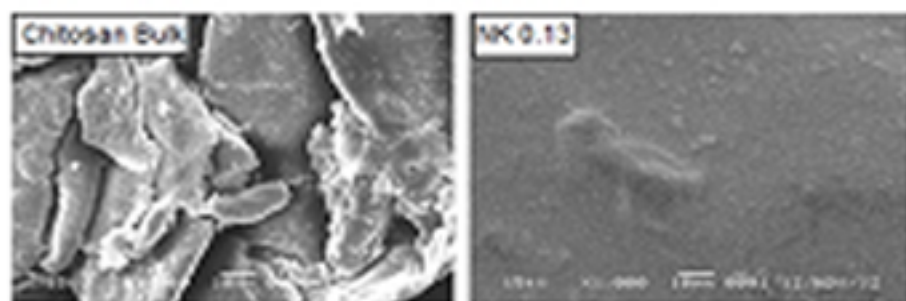
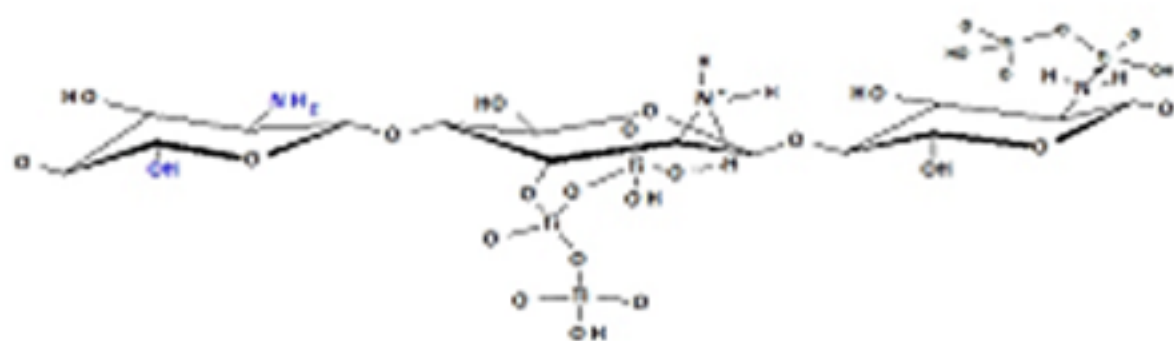


# Indonesian Journal of Chemistry

Vol. 14, No. 3, November, 2014



**Editor in Chief**

**Prof. Dr.rer.nat. Nuryono, MS** Email: nuryono\_mipa@ugm.ac.id  
 Department of Chemistry, Universitas Gadjah Mada  
 Sekip Utara, Yogyakarta Indonesia 55281, Tel/Fax (0062-274)-545188.  
 Website : <http://www.ijc.chemistry.ugm.ac.id>  
 Email : [ijc@ugm.ac.id](mailto:ijc@ugm.ac.id) or [ijcugm@yahoo.com](mailto:ijcugm@yahoo.com)

**Vice Editor in Chief**

Prof. Dr. Harno Dwi Pranowo, M.Si Email: harnodp@ugm.ac.id or harnopranowo@yahoo.com  
 Prof. Dr. Mudasir, M.Eng Email: mudasir@ugm.ac.id or m\_mudasir@hotmail.com

**Editorial Board**

Prof. Dr. Karna Wijaya, M.Eng. (*Physical/Material Chemistry*)  
 Drs. Iqmal Tahir, M.Si. (*Computational/Physical Chemistry*)  
 Dr. Ria Armunanto, M.Si. (*Computational/Physical Chemistry*)  
 Dr. Tri Joko Raharjo, M.Si. (*Biochemistry/Bioanalysis*)  
 Dr. Nurul Hidayat Aprilita (*Analytical/Environmental Chemistry*)

**Advisory Editorial Board**

Dr. Brian Williams (Adelaide University Australia)	Prof. Dr. Muhammad Idris Saleh (University Sains Malaysia)
Prof. Dr. Dr. Hc. Bernd M Rode (University of Innsbruck, Austria)	Dr. Hery Haerudin (Pertamina, Indonesia)
Dr. Dirk Bax (Utrecht University, Netherlands)	Prof. Dr. Iip Izul Falah (Universitas Gadjah Mada, Indonesia)
Prof. Dr. M. Gross (Louis Pasteur University, France)	Prof. Sri Juari Santosa, M.Eng, Ph.D. (Universitas Gadjah Mada, Indonesia)
Prof. Dr. Hardjono Sastrohamidjojo (Universitas Gadjah Mada, Indonesia)	Prof. Dr. Endang Tri Wahyuni, MS (Universitas Gadjah Mada, Indonesia)
Prof. Dr. David. St. C. Black (University of New South Wales, Australia)	Prof. Dr. Bambang Rusdiarso, DEA (Universitas Gadjah Mada, Indonesia)
Prof. Dr. Max Lu (University of Queensland, Australia)	Prof. Dr. Wega Trisunaryanti, M.S., Ph.D. Eng (Universitas Gadjah Mada, Indonesia)
Prof. Dr. Buchori (Bandung Institute of Technology, Indonesia)	Prof. Dr. Sabirin Matsjeh (Universitas Gadjah Mada, Indonesia)
Prof. Dr. Abdul Ra'uf Pathong (Hasanudin University, Indonesia)	Dr. Winarto Haryadi, M.Si. (Universitas Gadjah Mada, Indonesia)
Prof. Dr. Naoki Yoshioka (Keio University, Japan)	
Assoc. Prof. Dr. Wan Ahmad Kamil Mahmood (University Sains Malaysia)	

**Administrator**

Dr. Akhmad Syoufian  
 Warakustarti Listyariwangi, A.Md  
 Djoko Prihandono

Robby Noor Cahyono, S.Si., M.Sc  
 Nurzanah Hidayanti, A.Md

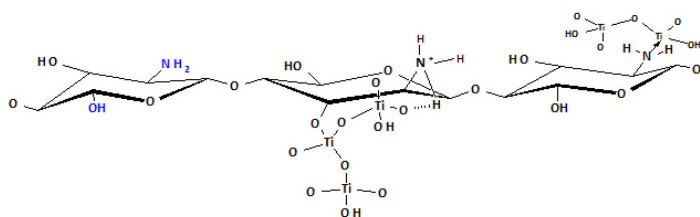
**Aims and Scope**

Indonesian Journal of Chemistry is an international journal covering all aspects of Chemistry, including Chemical Education and Chemical Engineering. The journal publishes original research papers, short communications, and review articles, and has been indexed by SCOPUS since 2012. The paper published in this journal implies that the work described has not been, and will not be published elsewhere, except in abstract, as part of a lecture, review or academic thesis.

Indonesian Journal of Chemistry (ISSN 1411-9420) is published by the Department of Chemistry, Faculty of Mathematics and Natural Sciences, Universitas Gadjah Mada, Sekip Utara, Yogyakarta, Indonesia. All orders accompanied by payment should be sent directly to The Department of Chemistry, Universitas Gadjah Mada. Annual subscription rate is IDR 200,000.00 (Java-Bali), IDR 250,000.00 (outside Java-Bali) (shipping included). Reprint order price is IDR 50,000.00 per article (shipping not included). Customers may make payments by transfer on **Mandiri Universitas Gadjah Mada (Dr. Nuryono, Account Nr. 137-00-1061199-0)**.

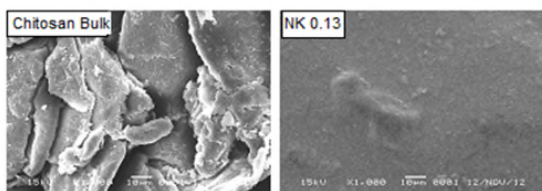
## CONTENTS

<b>Photocatalytic Decolorization Study of Methyl Orange by TiO<sub>2</sub>-Chitosan Nanocomposites</b> by Imelda Fajriati, Mudasir, and Endang Tri Wahyuni	209-218
<b>Simulation of Pollutant Gas Dispersion on Case Study of <i>Lontar 3</i> Coal Fired Power Plant in Addition into 1 X 660 MW Capacity in Kemiri, Tangerang District, Banten Province</b> by Eko Sugiharto, Taufik Abdillah Natsir, and Abdul Rozaq	219-225
<b>Adsorption Isotherm Studies on Acid Orange-10 Dye Removal Using Cerium Dioxide Nanoparticles</b> by Harry Budiman and Oman Zuas	226-232
<b>Production of Reducing Sugar from Cassava Solid Waste by Simultaneous Ultrasonication and Acid Hydrolysis</b> by Wasinton Simanjuntak, Heri Satria, and Nurul Utami	233-238
<b>Theoretical Analysis of Interaction Energy in Alginate-Capped Gold Nanoparticles Colloidal System</b> by Foliatini, Yoki Yulizar, and Mas Ayu Elita Hafizah	239-245
<b>Fast Swelling Superabsorbent Hydrogels Starch Based Prepared by Gamma Radiation Techniques</b> by Erizal, Dian Pribadi Perkasa, Basril Abbas, Sudirman, and Sulistioso G.S.	246-252
<b>The Effect of Caramelization and Carbonization Temperatures toward Structural Properties of Mesoporous Carbon from Fructose with Zinc Borosilicate Activator</b> by Tutik Setianingsih, Indriana Kartini, and Yateman Arryanto	253-261
<b>Hydrogel Based on Crosslinked Methylcellulose Prepared by Electron Beam Irradiation for Wound Dressing Application</b> by Ambyah Suliwarno	262-268
<b>Oxidation and Acetylation of Ursolic and Oleanolic Acids Isolated from <i>Fragraea fragrans</i> Fruits: Antiproliferation of P388 Leukemia Cells</b> by Dasril Basir, Julinar, Eva Agustriana, and Budi Untari	269-276
<b>Cytotoxic Isobractatin (Prenylated Xanthone) Epimer Mixture of <i>Garcinia eugenifolia</i></b> by Sri Hartati, I Ketut Triyono, and Sri Handayani	277-282
<b>Synthesis and Thermal-Stability Study of Polybutylene Itaconate Modified with Divinyl Benzene and Glycerol</b> by Atmanto Heru Wibowo, Ninis Makhnunah, Deny Irawati, Candra Purnawan, Nanik Dwi Nurhayati, and Henning Storz	283-289
<b>Di-(2-Ethylhexyl)Phthalate and Pyranon Derivated from Endophytic Fungi <i>Penicillium</i> sp the Leave of Kunyit Putih (<i>Curcuma zedoaria</i>)</b> by Muharni, Fitriya, Milanti Okta Ruliza, Dwi Anjar Susanti, and Elfita	290-296
<b>Major Anthocyanin Pigments in the <i>Ficus padana</i> Fruits: HPLC-DAD-ESI-MS Identification and Antioxidant Activity</b> by Daimon Syukri, Djaswir Darwis, and Adlis Santoni	297-303
<b>Isolation of Bioactive Compounds from <i>Aspergillus terreus</i> LS07</b> by Rizna Triana Dewi, Sanro Tachibana, Puspa Dewi, L.B.S. Kardono, and Muhammad Ilyas	304-310
<b>Short Communication: Synthesis and Characterization of [Fe(Picolinate)<sub>3</sub>][MnNi(Oxalate)<sub>3</sub>].CH<sub>3</sub>OH Polymeric Complex</b> by Fahimah Martak, Djulia Onggo, Ismunandar, and Agung Nugroho	311-314

CONTENTS (*Continued*)**Cover picture :**

See Imelda Fajriati et al., page 212 & 215

The hypothetic interaction between  $\text{TiO}_2$  and chitosan (above) & SEM images of chitosan bulk and  $\text{TiO}_2$ -chitosan nanocomposite (NK 0.13) (below)



### Organization of Manuscripts

The submitted manuscripts are classified into three categories: original paper which presents original works in detail, notes and/or short communications which present novel and/or valuable information and reviews which present a general survey of specialized subject in chemistry. All manuscripts should be written in concise and clear English and suggested to be typed with full justification, singled spaced for abstract, references, figure captions and tables (tables and figures should be typed on separate sheets at the end of the manuscript): double spaced for text, in Arial 11, using no more than 20 pages for original papers, 10 pages for notes and/or short communication and 30 pages for reviews. Left and right margins should be 3.0 cm length. The title should be typed in Arial 12 bold. The names of the authors and addresses at which the research was done, including postal code, should appear under the title. Use Arabic number typed as superscript to link authors to their addresses and asterisk to indicate the author(s) to whom correspondence should be addressed. Main headings (Abstract, Introduction, Experimental, Results and Discussion, Conclusions) are typed in bold and capital italics. Type all headings aligned left and lower case except the first letter of the first word or any proper name. The manuscripts should be written in English or in Indonesian, but the abstract must be written in English and contains no more than 200 words followed by 3-5 keywords. All references should be prepared according to the following style: **Article in Journal:** Barrer, R.M. and Craven, R.J.B., 2000, *Phys. Chem.*, 2, 545–550. **Chapter in a Book:** Rao, C.N.R, and Rao, K.J., "Ferroics" in *Solid State Chemistry Compounds*. Eds. Cheetam, A.K., and Day, P, P., Clarendon Press, Oxford, 1992, 281-96. **Whole Book:** Barrer, R.M. and Craven, R.J.B., 1986, *New Developments in Zeolite Science and Technology*, ed. Murakame, Y, Iijima, A. and Ward, J.W., Kodansha, Tokyo, p.521. Text references to the literature must be numbers in square brackets. Journals titles should be abbreviated according to the Chemical Abstract Service Source Index (CASSI).

Template file of the article could be downloaded in the website:

<http://www.ijc.chemistry.ugm.ac.id/author/ijctemplate.doc>

### Acknowledgment

All other contributing individuals should be typed and acknowledged at the end of the manuscript.

### Submission of Manuscripts

Please submit your article by online submission at <http://pdm-mipa.ugm.ac.id/ojs/index.php/ijc/> or via email: [ijcugm@yahoo.com](mailto:ijcugm@yahoo.com) or [ijc@ugm.ac.id](mailto:ijc@ugm.ac.id)

An IDR 800,000.00 (Java-Bali) and IDR 1,000,000.00 (outside Java-Bali) fee per article may be paid for papers published in this journal. This fee includes automatic subscription of the journal for one volume (3 issues). Subscription fee charged to overseas is USD 100 for 1 (one) copy of the journal and the article in pdf format. Articles of more than 7 pages (after layout) are subject to additional cost of IDR 75,000.00 per extra page. Color printing fee is IDR 100,000.00 per page.

Author may reproduce/republish portions of their published contribution without seeking permission from the Dept. of Chemistry, Universitas Gadjah Mada (UGM), provided that any such republication is accompanied by an acknowledgement in the form: (Original Citation-Reproduced by Permission of The Dept. of Chemistry, Universitas Gadjah Mada).

### Advertising

Inquires concerning advertising should be addressed to Dr. Tri Joko Raharjo, M.Si, Department of Chemistry, Universitas Gadjah Mada, Sekip Utara, Yogyakarta, Indonesia. Email: [trijr\\_mipa@ugm.ac.id](mailto:trijr_mipa@ugm.ac.id) or [trijrarin@yahoo.com](mailto:trijrarin@yahoo.com)

## DI-(2-ETHYLHEXYL)PHTHALATE AND PYRANON DERIVATED FROM ENDOPHYTIC FUNGI *Penicillium* sp THE LEAVE OF KUNYIT PUTIH (*Curcuma zedoaria*)

Muharni<sup>1,\*</sup>, Fitrya<sup>2</sup>, Milanti Okta Ruliza<sup>1</sup>, Dwi Anjar Susanti<sup>1</sup>, and Elfita<sup>1</sup>

<sup>1</sup>Department of Chemistry, Faculty of Mathematics and Natural Sciences, Sriwijaya University, Jl. Raya Palembang Prabumulih Km 32, Indralaya, Ogan Ilir, South Sumatera 30662, Indonesia

<sup>2</sup>Department of Pharmacy, Faculty of Mathematics and Natural Sciences, Sriwijaya University, Jl. Raya Palembang Prabumulih Km 32, Indralaya, Ogan Ilir, South Sumatera 30662, Indonesia

Received October 24, 2013; Accepted July 18, 2014

### ABSTRACT

Two compounds from cultivation of the endophytic fungi *Penicillium* sp of leaves of kunyit putih (*Curcuma zedoaria*) have been isolated. The endophytic fungus was cultivated on 5 L of Potatos Dextrose Broth (PDB) medium at room temperature (no shaking) for 3 weeks. The cultures were extracted with ethyl acetate to afford 3.0 g of residue after removal of the solvent under reduced pressure. The extract was separated and purified by silica gel column chromatography (CC) and afforded two pure compounds as colorless oily liquid (compound 1) and yellow crystal (compound 2). The structure of these compounds were characterized by detailed UV, IR, and NMR spectroscopic analysis and compound 1 as well as comparison with the reported data. Base on spectra analysis the compound 1 was determined as Di-(2-ethylhexyl)phthalate and compound 2 as 5-(4'-ethoxy-2'-hydroxy-5'-methyl-2',3'-dihydrofuran-3'-il (hydroxy) methyl-4-isopropyl-3-methyl-2-pyran-2-on). Compound 1 is not new compound, but it is new for endophytic fungus from *C. zedoaria* and compound 2 is new compound.

**Keywords:** endophytic fungi; *Penicillium* sp; *Curcuma zedoaria*

### ABSTRAK

Telah dilakukan isolasi dua senyawa dari kultifit jamur endofitik *Penicillium* sp dari daun kunyit putih (*Curcuma zedoaria*). Jamur endofit dikultur dalam 5 L medium Potatos Dextrose Broth (PDB) pada suhu kamar (keadaan statis) selama 3 minggu. Kultur kemudian diekstraksi dengan etil asetat dan dipekatkan dengan rotary evaporator sehingga didapatkan ekstrak pekat etil asetat 3,0 g. Ekstrak dipisahkan dan dimurnikan dengan kromatografi kolom menggunakan fasa diam silika gel dan didapatkan dua senyawa murni berupa cairan minyak bening (senyawa 1) dan kristal kuning (senyawa 2). Struktur senyawa hasil isolasi ditentukan berdasarkan analisis data spektroskopi UV, IR, dan NMR, dan senyawa 1 juga dikonfirmasi dengan membandingkan data yang telah dilaporkan. Berdasarkan analisis data spektroskopi disimpulkan senyawa 1 adalah Di-(2-ethylhexyl)phthalate dan senyawa 2 adalah 5-(4'-etoksi-2'-hidroksi-5'-metil-2',3'-dihidrofuran-3'-il (hidroksi) metil-4-isopropil-3-metil-2-piran-2-on). Senyawa 1 bukan merupakan senyawa baru, tetapi untuk pertama kalinya ditemukan pada jamur endofitik pada *C. zedoaria* dan senyawa 2 merupakan senyawa baru.

**Kata Kunci:** jamur endofitik; *Penicillium* sp; *Curcuma zedoaria*

### INTRODUCTION

Endophytic microorganisms that reside in the tissues of living plants and may produce secondary metabolites of biologically active [1]. Novel antibiotics, antimycotics, immunosuppressants, anticancer compound are only a few examples of what has been found after the isolation, culture and purification and characterization of some choice endophytes in the recent past. Isolation of their bioactive secondary metabolites of endophytic fungus from plant could be

selected mainly something on ethnobotanical history [2].

*Curcuma zedoaria*, a medicinal tuber plant belonging to the family Zingiberaceae has been used in the traditional system of medicine [3]. These plants were used for curing stomach diseases, toothache, blood stagnation, leucoderma, tuberculosis, enlargement of spleen, and for promoting menstruation in traditional medicine in Asia [4]. Antiinflammatory activity [5], antiulcer activity [6], and antimicrobial effect [7], of this plant rhizome have been reported.

\* Corresponding author.

Email address : muharnimyd@yahoo.co.id

In our research of endophytic fungus, many bioactive compounds and new compounds were isolated [8-9]. In this paper we reported the isolation and structural identification one known compound namely Di-(2-ethylhexyl)phthalate (**1**) and one new compound as 5-(4'-ethoxy-2'-hydroxy-5'-methyl-2',3'-dihydrofuran-3'-yl (hydroxy) methyl-4-isopropyl-3-methyl-2-pyran-2-on) (**2**) of *Penicillium* sp from the leaves of *C. zedoaria*. *Penicillium* species isolated as endophytic usually be found in plants zingiberaceae [10] and meliaceae family, although in marine organisms, three meroterpenes preaustinoids, A, B, A1, A2, and B1 have been reported to be isolated from *Penicillium* sp associated with the *Melia azedarach* [11]. *Penicillium* commune from the semi-mangrove plant *Hibiscus tiliaceus*, have been isolated one new compound 1-O-(2,4-dihydroxy-6-methylbenzoyl)-glycerol along with thirteen known products including 1-O-acetylglycerol, N-acetyl tryptophan, 3-indolylacetic acid methyl ester, 1-(2,4-dihydroxy-3,5-dimethylphenyl)ethanone, 2-(2,5-dihydroxy phenyl)acetic acid, (4R,5S)-5-hydroxyhexan-4-olide, thymidine, uracil, thymine, ergosterol,  $\beta$ -sitosterol,  $\beta$ -daucosterol, and ergosta-7,22-dien-3 $\beta$ ,5 $\alpha$ ,6 $\beta$ -triol [12].

## EXPERIMENTAL SECTION

### Materials

The leaves of kunyit putih were collected on May 2013 from the Indralaya, Ogan Ilir, South Sumatra. Material for isolation endophytic fungi: ethanol 70%, NaOCl, chloramphenicol, potato dextrose broth (PDB), potato dextrose agar (PDA), silica gel 60 (70-230 mesh), thin layer chromatography (TLC) from Merck (Art.5554) silica gel 60 F<sub>254</sub>, *n*-hexane, ethyl acetate, and methanol. The organic solvents were used from distilled technical grade.

### Instrumentation

The apparatus in the research were counter colony, autoclave, incubator, water bath, microscope, magnetic hotplate, UV lamp, column chromatography and generally apparatus in organic and microbiology laboratory, melting point was determined using Fisher John Apparatus. UV spectra were determined with Varian Conc 100 instrument. IR spectra were determined on FTIR-Perkin Elmer-Spectrum One and NMR spectra were recorded at 500 MHz (<sup>1</sup>H) and 125 MHz (<sup>13</sup>C) on JEOL JNM ECA-500 spectrometer, UV light at  $\lambda$  254 nm and 365 nm.

### Procedure

#### Isolation of endophytic fungus

The leaves sample was washed before it was processed and surface sterilized in 70% ethanol for 3 min and 0.5% NaOCl for 1 min and rinsed thoroughly with sterile distilled water. The segment sample placed on petri-plates containing potato dextrose agar medium (PDA) (200 g potato, 20 g dextrose, and 15 g agar in 1 L of H<sub>2</sub>O, supplemented with 100 mg/L of chloramphenicol to suppress bacterial growth). The plates were incubated at 25  $\pm$  2 °C until fungus growth appeared. The plant segments were observed once a day for the growth of endophytic fungus. Colony fungus showed difference characteristic furthermore to pure with the plated segments were immediately transferred into new PDA plates and then subcultured until pure cultures were obtained [13].

#### Identification of the endophyte

The endophytic fungal strain was identified by the morphological method. The morphological examination was performed by scrutinizing the fungal culture, the mechanism of spore production, and the characteristics of the spores. All experiments and observations were repeated at twice [14].

#### Cultivation of pure fungal strain

The purified fungus (a small park) was transferred under sterile conditions to the PDB medium. For chemical investigations, the fungal strains were static cultivated into 15 flasks (1 L each) containing 400 mL of PDB medium for 3 weeks at room temperature [12-14].

#### Extraction, isolation, purification, and structure elucidation

Fungus in the 3 weeks cultures were vacuum-filtered and the filtrate fractionated thrice by liquid-liquid partition with ethyl acetate (1:1). Then the solvent phase was evaporated under reduced pressure using rotary vacuum evaporator at 40 °C to produce the ethyl acetate fraction of liquid cultures. The EtOAc fraction (3.0 g) was preabsorbed on silica gel and then purification by column chromatography (silica gel, eluted *n*-hexane : EtOAc = 5:5 – 1:9), EtOAc 100%, EtOAc : MeOH = 9:1 – 1:9 and MeOH 100%). Based on detection by TLC using the eluent system, to give five fractions F1–F5. The 1<sup>st</sup> fraction to yield compound **1** (20 mg). Furthermore, fraction 2<sup>nd</sup> (0.2 g) was rechromatographed using the same method (silica gel, eluted with EtOAc : MeOH (8:2 – 1:9) and MeOH (100%) to yield three fractions F2.1 – F2.3. Fraction F2.1 to yield pure compound **2** (10 mg). The molecular structure of compounds were established on the basis

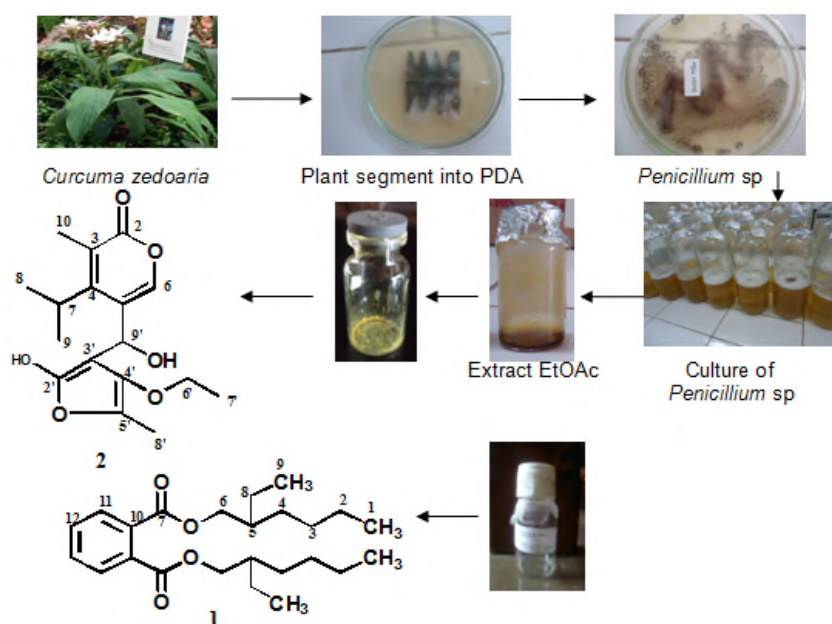


Fig 1. Isolation of the compounds from ethyl acetate extract of *Penicillium sp* from the leaves of *C. zedoaria*

Table 1.  $^1\text{H-NMR}$  (500 MHz) and  $^{13}\text{C-NMR}$  (125 MHz), spectral data of compound 1, recorded in  $\text{CD}_3\text{OD}$

Carbon no.	$\delta_{\text{H}}$ ppm ( $\Sigma\text{H}$ , multiplicity, $J$ in Hz)		$\delta_{\text{C}}$ (ppm)	
	1	1*	1	1*
1	0.89 (3H, <i>t</i> )	0.84 (3H, <i>t</i> , 4.3)	14.1	14.1
2	1.30 (2H, <i>m</i> )	1.23 - 1.44 (2H, <i>m</i> )	23.1	24.8
3	1.29 (2H, <i>m</i> )	1.23 - 1.44 (2H, <i>m</i> )	29.0	22.7
4	1.38 (2H, <i>m</i> )	1.23 - 1.44 (2H, <i>m</i> )	23.8	29.5
5	1.67 (1H, <i>m</i> )	2.60 (1H, <i>m</i> )	38.8	40.8
6	4.21 (2H, <i>m</i> )	4.15 (2H, <i>m</i> )	68.1	65.2
7			167.8	171.1
8	1.34 (2H, <i>m</i> )	2.30 (2H, <i>dq</i> , 4.3)	30.4	29.7
9	0.91 (3H, <i>t</i> )	0.93 (3H, <i>t</i> , 4.3)	11.0	20.8
10			132.5	124.8
11	7.69 (1H, <i>dd</i> , 5.9 - 3.3)	6.96 (1H, <i>dd</i> , 6.3 - 2.2)	128.8	119.0
12	7.51 (1H, <i>dd</i> , 5.9 - 3.3)	7.11 (1H, <i>dd</i> , 6.3 - 2.2)	130.9	132.6

1 [19]

of spectroscopic analysis including UV, IR,  $^1\text{H-NMR}$ ,  $^{13}\text{C-NMR}$ , DEPT, HMQC, HMBC, and COSY.

## RESULT AND DISCUSSION

The fungus strain was identified as *Penicillium sp.* *Penicillium* species isolated as endophytes were obtained from several plant species such as, *Melia azedarach* [15-16]. Zingiberaceae family [14] meliaceae family, although in marine organisms as the semi-mangrove plant *Hibiscus tiliaceus* [17]. Fungus *Penicillium sp* after that cultivated on 5 L of PDB medium and then extracted with ethyl acetate to afford 3.0 g of residue. The extract (3.0 g) was separated by column chromatography to yield compound 1 (20 mg) and compound 2 (10 mg). The isolation of the compounds

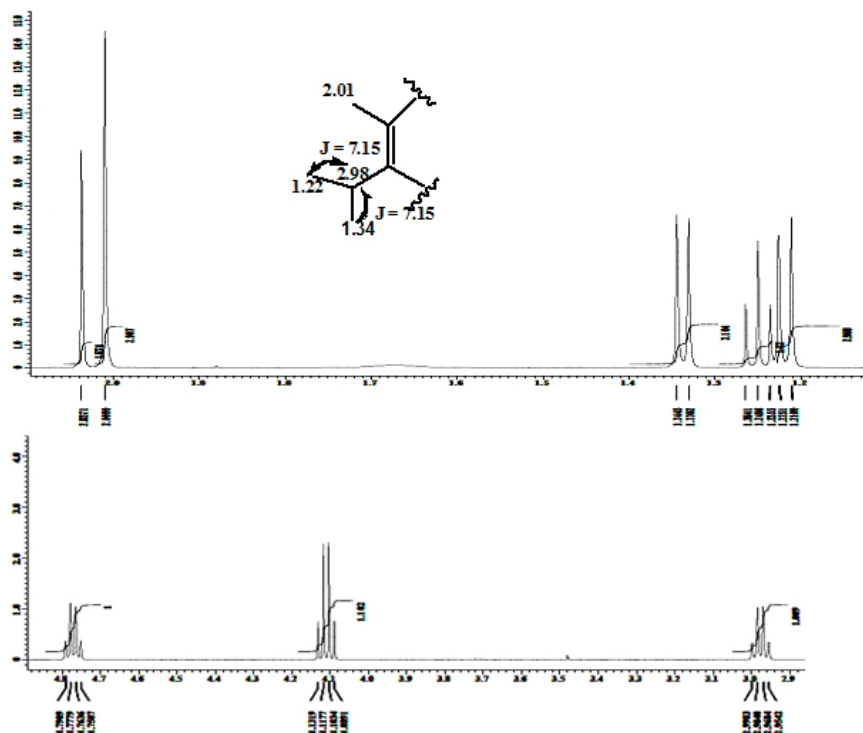
from ethyl acetate extract of *Penicillium sp* from the leaves of *C. zedoaria* described in Fig. 1.

Compound 1 was obtained as colorless oil liquid. The UV spectra of 1 exhibited absorption at  $\lambda_{\text{max}}$  nm : 206, 225, and 274. The bathochromic shift in addition of NaOH showed there is no wave length shift, it can concluded that there was no phenolic group. The IR spectrum showed the functional group such as carbonyl ester ( $1722\text{ cm}^{-1}$ ), C=C aromatic ( $1598\text{--}1462\text{ cm}^{-1}$ ), C-O ( $1273\text{ cm}^{-1}$ ), C-H aromatic ( $3070\text{ cm}^{-1}$ ), and C-H aliphatic ( $2927\text{--}2860\text{ cm}^{-1}$ ). The  $^1\text{H-NMR}$  data (Table 1) disclosed the presence of two protons as AB spin system at  $\delta_{\text{H}}$  7.69 (1H, *dd*, 5.9 & 3.3 Hz) and 7.51 (1H, *dd*, 5.9 & 3.3 Hz) that characteristic for aromatic proton at ortho substituted ring. The proton signal at  $\delta_{\text{H}}$  4.21 ppm (2H, *m*) is assigned to a methylene group geminal to the ester



**Table 2.**  $^1\text{H-NMR}$  (500 MHz) and  $^{13}\text{C-NMR}$  (125 MHz), spectral data of compound **2** recorded in  $\text{CDCl}_3$ 

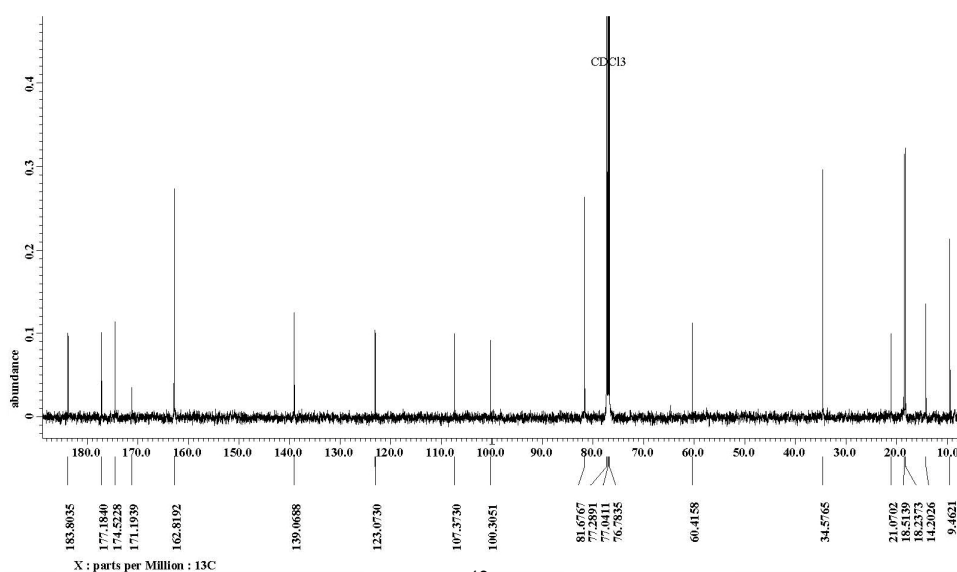
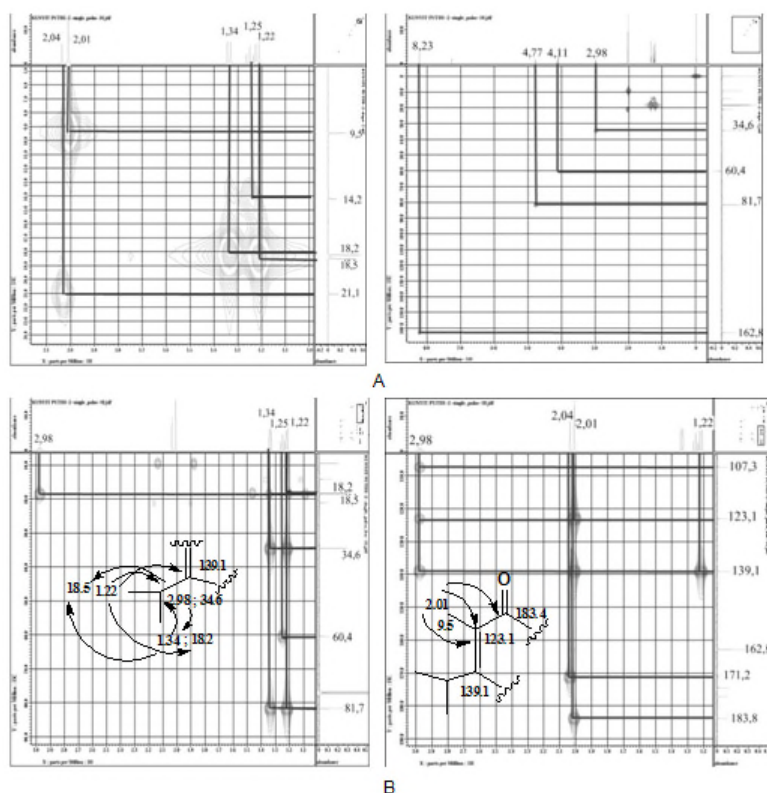
Carbon no.	$\delta_{\text{C}}$ ppm	DEPT	$\delta_{\text{H}}$ ppm ( $\Sigma\text{H}$ , multiplicity, $J$ in Hz)	HMBC	COSY
2	183.4	C			
3	123.1	C			
4	139.1	C			
5	107.4	C			
6	162.8	CH	8.23 (1H, s)	81.7, 107.4, 139.1	
7	34.6	CH	2.98 (1H, q, $J = 7.15$ Hz)	107.4, 123.1, 139.1	1.22
8	18.5	$\text{CH}_3$	1.22 (3H, d, $J = 7.15$ Hz)	139.1, 34.6, 81.7	
9	18.2	$\text{CH}_3$	1.34 (3H, d, $J = 7.15$ Hz)	34.6	
10	9.5	$\text{CH}_3$	2.01 (3H, s)	123.1, 139.1, 183.8	
2'	174.5	C			
3'	100.3	C			
4'	171.2	C			
5'	177.2	C			
6'	60.4	$\text{CH}_2$	4.11 (2H, q, $J = 7.15$ Hz)	171.2	1.25
7'	14.2	$\text{CH}_3$	1.25 (3H, t)	60.4	
8'	21.1	$\text{CH}_3$	2.04 (3H, s)	171.2	
9'	81.7	CH	4.77 (1H, q, $J = 7.15$ Hz)	139.1, 162.8	1.34

**Fig 2.** The  $^1\text{H-NMR}$  spectrum of compound **2**

alcohol group. Furthermore, the presence proton signal at  $\delta_{\text{H}}$  1.67 ppm (1H, *m*) for proton methine, signal at 1.2–1.4 ppm for four methylene group, and signal at 0.89 and 0.91 as pair of multiplet (3H, *m*) for two methyl groups.

The  $^{13}\text{C-NMR}$  spectrum of compound **1** (Table 1), confirming the symmetry of the molecule, exhibited the expected 12 carbon resonance. DEPT spectrum showed to two quaterner, three methane, five methylene carbons, and two methyl groups. These spectroscopic data, by comparison of  $^1\text{H}$  and  $^{13}\text{C-NMR}$  data to those

published in literature [19] and showed similarity, in conclusion compound **1** was identified as Di-(2-ethylhexyl)phthalate (DEHP). DEHP (compound **1**) is a well known synthetic plasticizers, so already reported to be present in *Calotropis gigantean* [15], *Alchornea cordifolia* [16], and *Aloe vera* [17]. The effective presence of compound **1** in endophytic fungus *Penicillium* sp of leave *C. zedoaria*, not as a contaminant from solvents and endophytic fungus *Penicillium* sp not cultivated in plastic bags, so these could be discounted as a source of DEHP.

Fig 3. Spectrum  $^{13}\text{C}$ -NMR compound 2Fig 4. HMQC correlation of proton at  $\delta_{\text{H}}$  1.23–8.23 ppm (A) and HMBC correlation of proton at  $\delta_{\text{H}}$  1.23–2.98 ppm (B) compound 2

Compound 2 was obtained as a yellow crystal, mp. 171–172 °C. The Spectra UV (MeOH) of 2 exhibited absorption at  $\lambda_{\text{max}}$  nm: 213, 253, and 321. The bathochromic shift in addition of NaOH exhibited absorption at  $\lambda_{\text{max}}$  nm: 213, 253, and 321. Base on Spectroscopic data UV indicate this compound was no

phenolic group. The IR spectra (KBr) showed  $\nu_{\text{max}}$   $\text{cm}^{-1}$ : 3466.08 (OH), 2980.02 and 2935.66 (C-H aliphatic), 1625.99 (conjugated C=O), 1579.70; 1521.12; 1438.90 (C=C conjugation), and 1180.44 (C-O ether).  $^1\text{H}$ -NMR (DMSO, 500 MHz)  $\delta_{\text{H}}$  ppm and  $^{13}\text{C}$ -NMR (DMSO, 125 MHz)  $\delta_{\text{C}}$  ppm (see Table 2).

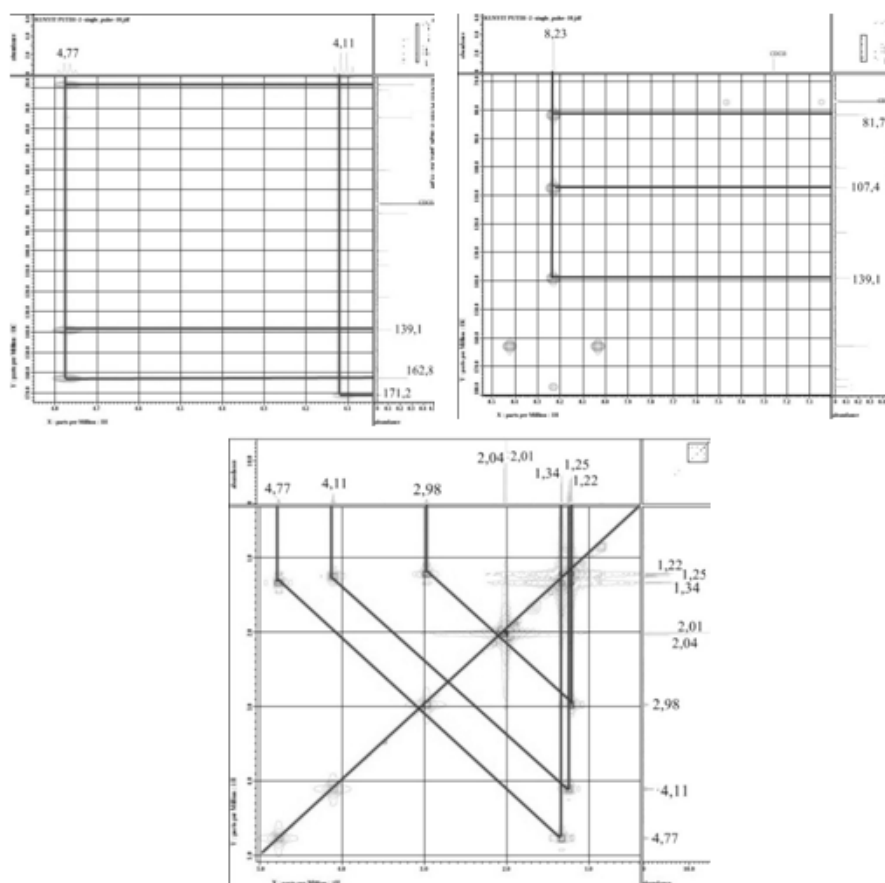


Fig 5. HMBC correlation of proton at 4.01–8.23 ppm and COSY correlation compound 2

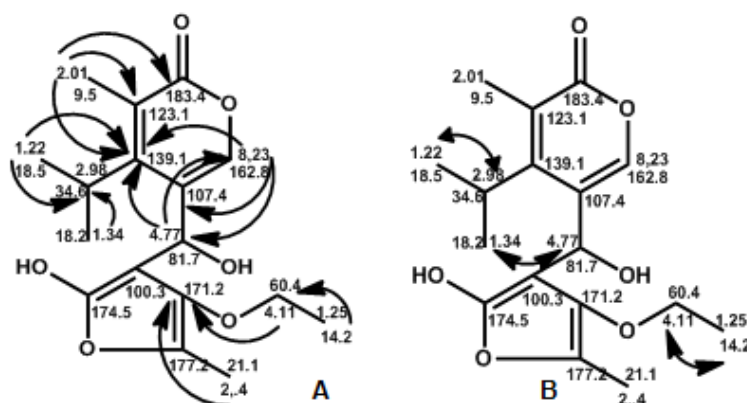


Fig 6. HMBC (A), and COSY (B) correlations and  $\delta$ -assignment of compound 2

The  $^1\text{H-NMR}$  spectrum, (Fig. 2) showed signal two methyl doublet at  $\delta_{\text{H}}$  1.22 and 1.34 ppm (3H, *d*, 7.15 Hz) and signal methine quartet at  $\delta_{\text{H}}$  2.98 ppm (1H, *q*, 7.15 Hz) and one methine singlet at  $\delta_{\text{H}}$  8.23 (1H, *s*). At spectrum also showed signal for methyl triplet at  $\delta_{\text{H}}$  1.25 ppm (3H, *t*, 7.15 Hz), and two methyl singlet at  $\delta_{\text{H}}$  2.01 and 2.04 ppm, (3H, *s*) and one signal methylene quartet at  $\delta_{\text{H}}$  4.11 ppm (2H, *q*, 7.15 Hz).

The  $^{13}\text{C-NMR}$  (Fig. 3), DEPT 135 spectrum, and HMQC spectrum (Fig. 4) showed 17 signal consist that nine signal as  $\text{C sp}^2$  and 8 signal as  $\text{C sp}^3$ . Analysis spectrum DEPT 135 showed 8 signal C quarternary at  $\delta_{\text{C}}$  100.3; 107.4; 123.1; 139.1; 171.2; 174.5; 177.2 and 183.8 ppm, 5 signal methyls carbon at  $\delta_{\text{C}}$  9.5; 14.2; 18.2; 18.5 and 21.1 ppm, 3 signal methines carbon at  $\delta_{\text{C}}$  34.6; 81.7 and 162.8 ppm, and one signal

methylene carbon at  $\delta_C$  60.4 ppm. Signal carbon at  $\delta_C$  183.4 ppm indicated these compound have C=O carbonyl.

NMR 2D analysis for HMQC spectrum (Fig. 4) showed the proton at  $\delta_H$  1.34 ppm correlation to carbon at  $\delta_C$  18.2 ppm and proton at  $\delta_H$  1.22 ppm correlation to carbon at  $\delta_C$  18.5. HMBC spectrum showed correlation from proton at  $\delta_H$  1.22 and 1.34 ppm to carbon at  $\delta_C$  34.6 and 139.1 ppm. Proton at  $\delta_H$  1.22 also correlation to carbon at  $\delta_C$  18.2 ppm and proton at  $\delta_H$  1.34 ppm showed correlation with carbon at  $\delta_C$  18.5. This data to indicated that proton  $\delta_H$  1.22 and 1.34 ppm bound to carbon fasten carbon  $\delta_C$  34.6 ppm. Proton at 1.25 ppm correlation to carbon at  $\delta_C$  60.4 ppm. Further HMBC spectrum showed correlation proton at  $\delta_H$  2.01 (3H, s) to carbon at  $\delta_C$  123.1; 139.1 and 183.8 ppm, and correlation proton at  $\delta_H$  2.04 ppm to carbon at  $\delta_C$  171.2 ppm. Proton at  $\delta_H$  2.01 and 2.04 (3H, s) at HMQC spectrum showed fastened with carbon at  $\delta_C$  9.5 and 21.1 ppm.

Proton at  $\delta_H$  4.11 correlation to carbon at  $\delta_C$  171.2, proton at  $\delta_H$  4.77 ppm showed correlation to carbon at  $\delta_C$  139.1; 162.8 ppm, while proton at  $\delta_H$  8.23 ppm to correlation to carbon at  $\delta_C$  81.7; 107.4 and 139.1 ppm. Analysis of  $^1\text{H}$ - $^1\text{H}$  COSY spectrum (Fig. 5) also to indication of two proton spin system corresponding at  $\delta_H$  1.22 with proton at  $\delta_H$  2.98 ppm. And proton at  $\delta_H$  1.25 to correlation to proton at 4.11 ppm. HMBC and COSY correlation and  $\delta$ -assignment of compound showed Fig. 6. These spectroscopic data, therefore suggested that compound **2** is 5-(4'-ethoxy-2'-hydroxy-5'-methyl-2',3'-dihydrofuran-3'-il (hydroxy) methyl-4-isopropyl-3-methyl-2-pyran-2-on).

Compound **1** is not new compound, but it is new for endophytic fungus from *C. zedoaria* and base on Dictionary Natural Products data base, 5-(4'-ethoxy-2'-hydroxy-5'-methyl-2',3'-dihydrofuran-3'-il (hydroxy) methyl-4-isopropyl-3-methyl-2-pyran-2-on) (**2**) is new compound. Exploration of secondary metabolites research needs to be done in order to get the profile of organic compounds produced by endophytic fungus of *C. zedoaria*.

## CONCLUSION

Two compounds have been isolated from the endophytic fungus *Penicillium* sp from the leaves of kunyit putih (*C. zedoaria*). Based on spectroscopic analysis and comparison data to those published in literature compound **1** was identified as Di-(2-ethylhexyl)phthalate and compound **2** as

5-(4'-ethoxy-2'-hydroxy-5'-methyl-2',3'-dihydrofuran-3'-il (hydroxy) methyl-4-isopropyl-3-methyl-2-pyran-2-on).

## ACKNOWLEDGEMENT

The authors are statement grateful to the Directorate General of Higher Education which research grant Fundamental 2013 was supported this research.

## REFERENCES

1. Strobel, G., Daisy, B., and Castillo, U., 2005, *Plant Pathol. J.*, 4 (2), 161–176.
2. Premjanu, N., and Jayanthi, C., 2012, *Int. J. Inst. Pharm. Life Sci.*, 2 (1), 135–162.
3. Lakshmi, S., Padmaja, G., and Remani, P., 2011, *Int. J. Med. Chem.*, 2011, 1–13.
4. Saikia, N., and Nath, S.C., 2003, *J. Econ. Taxon. Bot.*, 27, 430–433.
5. Jang, M.K., Sohn, D.H., and Ryu, J-H., 2001, *Planta Med.*, 67 (6), 550–552.
6. Wilson, B., Abraham, G., Manju, V.S., Mathew, M., Vimala, B., Sundaresan, S., and Nambisan, B., 2005, *J. Ethnopharmacol.*, 99 (1), 147–151.
7. Bugno, A., Nicoletti, M.A., Almodóvar, A.A.B., Pereira, T.C., and Auricchio, M.T., 2007, *Braz. J. Microbiol.*, 28, 440–445.
8. Elfita, Muharni, Munawar, Legasari, L., and Darwati, 2011, *Indo. J. Chem.*, 11 (1), 53–58.
9. Elfita, Muharni, Munawar, and Aryani, S., 2012, *Indo. J. Chem.*, 12 (2), 195–200.
10. Xu, L., Zhou, J., Zhao, J., Li, X., and Wang, J., 2008, *Lett. Appl. Microbiol.*, 46 (1), 68–72.
11. dos Santos, R.M.G., and Rodrigues-Fo, E., 2003, *Z. Naturforsch.*, 58c, 663–669.
12. Yan, H-J., Gao, S-S., Li, C-S., Li, X-M., and Wang, B-G., 2010, *Molecules*, 15 (5), 3270–3275.
13. Barik, B.P., Tayung, K., Jagadev, P.N., and Dutta, S.K., 2010, *Eur. J. Biol. Sci.*, 2 (1), 8–16.
14. Guo, L., Wu, J-Z., Han, T., Cao, T., Rahman, K., and Qin, L-P., 2008, *Molecules*, 13 (9), 2114–2125.
15. Habib, M.R., and Karim, M.R., 2009, *Micobiology*, 37 (1), 31–36.
16. Mavar-Manga, H., Haddad, M., Pieters, L., Baccelli, C., Penge, A., and Quetin-Leclercq, J., 2008, *J. Ethnopharmacol.*, 115 (1), 25–29.
17. Lee, K.H., Kim, J.H., Lim, D.S., and Kim, C.H., 2000, *J. Pharm. Pharmacol.*, 52 (5), 593–598.