

Probiotic enterococci: current status

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- typical lactic acid bacteria (LAB)
- of importance in food and clinical microbiology
- ubiquitous microorganisms, but have predominant habitat in GIT of human and animals (Giraffa, 2002)





- former representative Group of the genus
 Streptococcus → fecal streptococci or
 Lancefield's group D streptococci
- later: separated from the genus Streptococcus on the basis of modern classification techniques and serological studies in the 1980s.
- the large conglomeration of streptococci was thus subdivided into three separate genera: Streptococcus, Lactococcus and Enterococcus





Introduction

- the typical pathogenic species (except S. thermophilus) remained in the genus Streptococcus and separated from the nonpathogenic and technically important species of the new genus Lactococcus (Devriese and Pot, 1995)
- the faecal ,streptococci' associated with the GIT of human and animals, with some fermented foods and a range of other habitats, constituted the new genus *Enterococcus*.



Introduction

- Two most prominent representatives of the genus Entrerococcus:
 - Enterococcus faecalis
 - Enterococcus faecium
- *E. faecalis* and *E. faecium* play the most important roles both in human disease and in fermented foods and probiotics (Franz et al., 1999)





Taxonomy and identification

Thirty Enterococcus species are currently recognized based on sequence analysis of 16S rRNA genes.





Taxonomy and identification

Genus Enterococcus can be distinguished into 7 Groups:

E. faecium-group: E. faecium, E. durans, E. hirae, E. mundtii, E. villorum, E. canis

E. avium-group: E. avium, E. malodoratus, E. pseudoavium, E. raffinosus, E. gilvus

E. gallinarum-group: E. gallinarum E. casseliflavus, E. flavescens

E. dispar-group: E. asini, E. dispar, E. pallens, E. hermanniensis, E. canintestini

E. saccharolyticus-group: E. saccharolyticus, E. sulfureus, E. saccharominimus, E. italicus, E. aquimarinus

E. cecorum-group: E. cecorum, E. columbae

E. faecalis-group: E. faecalis, E. haemoperoxidus, E. moraviensis, E. ratti

Physiological properties

	Growth at		Growth in the presence of					
Species	10°C	45°C	рН 9.6	6.5% NaCl	40% Bile	0.04% Sodium azide	Esculin hydrolysis	Group D antigen
E. asini	(+)	(+)	n.d.		+	n.d.	+	+
E. avium	V	+	+	V	V/+	n.d.	+	+
E. casseliflavus	+	+	+	V/+	+	+	+	+
E. cecorum		+	(+)	_	(+)		+	
E. columbae		n.d.	n.d.	_	(+)		+	
E. dispar	+		n.d.	+/-	+		+	
E. durans	+	+	+	+	+	+	+	(+)
E. faecalis	+	+	+	+-	+	+	+	+
E. faecium	+	+	+	+	+	+	+	v
E. flavescens	V/-	V/+	n.d.	+	+	+	+	+
E. gallinarum	+	+	+	+-	+	+	+	+
E. gilvus	+	+	n.d.	+	+	n.d.	+	+
E. haemoperoxidus	+		n.d.	+-	+	+	+	+
E. hirae	+	+	+	+-	+-	+	+	V
E, malodoratus	+		+	+	+	n.d.	+	+
E. moraviensis	+		n.d.	+-	+	+	+	+-
E. mundtii	+	+	+	-+-	+	+	+	+
E. pallens	+	+	n.d.	+	+	n.d.	+	+
E. phoeniculicola	n.d	n.d	n.d.			n.d.	n.d.	n.d.
E. porcinus	+	+	n.d.	+	n.d.	n.d.	+	+
E. pseudoavium	+	+	+	+/-	V/+	n.d.	+	
E. raffinosus	(+)	+	+	+	V/+	n.d.	+	n.d.
E. ratti	+	+	n.d.	+	n.d.	n.d.	+	(+)
E. saccharolyticus	+	+	n.d.	(+)	+	n.d.	+	_
E. solitarius	+	+	n.d.	+	+	n.d.	+	+
E. sulfureus	+-	_	n.d.	+	+	n.d.	+	
E. villorum	n.d.	n.d.	n.d.	+	+	+	+	n.d.

 Table 1
 Characteristic Physiological Properties of Validly Described Enterococcus Species

n.d., not determined, (+), weak positive; V, variable; +/-, differing reports in literature.



Environment

soil, surface waters, waste water, on plants,

- Gastrointestinal tract
- Foods
 - cheese
 - fermented vegetables



Habitat

Table 20.1 Numbers and predominant isolates of Enterococcus spp. in cheeses from Mediterranean countries

Cheese	Country of origin	Milk source	Enterococci in curd (log CFU/g)	Enterococci at end of ripening (CFU/g)	Predominant bacteria in end product (% of isolates)	Reference	
White-brined Greece Raw goat cheese or mixed g and ewes'		Raw goat milk or mixed goat and ewes' milk	4.0	6.7	L. plantarum (47%) ^b E. faecium (12%) L. paracasei subsp. paracasei (10%) E. faecalis (9%)	Litopoulou- Tzanetaki and Tzanetakis (1992	
Kefalotyri cheese	Greece	Ewes' milk, cow milk or mixed ewes' and goat milk	4.9	5.8	E. faecium (35.6%) L. plantarum (18.4%) L. casei subsp. casei (15.8%) E. durans (9.2%) pediococci (9.2%)	Litopoulou- Tzanetaki (1990)	
Teleme cheese	Greece	Pasteurised ewes' milk	n.r. ^a	n.r.	Lactobacilli Leuconostocs Enterococci	Tzanetakis and Litopoulou- Tzanetaki (1992)	
Orinotyri	Greece	Raw ewes' milk	n.r.	6.8	lactococci, enterococci, leuconostocs	Prodromou <i>et al.</i> (2001)	
La Serena ewes' milk	Spain	Raw ewes' milk	6.2	7.2	Lactobacilli Leuconostocs Enterococci	Del Pozo <i>et al.</i> (1988)	
Manchego	Spain	Raw ewes' milk	n.r.	n.r.	Enterococci	Ordoñez et al. (1978)	
Cebreiro	Spain	Raw cow milk	n.r.	6.5	E. faecalis (30.1%) E. faecalis (var liquifaciens) (11.9%) Lact. lactis (19.0%) W. (Leuc.) paramesenteroides (7.9%)	Centeno <i>et al.</i> (1996)	

Leuc. mesenteroides subsp. mesenteroides (6.3%) E. faecium (4.8%)

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Table 20.1 Continued

Cheese	Country of origin	Milk source	Enterococci in curd (log CFU/g)	Enterococci at end of ripening (CFU/g)	Predominant bacteria in end product (% of isolates)	Reference
San Símon cheese	Spain	Raw cow milk	5-6	6–7	E. faecalis, E. faecium, E. durans, Staph, spp. Micrococcus spp	García <i>et al.</i> (2002)
Tetilla cheese	Spain	Raw cow milk	n.r.	7.3	E. faecalis, L. casei subsp. casei, Leuconostoc mesenteroides subsp. mesenteroides	Menéndez <i>et al.</i> (2001)
Caprino d' Aspromonte	Italy	Raw or heated goat milk	4-6	5-7	enterococci, lactobacilli, mesophilic and thermophilic cocci	Caridi et al. (2003)
Montasio	Italy	Raw or heated cow milk	4	ca. 5–7	S. thermophilus, E. durans, E. faecalis, E. faecium	Marino et al.
Serra cheese	Portugal	Raw ewes' milk	n.r.	n.r.	Leuc. lactis, Lact. lactis, Leuc. mesenteroides subsp. mesenteroides/ dextranicum E. faecium	Macedo <i>et al.</i> (1995)
Picante da Beira Baixa cheese	Portugal	Mixture of raw goat and ewes' milk	n.r.	n.r.	E. faecium, E. faecalis, E. durans, L. plantarum, L. paracasei	Freitas <i>et al.</i> (1995)

^a n.r. = not reported: ^b : L. = Lactobacillus: E. = Enterococcus: Lact. = Lactococcus: Leuc. = Leuconostc; W. = Weissella, Staph. = Staphylococcus



Use of enterococci as probiotics

E.faecium SF68

diarrhoea treatment

- Causido[®] culture
 - consists of S. thermophilus and E. faecium
 - hypocholsterolaemic effect

Virulence factors

 Table 20.2
 Virulence factors which may be present in some Enterococcus strains, and (suggested) association with stage of virulence

Virulence determinant	(Suggested) association with stage of virulence				
Aggregation substance (AS)	Adhesion to eukaryotic cells (adhesin) promotes colonisation				
	Invasion of eukaryotic cells (invasin)				
	Adhesion to extracellular matrix proteins (may promote translocation)				
	Increases survival in immune cells (evasion of host immune response)				
Cytolysin (Cyl)	Eukaryotic cell toxin				
	Lyses immune cells (evasion of host immune response)				
Gelatinase (Gel)	Can hydrolyse various biological peptides, e.g. collagens and fibrin (role in translocation?)				
	of host innate immune response)				
Enterococcal surface protein	Adhesin, promotes colonisation				
(Esp _{fs} and Esp _{fm})	Exhibits characteristics of MSCRAMMs – role in evasion of immune response?				
Adhesin to collagen of E. faecalis	Adhesion to extracellular matrix proteins				
(Ace) or E. faecium (Acm)	(may promote translocation)				
	Exhibits MSCRAMM characteristics: role in evasion of immune response?				
Endocarditis antigen from <i>E. faecalis</i> or <i>E. faecium</i> (EfaA _{fs} and EfaA _{fm})	Adhesin: role in endocarditis				
Hyaluronidase	Degrades hyaluronic acid, a major extracellular matrix constituent: role in translocation?				
Pheromones	Cause inflammation, induce superoxide production				
E. faecium secreted antigen (Sag)	Adhesion to extracellular matrix proteins				
Superoxide and hydrogen peroxide	May cause cell/DNA damage, improves colonisation				
Capsule	Evasion of host immune response				

Incidence of virulence factors

	% incidence of virulence factor of enterococci from foods							
For <i>F. faecium</i> strains:	Fermented sausages ¹	Fermented sorghum ²	Cheese/other foods ³	Cheese ⁴	Foods ⁵			
	(n=55)	(n=22)	(n=48)	na	(11)			
EfaA	100	90.9	nd	na	82			
GelE	5.5	0	0	na	0			
AS ^a	5.5	0	0	na	0			
Esp	5.5	9.1	2.1	na	0			
Haemolysin or cytolysin ^b	nd	0	8.3	na	0			
For <i>E. faecalis</i> strains:	(n=5)	na	(n=47)	(n=10))	(9)			
EfaA	100	na	nd	100	89			
GelE	100	na	48	70	78			
AS	100	na	48.9	100	67			
Esp	99.3	na	36.2	50	33			
Haemolysin or cytolysin	nd	na	21.3	50	44			

Table 20.3 Reported incidences of virulence factors among E. faecalis and E. faecium strains from foods

nd: not determined: na: not applicable as no strains of the indicated species were investigated

a: aggregation substance: data pertaining to information on either the clumping phenotype or presence of the asal gene

b: haemolysin or cytolysin: data pertaining to information on either the presence of cytolysin genes or lysis on blood agar

1 data adapted from Martin et al. (2005)

2 data adapted from Yousif et al. (2005)

3 data adapted from Franz et al. (2001)

4 data adapted from Majhenic et al. (2005)

5 data adapted from Eaton and Gasson (2001)

Incidence of antibiotic resistance among food enterococci

Resistance to antibiotic	% incidence of resistance of enterococci from foods							
	Fermented sausages ¹	Fermented sorghum ²	Cheese/other foods ³	Retail meats (turkey/chicken) ⁴	Retail meats (pork/beef) ⁴	Produce ⁵	Ready to eat foods ⁶	
For E. faecium strains:	(n=55)	(n=22)	(n=48)	(n=213/245)	(n=114/245)	(n=97)	(n=47)	
Ampicillin	30.9	0	0	54/23	4.4/2.8	nd	6.4	
Penicillin	58.2	9.1	45.8	53/20	9.6/8.7	7	12.8	
Erythromycin	56.4	31.8	27.1	87/43	60/39	10	29.8	
Tetracycline	29.1	0	6.3	0.9/0.4	0.9/0.4	29	17	
Chloramphenicol	20	0	10.4	41/22	7/19	5	2.1	
Ciprofloxacin	54.5	13.6	56.3	nd	nd	28	25.5	
Gentamicin	0	0	2.1	nd	nd	0	nd	
Streptomycin	nd	0	4.2	nd	nd	3	nd	
Vancomycin	1.8	13.6	2.1	nd	nd	0	0	
Rifampin	69.1	nd	nd	nd	nd	nd	19.1	
For E. faecalis strains:	(n=5)	na	(n=47)	(n=110/51)	(n=161/66)	(n=38)	(n=52)	
Ampicillin		na	2.1	nd	nd	nd	0	
Penicillin		na	12.8	0	0/0	0.6/0	0	
Erythromycin	93.3	na	63.8	3	42/33	8.1/4.5	26.4	
Tetracycline	86.7	na	44.7	0	94/67	89/39	41.5	
Chloramphenicol	93.3	na	31.9	3	0/0	3.1/0	24.5	
Ciprofloxacin	46.7	na	27.7	5	0/0	0.6/0	0	
Gentamicin	0	na	25.5	0	nd	nd	nd	
Streptomycin	nd	na	46.8	0	nd	nd	nd	
Vancomycin	0	na	0	0	nd	nd	0	
Rifampin	100	na	nd	nd ·	nd	nd	11.3	

Table 20.4 Reported incidences of antibiotic resistances among E. faecium and E. faecalis strains isolated from foods

nd: not determined; na: not applicable as no E. faecalis strains were investigated

1 data adapted from Martin et al. (2005)

2 data adapted from Yousif et al. (2005)

3 data adapted from Franz et al. (2001)

4 data adapted from Hayes et al. (2003)

5 data adapted from Johnston and Jaykus (2004)

6 data adapted from Baumgartner et al. (2001

Conclusions

- it is still hard to make decision whether enterococci are risk for consumer or not.
- many enterococci occur in large numbers in foods; however there many strains can harbour (multiple) virulence determinants and/or antibiotis resistance genes.
- The ,host factor' appears to play a key or determining role in the establishment of an infection with enterococci.

Thank you!