium* Species. *Journal of Microbiology and Biotechnology*,

32(5), 541–550. <https://doi.org/10.4014/jmb.2201.01007>

- Tironi, L. S., Carletto, L. B., Silva, E. O., Schripsema, J., & Luiz, J. H. H. (2024). Endophytic Fungi Co-Culture: An Alternative Source of Antimicrobial Substances. *Microorganisms*, 12(12), 1–22. <https://doi.org/10.3390/microorganisms12122413>
- Tiwari, P., & Bae, H. (2020). *Horizontal Gene Transfer and Endophytes : An Implication for the Acquisition of Novel Traits*.
- Toghueo, R. M. K., & Boyom, F. F. (2020). Endophytic *Penicillium* species and their agricultural, biotechnological, and pharmaceutical applications. In 3 *Biotech* (Vol. 10, Issue 3). Springer International Publishing. <https://doi.org/10.1007/s13205-020-2081-1>
- Toppo, P., Kagatay, L. L., Gurung, A., Singla, P., Chakraborty, R., Roy, S., & Mathur, P. (2023). Endophytic fungi mediates production of bioactive secondary metabolites via modulation of genes involved in key metabolic pathways and their contribution in different biotechnological sector. In 3 *Biotech* (Vol. 13, Issue 6). Springer International Publishing. <https://doi.org/10.1007/s13205-023-03605-z>
- Tran, S. L., Lebreuilly, L., Cormontagne, D., Samson, S., Tô, T. B., Dervyn, R., Griebhammer, A., De, J., Cuesta-Zuluaga, L., Maier, L., Naas, T., Mura, S., Nicolas, J., Rognan, D., André, G., & Ramarao, N. (2025). An anti-virulence drug targeting the evolvability protein Mfd protects against infections with 1 antimicrobial resistant ESKAPE pathogens 2 3 4. *Nature Communications*. <https://doi.org/10.1038/s41467-025-58282-8>
- Tranbarger, T. J., & Tadeo, F. R. (2025). Abscission zone metabolism impacts pre- and post-harvest fruit quality: a very attaching story. *Frontiers in Plant Science*, 15(January), 1–20. <https://doi.org/10.3389/fpls.2024.1524893>
- Trautmann, E., Attin, T., Mohn, D., & Zehnder, M. (2021). Hydrogen Peroxide Versus Sodium Hypochlorite: All a Matter of pH? *Journal of Endodontics*, 47(2), 297–302. <https://doi.org/10.1016/j.joen.2020.10.016>
- Tripathi, A., Pandey, P., Tripathi, S. N., & Kalra, A. (2025). Plant growth-promoting endophytic consortium improved artemisinin biosynthesis via modulating antioxidants, gene expression, and transcriptional profile in *Artemisia annua* (L.) under stressed environments. *Plant Stress*, 15(December 2024), 100757. <https://doi.org/10.1016/j.stress.2025.100757>
- Tsuneda, T. (2020). Fenton reaction mechanism generating no OH radicals in Nafion membrane decomposition. *Scientific Reports*, 10(1), 1–13. <https://doi.org/10.1038/s41598-020-74646-0>
- Tu, N. H. K., Thuy, N. T. T., Vinh, D. T. T., Anh, N. P. Q., & Ly, H. D. (2021). Detection of antimicrobial, antioxidant and cytotoxicity activities of fusarium

- oxysporum f01 isolated from catharanthus roseus collected in vietnam. *Journal of Pure and Applied Microbiology*, 15(3), 1643–1654. <https://doi.org/10.22207/JPAM.15.3.61>
- Tuan Anh, H. Le, Le Ba, V., Do, T. T., Phan, V. K., Pham Thi, H. Y., Bach, L. G., Tran, M. H., Tran Thi, P. A., & Kim, Y. H. (2021). Bioactive compounds from *Physalis angulata* and their anti-inflammatory and cytotoxic activities. *J Asian Nat Prod Res*, 23(8), 809–817. <https://doi.org/10.1080/10286020.2020.1825390>
- Tuerdibieke, M., Tian, X., An, X., Feng, Y., & Liu, W. (2024). Isolation and identification of endophytic fungi from *Alhagi sparsifolia* Shap. and their antibacterial activity. *Heliyon*, 10(19), e39003. <https://doi.org/10.1016/j.heliyon.2024.e39003>
- Tulum, I., Tahara, Y. O., & Miyata, M. (2019). Peptidoglycan layer and disruption processes in *Bacillus subtilis* cells visualized using quick-freeze, deep-etch electron microscopy. *Microscopy*, 68(6), 441–449. <https://doi.org/10.1093/jmicro/dfz033>
- Tziros, G. T., Samaras, A., & Karaoglanidis, G. S. (2022). *Fusarium equiseti* as an Emerging Foliar Pathogen of Lettuce in Greece: Identification and Development of a Real-Time PCR for Quantification of Inoculum in Soil Samples.
- Ujamp, N. T., Ezep, P. M., Ejikeugwup, C., Festus, P., Okoyep, B. C., & Esimonep, C. O. (2020). Antimicrobial Activity of Metabolites of *Lasiodiplodia theobromae* isolated from *Psidium guajava*. *IJISSET-International Journal of Innovative Science, Engineering & Technology*, 7(2), 123–129. www.ijiset.com
- Ushie, O. A., Iyen, S. I., Abeng, F. E., Azuaga, T. ., Okpaegbe, U. C., & Aikhoje, E. F. (2019). Quantification of Alkaloids, Flavonoids and Saponins in *Physalis angulata* and *Mucuna Pruriens*. *FRsCS*, 1(02), 86–89.
- van den Brule, T., Punt, M., Teertstra, W., Houbraken, J., Wösten, H., & Dijksterhuis, J. (2020). The most heat-resistant conidia observed to date are formed by distinct strains of *Paecilomyces variotii*. *Environmental Microbiology*, 22(3), 986–999. <https://doi.org/10.1111/1462-2920.14791>
- van der Vaart, T. W., Prins, J. M., Soetekouw, R., van Twillert, G., Veenstra, J., Herpers, B. L., Rozemeijer, W., Jansen, R. R., Bonten, M. J. M., & van der Meer, J. T. M. (2022). All-Cause and Infection-Related Mortality in *Staphylococcus aureus* Bacteremia, a Multicenter Prospective Cohort Study. *Open Forum Infectious Diseases*, 9(12), 1–9. <https://doi.org/10.1093/ofid/ofac653>
- Van Hung, P. (2016). Phenolic Compounds of Cereals and Their Antioxidant Capacity. *Critical Reviews in Food Science and Nutrition*, 56(1), 25–35.

<https://doi.org/10.1080/10408398.2012.708909>

- Vargas-Arana, G., Torres-Benítez, A., Ortega-Valencia, J. E., Merino-Zegarra, C., Carranza-Rosales, P., & Simirgiotis, M. J. (2025). Untargeted Chemical Profile, Antioxidant, and Enzyme Inhibition Activity of *Physalis angulata* L. from the Peruvian Amazon: A Contribution to the Validation of Its Pharmacological Potential. *Antioxidants*, 14(3). <https://doi.org/10.3390/antiox14030246>
- Vaz, C., Nascimento, M., Carriço, J. A., Rocher, T., & Francisco, A. P. (2021). Distance-based phylogenetic inference from typing data: A unifying view. *Briefings in Bioinformatics*, 22(3), 1–16. <https://doi.org/10.1093/bib/bbaa147>
- Vázquez de Aldana, B. R., Arellano, J. B., Cuesta, M. J., Mellado-Ortega, E., González, V., & Zabalgogezcoa, I. (2021). Screening fungal endophytes from a wild grass for growth promotion in tritordeum, an agricultural cereal. *Plant Science*, 303(July 2020). <https://doi.org/10.1016/j.plantsci.2020.110762>
- Verma, A., Shameem, N., Jatav, H. S., Sathyanarayana, E., Parray, J. A., Poczai, P., & Sayyed, R. Z. (2022). Fungal Endophytes to Combat Biotic and Abiotic Stresses for Climate-Smart and Sustainable Agriculture. *Frontiers in Plant Science*, 13(July), 1–16. <https://doi.org/10.3389/fpls.2022.953836>
- Verma, S. C., Qian, Z., & Adhya, S. L. (2019). Architecture of the *Escherichia coli* nucleoid. In *PLoS Genetics* (Vol. 15, Issue 12). <https://doi.org/10.1371/journal.pgen.1008456>
- Vieceli, P. S., Juiz, P. J. L., Lauria, P. S. S., Couto, R. D., Tomassini, T. C. B., Ribeiro, I. M., Soares, M. B. P., & Villarreal, C. F. (2021). *Physalis angulata* reduces the progression of chronic experimental periodontitis by immunomodulatory mechanisms. *Journal of Ethnopharmacology*, 273(July 2020). <https://doi.org/10.1016/j.jep.2021.113986>
- Viogenta, P., Susanti, L., & Megasari, L. (2023). Antibacterial Activity and Bioautography of the Chloroform Fraction of Morel Berry (*Physalis angulata* L.) Root Against *Staphylococcus epidermidis* and *Pseudomonas aeruginosa*. *Journal of Multidisciplinary Applied Natural Science*, 3(1), 90–99. <https://doi.org/10.47352/jmans.2774-3047.148>
- Vuolo, M. M., Lima, V. S., & Maróstica Junior, M. R. (2019). Phenolic Compounds. In *Bioactive Compounds: Health Benefits and Potential Applications*. Elsevier Inc. <https://doi.org/10.1016/B978-0-12-814774-0.00002-5>
- Wahyudi, B. E., Elfita, Widjajanti, H., Salni, Eliza, Julinar, & Musifa, E. (2025a). Diversity and bioactivity of endophytic fungi isolated from various fruit organs of *Physalis angulata*. *Biodiversitas*, 26(3), 1485–1499. <https://doi.org/10.13057/biodiv/d260349>

- Wahyudi, B. E., Widjajanti, H., & Oktiansyah, R. (2025b). Bioactive Compound from the Endophytic Fungus *Paecilomyces variotii* Isolated from the Stems of *Physalis angulata* L. *Trends Sci*, 22(1), 1–16.
- Wahyuningsih, S. (2019). Biodegradation of pulp sludge by *Phanerochaete chrysosporium*, *penicillium oxalicum* and *penicillium citrinum* after six months incubation. *Indonesian Journal of Forestry Research*, 6(2), 95–105. <https://doi.org/10.20886/IJFR.2019.6.2.95-105>
- Wang, B., Cai, J., Huang, L., Chen, Y., Wang, R., Luo, M., Yang, M., Zhang, M., Nasihat, Chen, G., Huang, G., & Zheng, C. (2024). Significance of research on natural products from marine-derived *Aspergillus* species as a source against pathogenic bacteria. *Frontiers in Microbiology*, 15(September). <https://doi.org/10.3389/fmicb.2024.1464135>
- Wang, J., Ma, W., & Wang, X. (2021). Insights into the structure of *Escherichia coli* outer membrane as the target for engineering microbial cell factories. *Microbial Cell Factories*, 20(1), 1–17. <https://doi.org/10.1186/s12934-021-01565-8>
- Wang, L., Lu, S., Wang, L., Xin, M., Xu, Y., Wang, G., Chen, D., Chen, L., Liu, S., & Zhao, F. (2021). Anti-inflammatory effects of three withanolides isolated from *Physalis angulata* L. in LPS-activated RAW 264.7 cells through blocking NF- κ B signaling pathway. *Journal of Ethnopharmacology*, 276, 114186. <https://doi.org/https://doi.org/10.1016/j.jep.2021.114186>
- Wang, P., Yang, X.-M., Hu, Z.-X., Li, Y.-N., Yang, J., Hao, X.-J., Yuan, C.-M., & Yi, P. (2023). UPLC-Q-Orbitrap-MS/MS-Guided Isolation of Bioactive Withanolides from the Fruits of *Physalis angulata*. *Journal of Agricultural and Food Chemistry*, 71(44), 16581–16592. <https://doi.org/10.1021/acs.jafc.3c04311>
- Wang, S., Li, Y., He, L., Yang, J., Fernie, A. R., & Luo, J. (2022). Natural variance at the interface of plant primary and specialized metabolism. *Current Opinion in Plant Biology*, 67, 102201. <https://doi.org/https://doi.org/10.1016/j.pbi.2022.102201>
- Wang, W., † Z., Wu, C., Shahid, H., Wang, S., & Shan, T. (2021). Diversity and Antibacterial and Antioxidant Activities of Fungal Endophytes from the Roots of *Eucalyptus deglupta*. *Sains Malaysiana*, 51, 1687–1696. <https://doi.org/10.17576/jsm-2022-5106-07>
- Wang, X., Wang, X., Yao, Y., Chen, B., Zhang, M., Zhang, M., Liu, Z., Wang, Q., Wang, Q., & Ma, J. (2020). *Paecilomyces variotii* Extracts and Controlled-Release Urea Synergistically Increased Nitrogen Use Efficiency and Rice Yield. *ACS Omega*, 5(22), 13303–13311. <https://doi.org/10.1021/acsomega.0c01348>
- Wang, Y., Li, Z., Xu, M., Xiao, Z., Liu, C., Du, B., Xu, D., & Li, L. (2023). Signal

- Molecules Regulate the Synthesis of Secondary Metabolites in the Interaction between Endophytes and Medicinal Plants. *Processes*, 11(3). <https://doi.org/10.3390/pr11030849>
- Wei, W., Khan, B., Dai, Q., Lin, J., Kang, L., Rajput, N. A., Yan, W., & Liu, G. (2023). Potential of Secondary Metabolites of Diaporthe Species Associated with Terrestrial and Marine Origins. *Journal of Fungi*, 9(4). <https://doi.org/10.3390/jof9040453>
- Wen, J., Okyere, S. K., Wang, S., Wang, J., Xie, L., Ran, Y., & Hu, Y. (2022). Endophytic Fungi: An Effective Alternative Source of Plant-Derived Bioactive Compounds for Pharmacological Studies. *J Fungi*, 8(2). <https://doi.org/10.3390/jof8020205>
- Wen, L., Chen, G., She, Z., Yan, C., Cai, J., & Mu, L. (2010). Two new paeciloxocins from a mangrove endophytic fungus Paecilomyces sp. *Russian Chemical Bulletin*, 59(8), 1656–1659. <https://doi.org/10.1007/s11172-010-0290-1>
- Wen, Q. H., Wang, R., Zhao, S. Q., Chen, B. R., & Zeng, X. A. (2021). Inhibition of biofilm formation of foodborne staphylococcus aureus by the citrus flavonoid naringenin. *Foods*, 10(11). <https://doi.org/10.3390/foods10112614>
- Weng, W., He, M., Zheng, Z., Lin, T., Lai, Z., Zheng, S., & Wu, X. (2024). Tomato Pedicel Physical Characterization for Fruit-Pedicel Separation Tomato Harvesting Robot. *Agronomy*, 14(10), 1–20. <https://doi.org/10.3390/agronomy14102274>
- Wiraswati, H., Ekawardhani, S., Rohmawaty, E., Laelalugina, A., Zuhrotun, A., Hendriani, R., Wardhana, Y., Bestari, M., Sahirdjan, E., & Dewi, S. (2024). Antioxidant, Antiinflammation, and Antifibrotic Activity of Ciplukan (*Physalis angulata* L). Extract. *Journal of Inflammation Research*, Volume 17(September), 6297–6306. <https://doi.org/10.2147/jir.s470318>
- Wong, H. J., Mohamad-Fauzi, N., Rizman-Idid, M., Convey, P., & Alias, S. A. (2019). Protective mechanisms and responses of micro-fungi towards ultraviolet-induced cellular damage. *Polar Science*, 20(September 2018), 19–34. <https://doi.org/10.1016/j.polar.2018.10.001>
- Wu, J., Zhao, J., Zhang, T., Gu, Y., Khan, I. A., Zou, Z., & Xu, Q. (2021). Naturally occurring physalins from the genus *Physalis*: A review. *Phytochemistry*, 191(June). <https://doi.org/10.1016/j.phytochem.2021.112925>
- Wu, Y. M., Zhou, Q. Y., Yang, X. Q., Luo, Y. J., Qian, J. J., Liu, S. X., Yang, Y. Bin, & Ding, Z. T. (2019). Induction of Antiphytopathogenic Metabolite and Squalene Production and Phytotoxin Elimination by Adjustment of the Mode of Fermentation in Cocultures of Phytopathogenic *Nigrospora oryzae* and *Irpex lacteus*. *Journal of Agricultural and Food Chemistry*. <https://doi.org/10.1021/acs.jafc.9b04209>

- Wulandari, S. L., Kaloik, P., Lakobal, P., & Wenda, N. (2021). *Isolasi dan Seleksi Mikrob Penghasil Asam Sitrat dari Buah-buahan Busuk*. 1(1), 9–12.
- Xiao, J., Huang, J., Xiao, K., Li, G., Yang, S., & He, Y. (2024). How different of the rhizospheric and endophytic microbial compositions in watermelons with different fruit shapes. *PLoS ONE*, 19(5 May), 1–15. <https://doi.org/10.1371/journal.pone.0302462>
- Xiao, Y., Liang, W., Zhang, Z., Wang, Y., Zhang, S., Liu, J., Chang, J., Ji, C., & Zhu, D. (2022). Polyketide Derivatives from the Endophytic Fungus *Phaeosphaeria* sp. LF5 Isolated from *Huperzia serrata* and Their Acetylcholinesterase Inhibitory Activities. *Journal of Fungi*, 8(3). <https://doi.org/10.3390/jof8030232>
- Xu, K., Li, X. Q., Zhao, D. L., & Zhang, P. (2021). Antifungal Secondary Metabolites Produced by the Fungal Endophytes: Chemical Diversity and Potential Use in the Development of Biopesticides. *Frontiers in Microbiology*, 12(June), 1–17. <https://doi.org/10.3389/fmicb.2021.689527>
- Xu, W., Shi, D., Chen, K., & Popovich, D. G. (2024). TLC-Bioautography-Guided Isolation and Assessment of Antibacterial Compounds from Manuka (*Leptospermum scoparium*) Leaf and Branch Extracts. *Molecules*, 29(717), 1–13.
- Yan, F., Li, K., Xing, W., Dong, M., Yi, M., & Zhang, H. (2022). Role of Iron-Related Oxidative Stress and Mitochondrial Dysfunction in Cardiovascular Diseases. *Oxidative Medicine and Cellular Longevity*, 2022. <https://doi.org/10.1155/2022/5124553>
- Yan, J., Manawasinghe, I., Jayawardena, R., Hl, L., Yy, Z., Zhang, W., Phillips, A., Wanasinghe, D., Dissanayake, A., Xh, L., Yh, L., Kd, H., & Jy, Y. (2021). Microfungi associated with *Camellia sinensis*: A case study of leaf and shoot necrosis on Tea in Fujian, China. *Mycosphere*, 12, 430– 518. <https://doi.org/10.5943/mycosphere/12/1/6>
- Yap, P. S. X., Yusoff, K., Lim, S. H. E., Chong, C. M., & Lai, K. S. (2021). Membrane disruption properties of essential oils-a double-edged sword? *Processes*, 9(4), 1–11. <https://doi.org/10.3390/pr9040595>
- Yapıcı, İ., Altay, A., Öztürk Sarıkaya, B., Korkmaz, M., Atila, A., Gülçin, İ., & Köksal, E. (2021). In vitro Antioxidant and Cytotoxic Activities of Extracts of Endemic *Tanacetum erzincanense* Together with Phenolic Content by LC-ESI-QTOF-MS. *Chemistry and Biodiversity*, 18(3). <https://doi.org/10.1002/cbdv.202000812>
- Yavuz, O., Dincel, G. C., Yildirim, S., El-Ashram, S., & Al-Olayan, E. (2024). Impact of apoptosis and oxidative stress on pancreatic beta cell pathophysiology in streptozotocin-induced Type 1 diabetes mellitus. *Tissue and Cell*, 91, 102552.

<https://doi.org/https://doi.org/10.1016/j.tice.2024.102552>

- Yehia, R. S. (2023). Multi-Function of a New Bioactive Secondary Metabolite Derived from Endophytic Fungus *Colletotrichum acutatum* of *Angelica sinensis*. *Journal of Microbiology and Biotechnology*, 33(6), 806–822. <https://doi.org/10.4014/jmb.2206.06010>
- Yin, W., Wei, Y., Li, G., Wang, Y., Lou, Y., Pan, H., Yang, Q., Hu, G., Wang, H., & Zhuge, Y. (2023). Low concentration of *Paecilomyces variotii* extracts promote the growth and disease resistance of pepper. *Cogent Food and Agriculture*, 9(1). <https://doi.org/10.1080/23311932.2023.2195975>
- Yip, D. W., & Gerriets, V. (2024). *Penicillin* (pp. 1–8). StatPearls Publishing.
- Yohanna, C., Kwaji, A., & Atiko, R. (2021). Antibacterial Activity, Antioxidant Potential and Stigmasterol Isolation from *Laggera aurita* Linn (Asteraceae). *International Journal of Biochemistry Research & Review*, 30(9), 24–39. <https://doi.org/10.9734/ijbcr/2021/v30i930288>
- Yoshikawa, T., & You, F. (2024). *Oxidative Stress and Bio-Regulation*. 1–17.
- Yu, J., & Yin, C. (2025). *Community Structure Diversity of Endophytic Fungi in Cissampelos pareira from Different Habitats and Their α -Glucosidase Inhibitory Activity*. 1–19.
- Yu, X., Müller, W. E. G., Frank, M., Gao, Y., Guo, Z., Zou, K., Proksch, P., & Liu, Z. (2023). Caryophyllene-type sesquiterpenes from the endophytic fungus *Pestalotiopsis lespedezae* through an OSMAC approach. *Frontiers in Microbiology*, 14(January), 1–9. <https://doi.org/10.3389/fmicb.2023.1248896>
- Zahra, Z. S. A. (2024). Nuclear Magnetic Resonance (NMR): Principle, Applications, Types, and Uses in Metabolite Identification and Medical Biotechnology. *Current Clinical and Medical Education*, 02(May), 33–48.
- Zaid, R., Koren, R., Kligun, E., Gupta, R., Leibman-Markus, M., Mukherjee, P. K., Kenerley, C. M., Bar, M., & Horwitz, B. A. (2022). Gliotoxin, an Immunosuppressive Fungal Metabolite, Primes Plant Immunity: Evidence from *Trichoderma virens*-Tomato Interaction. *MBio*, 13(4), 1–19. <https://doi.org/10.1128/mbio.00389-22>
- Zainudin, N. A. I. M., Murad, N. B. A., Aris, A., & Hussain, N. H. (2023). Disease Note: Diseases Caused by Fungi and Fungus-Like Organisms First. *Plant Disease*. <https://doi.org/10.13039/501100003093>
- Zakaria, L. (2024). An Overview of *Aspergillus* Species Associated with Plant Diseases. *Pathogens*, 13(9). <https://doi.org/10.3390/pathogens13090813>
- Zakariyah, R. F., Ajijolakewu, K. A., Ayodele, A. J., Folami-A, B. I., Samuel, E. P., Otuoze, S. O., Abdulrauf, L. B., & Ahmed, R. N. (2024). Progress in endophytic fungi secondary metabolites: biosynthetic gene cluster reactivation

- and advances in metabolomics. *Bulletin of the National Research Centre*, 48(1). <https://doi.org/10.1186/s42269-024-01199-x>
- Zambrano, C., Kerekes, E. B., Kotogán, A., Papp, T., Vágvölgyi, C., Krisch, J., & Takó, M. (2019). Antimicrobial activity of grape, apple and pitahaya residue extracts after carbohydrase treatment against food-related bacteria. *LWT*, 100, 416–425. <https://doi.org/https://doi.org/10.1016/j.lwt.2018.10.044>
- Zaritsky, A., Vollmer, W., Männik, J., & Liu, C. (2019). Does the nucleoid determine cell dimensions in *Escherichia coli*? *Frontiers in Microbiology*, 10(JULY). <https://doi.org/10.3389/fmicb.2019.01717>
- Zeece, M. (2020). *Chapter Five - Vitamins and minerals* (M. B. T.-I. to the C. of F. Zeece (ed.); pp. 163–212). Academic Press. <https://doi.org/https://doi.org/10.1016/B978-0-12-809434-1.00005-0>
- Zehiroglu, C., & Ozturk Sarikaya, S. B. (2019). The importance of antioxidants and place in today's scientific and technological studies. *Journal of Food Science and Technology*, 56(11), 4757–4774. <https://doi.org/10.1007/s13197-019-03952-x>
- Zeliger, H. I. (2023). *Chapter 18 - Free radicals* (H. I. B. T.-O. S. Zeliger (ed.); pp. 211–226). Academic Press. <https://doi.org/https://doi.org/10.1016/B978-0-323-91890-9.00019-2>
- Zeng, L., Zhou, X., Liao, Y., & Yang, Z. (2021). Roles of specialized metabolites in biological function and environmental adaptability of tea plant (*Camellia sinensis*) as a metabolite studying model. *Journal of Advanced Research*, 34(723), 159–171. <https://doi.org/10.1016/j.jare.2020.11.004>
- Zhang, H., Shen, X., Shen, W., Zhang, D., Huang, X., Zhu, K., Liu, J., & Li, G. (2024). Glutathione peroxidase LtGPX3 contributes to oxidative stress tolerance, virulence, and plant defense suppression in the peach gummosis fungus *Lasiodiplodia theobromae*. *Phytopathology Research*, 6(1), 1–15. <https://doi.org/10.1186/s42483-024-00224-1>
- Zhang, J., Lu, J., Zhu, Y., Shen, X., Zhu, B., & Qin, L. (2024). Roles of endophytic fungi in medicinal plant abiotic stress response and TCM quality development. *Chinese Herbal Medicines*, 16(2), 204–213. <https://doi.org/10.1016/j.chmed.2023.02.006>
- Zhang, T., Alexa, E.-A., Liu, G., Berisha, A., Walsh, R., & Kelleher, R. (2025). *Lycium barbarum* for Health and Longevity: A Review of Its Biological Significance. *Obesities*, 5(2), 35. <https://doi.org/10.3390/obesities5020035>
- Zhang, W., Pan, X., Fu, J., Cheng, W., Lin, H., Zhang, W., & Huang, Z. (2024). Phytochemicals derived from *Nicotiana tabacum* L. plant contribute to pharmaceutical development. *Frontiers in Pharmacology*, 15(April), 1–19. <https://doi.org/10.3389/fphar.2024.1372456>

- Zhao, L. Y., Shi, J., Xu, Z. Y., Sun, J. L., Yan, Z. Y., Tong, Z. W., Tan, R. X., Jiao, R. H., & Ge, H. M. (2024). Hybrid Type I and II Polyketide Synthases Yield Distinct Aromatic Polyketides. *Journal of the American Chemical Society*, 146(43), 29462–29468. <https://doi.org/10.1021/jacs.4c08803>
- Zheng, J., Wang, H., Wang, X., Zeng, S., Yuan, S., & Yin, T. (2025). Genome mining and OSMAC strategies unveil diverse secondary metabolites from the endophytic fungus *Diaporthe kyushuensis* ZMU-48-1 with antifungal activity. *Frontiers in Microbiology*, 16(June). <https://doi.org/10.3389/fmicb.2025.1604639>
- Zheng, M., Liu, Y., Zhang, G., Yang, Z., Xu, W., & Chen, Q. (2023). The Applications and Mechanisms of Superoxide Dismutase in Medicine, Food, and Cosmetics. *Antioxidants*, 12(9). <https://doi.org/10.3390/antiox12091675>
- Zin, N. A., & Badaluddin, N. A. (2020). Biological functions of *Trichoderma* spp. for agriculture applications. *Annals of Agricultural Sciences*, 65(2), 168–178. <https://doi.org/10.1016/j.aos.2020.09.003>