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Study on Bacillus thuringiensis Indigenous Highland of South Sumatera-Based Bioinsecticide Towards Lepidopteran Insect Pests

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Abstract— The objectives of research were 1) to explore the presence of Bacillus thuringiensis from highland soil of South Sumalin 2) to investigate crystal proteins and their toxicity against diamondback moth Plutella xylostella and armyworm Spodoptera little and 3) to produce B. thuringiensis – based product of the most promising B. thuringiensis isolate. Exploration of soil resulted 331 thuringiensis isolates in which 21 isolates were toxic against P. xylostella and 15 isolates were toxic towards S.litura. Two isolates namely SASU and KATB, were very toxic to both insects. Developing of those isolates as bio-insecticide was done in three mingrowth media i.e. coconut water, soybean soaking water, tofu liquid waste, mixtures of coconut water and soybean soaking water (15 v/v), mixtures of coconut water and tofu liquid waste (1:1. v/v), and Nutrient Broth, as control. Total Viable Spore Count (TVS) showed spore product was ranged from 2.22 x 106 until 8.98 x 108spores/ml resulted in high mortality of P. xylostella and of S. line indicating the presence of toxic crystal protein.

Keywords--- Bacillus thuringiensis; bio-insecticide; Plutella xylostella; Spodoptera litura.

I. INTRODUCTION

Bacillus thuringiensis(Bt) is a gram-positive, rod-shaped, aerobic and spore-forming bacteria. The presence of inclusions in B. thuringiensis have been found and detected in the inclusion parasporal crystal structure containing more than one type of insecticidal crystal proteins (insecticidal crystal protein, ICP) or also called delta endotoxins [1] and it will be produced by Bt during sporulation [2]. This bacterium can be found in soil, various plants, including vegetables, cotton, tobacco, and forest plants. In the environment with good conditions and adequate nutrition, bacterial spores can survive and continue the vegetative growth [3] [4] [5]. Various strains of Bt isolates have been demonstrated to control various plant pests. Some member of these orders Lepidoptera, Coloeptera, Diptera, Hymenoptera, Homoptera, Molophoga, and Acari are target of Bt [6]. In Indonesia, important insect pests are armyworm Spodoptera linera (Lepidoptera: Noctuidae) diamondback moth Plutella xylostella (Lepidoptera :Plutellidae) because of their characteristic of life.[7].

Bt-based product (known as bio-insecticide) was made from waste material content of carbohydrate, nitrogen, protein and some other minerals. Devi et al. [8] used when bran based media to produce bioinsecticide toxic to larvae of castor semilooper, Achaea jamata L. Chilcott and Pillai [8] used coconut wastes for production of B. thuringiensis we israelensis. High production of Bt based -bioinsecticide we depended on carbon, nitrogen, water content, mineral element and suitable growth condition. The strain of local B. thuringiensis also played a role in achievement of manufacturing process. This paper presented earlier observation of B. thuringiensis isolated from highland of South Sumatera and their effectiveness to kill insect pests.

II. MATERIAL AND METHOD

A. Soil Sample Collection

Samples were collected from soil in location had not bee treated by *B. thuringiensis*-bio insecticide. Location was a high land of South Sumatera. Samples were collected by scrapping off material by an sterile spatula and obtaining g soil below 5 cm from surface. Samples were kept on 42 until use.

B. Isolation of B. thuringiensis

Five g of soil samples is diluted well in 15 ml dH₂0 in to tube. Shaked well until perfectly diluted. One ml of upper part of dilution is taken in eppendorf tube, added by 1 µl Triton X-100, and heated in waterbath 85°C 15 minutes. With a sterile spatula, the solution was streaked on the medium NaCl Glycine Kim and Goepfert (NGKG) on petridish. Petridish incubated at 30°C, for 24-72 hours. Colonies of Bt will grow in white colour. After 24-72 hours incubation, proteinaceous parasporal inclusion bodies will presence. Identification of B. thuringiensis refers to Thiery and Frachon [10].

C. Insect Test mass-rearing

Groups of eggs of armyworm S. litura and diamondback worm P. xylastella were obtained from the field and subsequently maintained in the laboratory. Larvae reared in a plastic container maintenance (d = 15 cm and h = 9 cm). Depending on species, food used were the leaves of water spinnel (Ipomoca reptana) grown without pesticide peatment for mass rearing for S. litura, and brassica leaves for P. xylostella, as well. Temperature and relative humidity were maintained. Maintenance of container was done by cleaning of residual dirt and food remains to ensure the availability of food and cleanliness. At the bottom of the box was placed maintenance of sterile soil that had been sterilized as a place of S. litura to become pupae. If the caterpillar has reached prepupa phase characterized by no activity, meaning caterpillar will enter the pupa stage. Larvae of S. litura reared to be a phase of insect pupae, and imago. Insect samples used were second generation (F2).

D. Preliminary Test of Bt isolate (Screening test)

Leaves of spinnch and leaves of brassica were prepared for screening test. Bt isolates were prepared in single dose of 10⁶ spores/ml. Leaves were dipped in Bt about 3 minutes, dried-air and transferred into petri dish. Second-instar larvae of Slitura were placed in petri dish with Bt treated spinach leaves, and third instar larvae of P. xylostella were place in treated leaves of brassica. Each isolates was tested by 20 larvae. Mortality of larvae was observed and counted.

E. Mass Production of Bt spores in Various Media

Two isolates will be chosen for mass-production of Bt spores with criteria they showed the highest mortality towards both insect pests. Media used for mass production was 1). Coconut water, 2). Soybean soaking water, 3) Tofu liquid waste, 4). Mixture of coconut water and soybean soaking water (1:1. v/v), 5). Mixture of coconut water and tofu liquid waste (1:1. v/v), and 6). Nutrient Broth (Control). The media were individually added by 0.3 g/l MgSO4.7H2O, 0.02 g/l FeSO4.7H2O, 0.02 g/l MnSO4.7H2O, 0.02 g/l SnSO4.7H2O and 0.01 g/l CaCO3 following the method of Dulmage and Rhodes [11] Those media were shaken 300 rpm for 72 days. Total Viable Spore Count (TVSC) was observed. Two isolates chosen were checked their protein shape by SEM (Scanning Electron Microscope)

F. Bioassay of Bt-product towards S.litura and P. xylustella

TVSC of Bt product was used as treatment for bioassay towards S. linura and P. xylostella. Experiment was done by Completely Randomized Design (CRD) with 6 treatments and 5 replications. Leaves were dipped in Bt about 3 minutes. dried-air and transferred into petri dish. Second-instar larvae

of S.litura were placed in petri dish with Bt treated spinach leaves, and third instar larvae of P. sydostella were place in treated leaves of brassica. Each replication was tested by 10 larvae. Mortality was observed until 5 days.

III. RESULT AND DISCUSSION

A. Isolation of Bacillus thuringiensis

Soil sampling was conducted in highland of South Sumatera consisted of 4 districts namely Pagaralam district (985m asl), Lahat district (925 m asl), OKU Selatan district (950 m asl) and Munra Enim district (915 m asl). Exploration of soil resulted 33 B. thereingiensis isolates in which 21 isolates were toxic against P. sylostella and 15 isolates were toxic towards S.litura. Data was shown in Table 1.

TABLE I
SCREENING TEST OF BY BOLATED FROM HIGHLAND OF SOUTH SUMATERA
AGAINST SPODOPTERA LITURA AND PLUTELLA VILOSTELLA

No.	Isolate code	Location	Mortality (%)	
			S.litma	P. xylostella
1	BAK	Pagaralam (985m asi)	35	*40
2	BAC		6.5	0
3	PWP		35	20
4	KDu		40	45
5	KRa		0	0
6	PKs		0	0
7	PKe		30	45
8	PKo		0	40
9	PCe	Labat	0	0.
10	DMS	(925 m asl)	40	0.
11	DMA		30	50
12	DMP		0	65
13	DMK		0	0
14	SRK		0.	0
15	SRA		0	0
16	SRJ		0	55
17	SKD		0	50
18	PGK	OKU Selatan (950 m asil)	0	40
10	APLB		55	0
20	CELB		0	60
21	PILB		70	0
22	DRPD		0	0
23	DUPD		60	45
24	MAPD		- 55	50
25	JABD	Musra Enim (985 m asl)	0	0
26	KHBD		0	65
27	SEBD		35	40
28	KATH		90	95
29	PITB		0	40
30	MATB		45	45
31	PELM		0	45
32	SASU		95	95
33	RASU		50	40

Two isolates toxic to both S.litura and P. xylostella were chosen among 33 isolates, i.e. SASU and KATB. Their toxicity towards S.litura was 90 and 95 % (SASU-product) and 95 and 95 % on P. xylostella. There was any possibility that these isolates contained of cry: I gene and specific shape of crystal proteins. Asano et al [12] showed that B. thuringiensis toxic to S. litura belongs to cry I gene group. These two isolates will be used as material to mass-production of Bt. The shape of two proteins was observed by Scanning Electron Microscope (SEM). The photographs of these proteins were shown in Figure 1.

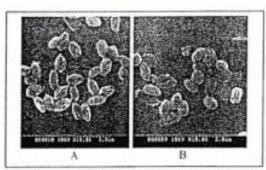


Fig 1.Crystal protein of SASU Bt isolate (A) and KATB Bt isolate (B)

B. Number of Total Viable Spores Count (TVSC) SASU dan KATB isolate-based product

TVSC of SASU-based product was in the range of 5.18 x 10⁴ - 8.98 x 10⁸ spores/ml and KATB-based product was 5.29 x 10⁵ - 7.34 x 10⁸ spores/ml. Content of growth media for culturing *B. thuringiensis* was very important. It can be seen in the media of mixture coconut water and soybéan soaking water, in SASU and KATB isolates, produced the highest spores. Compare with standard growth medium (nutrient broth), spores produced was similar. It indicated that crystal protein content was high, as well. Data was shown in Table 2.

- TABLE II
TOTAL VIABLE SPORES COUNT (TVSC) OF SASU AND KATB
ISOLATE-BASED PRODUCT

F/Z	TVSC (spore/ml)	
Treatment	Br - SASU	Bt - KATB
A. coconut water	2.22 x 10°	3.02 x 10°
B. soybean soaking water	3.14 x 10°	3.40 x 10
C. tofu liquid waste	5.18 x 10°	5.29 x 10°
D. mixture A and 8 (1:1,v/v)	8.98 x 10°	7.34 x 10°
E. mixture A and C (1:1, v/v)	3.67 x 10°	4.18 x 10 ^h
F. nutrient broth	3.56 x 10 ¹	5.09 x 10 ^x

C. Taxicity of SASU dan KATB isolate-based product

Mortality of insect pest (S.litura and P. xylostella) was the highest on media coconut water and soybean soaking water. Carbon and nutrient content of this media could be factor affect the growth of spores. The more number of spores consumed by larvae, the more number larvae will die, since Bt played a role as stomach poisons. Prabakaran et al [13] also showed coconut waste media could produce high number of spores, similar with production of spores in NYS medium.

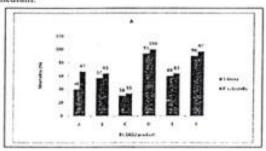
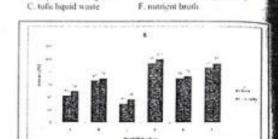


Fig. 2. Mortality of Speakgrova linear and Placella sydostellar on various media growth of Bi-SASU-based product.



D. mixture A and B (1:1,v/v)

E. mixture A and C (1:1, w/v)

А сосония жасе

B. soybean soaking water

Fig. 3. Mortality of Spodoptera litura and Plutella sylostella on variamedia growth of Bt- KATB-based product

Note:	
A. euconut water	D. mixture A and B (1:1,w/v)
B. soybean soaking water	E. misture A and C (1:1, v/v)
C. tofu liquid waste	F. nutrient broth

IV. CONCLUSIONS

Exploration of soil resulted 33 B. thuringiensis isolars which 21 isolates were toxic against P. xylostella and isolates were toxic towards S. litura. The highest mortality both S. litura and P. xylostella was occurred on treatment mixture of coconut water and soybean soaking water. It highest spores produced was Mixture of coconut water soybean soaking treatment indicated as the best media producing bio insecticide.

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REFERENCES

- [1] Aranda E. Sanchez, J., Peferoen, M. Guerrea, L., and Bravo A. E. Interaction of Bacillus thuringiensis crystal proteins with the miepithelial cells of Synchysters fragiperals (Lepidoptera:Noctain J. Invert. Pathol. 68: 203-212
- [2] Bravo, A., Sill, S. S., and Soberón, M. 2007. Mode of action Bacillus rharingiousis. Cry. and Cyt toxins and their potential, insect control. Toxicon. 49: 423-435.
- [3] Ferre, J. 2006. Toxicity and Mode of action of Boeiflus Muring Cry Toxin in Mediterranean Corn Borer, Sesamia managrio (Lefebvre). App. Environmental Microbiology, 72:4:2594-2600.
- [4] Martin, P. W and R. S. Travers. 1989. Worldwide abundance distribution of Bacillus theringsivesis isolates. Applied Environmental Microbiology, 55:2437-2442.
- [5] Pojiastuti, Y., Shin-ichiro Asano, Ken Sahara, Hisanon Bando Toshihiko lizuka. 1999a. Toxicity of Bucillus thuringiousis subunharensis crystal protein to Bosecx meet and Spodoptera lib 1999. J.Scric.Sci.Jpn. 68(3): 195-199.
- Feitelson, J. S., J. Payne, and L.Kim. 1992. Bacillus thuringies insects and beyond. Bio Technology, 10. 271-275.
 Kalshoven, L.G.E. 1981. The Pests of Crops in Indonesia. Rev8
- [7] Kalshoven, L.G.E. 1981. The Pests of Crops in Indonesia. Reve and Translated by P.A. Van der Lann. PT Ichtiar Baru-van Hot Jakarta. 701 p.

- Devi. P.S.Vimala, T. Ravinder, C. Jaidev. 2005. Cost-Effective Production Of Boxillar Murringionair By Solid-State Fermentation Journal of Invertebrate Pathology 88: 163–168.

 Chilcoti. C.N. and J.S.Pillai 1985. The use cocosus waste for production of Bocillos theringions's var. interferons. J. Misson of Production of Proceedings 1.
- production of Hacillus thuringiensis var. israelennis, J. Mircen. 1: 327-332
- 327-332 Thiery and Friction, 1997. Bacteria: Identification, Isolation, culture and preservation of entomopathogenic bacteria. In Manual of Techniques in Issect Pathology. Edited by L. Loccy. Academic Press San Diego USA.
- [41] Dubinge, HT and Rhodes, RD (1971). In: Alterabiat Control of Junetti and Asses, Eds. Burgess. HD and Huesey, NW. Academic Press, New York, pp.507-539.
- [12] Asono S., Yulix Pajiastuti, Ken SAHARA, Hisanovi BANDO, H. KIKUTA and Toshihiku BZUKA. 1998. Identification of cryl power from Bacillos thuringiensis strains which have activity toward Speakaptevic times. J Seric, Sci. Tjm. 60 (3): 237-242.
- [13] Prabakaran G, Hoti SL, Manonmans AM, and Balaramon K. 7007. Coconut water as a cheap source for the production of delta endotoxin of Bacillus thuringiensis var. israelensis, a mosquito control agent. Acta Trop. 105(1):35-8.