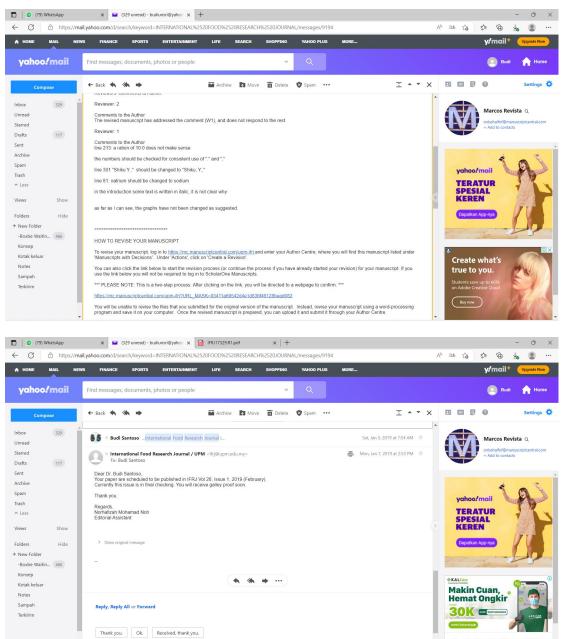


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Gratis Ongkir Hingga 30K

1 The effect of eel's protein extract on the characteristics of edible film

from crosslinked modified canna starch

3

2

4 Abstract

5 This study aimedtostudy theeffect of eel's proteinextractoncharacteristics ofmodified Canna's starch ediblefilm.Research method was addition of 6 7 eel's protein extract at concentrations of 2%(v/v), 4%(v/v), and 6%(v/v) in 8 the formulation of modified Canna's starch ediblefilm. The observed 9 parameters were percentelongationpercentage, compressive strength, water vapor transmission rateand themicrostructure. The addition of eel's 10 protein extracts increases the elongation percentage and decreases water 11 vapor transmission rate of edible flim. The edible compressive strength of 12 the film decreases after the addition of eel's protein extract, but addition of 13 14 higher concentration of eel's protein extracts had increased compressive strength of edible film. 15

Comment [W1]: There is no discussion of microstructure in the manuscript.

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Keywords: edible film, Canna's starch, eel's extract protein, modified, POCl_{3.}

19 Introduction

The use of Canna's starch as raw material for edible flim is still limited due to low values of elongation percentage, compressive strength and water vapor transmission rate. One of effort that can be done is by using modified Canna's starch through crosslinking of POCl₃because the amylose and amylopectin compositions are relatively balance with
magnitude of 25% and 75%, respectively.

26 The use of modified starch is very important to produce edible film matrix because POCI₃ compound creates a cross link between one 27 amylose chain to anothers through phosphate link. This phosphate link can 28 29 produce three dimension continous net and this net can trap water through 30 reactive OH which not bound with POCI₃. The cross link of starch polymer chains is occurred at groups containing more OH reactive, especially at OH 31 32 group numbers 2, 3 and 6 (Yoshida et al., 2002). The numbers of reactive OH group is affected by cross-linking degree and starch concentration. 33 Granulair size and ratio of amylose and amylopectin numbers of starch may 34 35 also affect the numbers of reactive OH group. The characterisics of these starches may have effect on the producing of modified starch. According 36 37 toSantosoet al.(2011)the modified starch through cross linking by POCl₃ may produce good compressivestrengthbut low percent elongation and 38 water vapor transmission rate edible film. This low percent elongation and 39 40 water vapor transmission rate characteristics can be improve by the addition of other compound such as glycerol and beeswax. 41

Glycerol and bees wax not only has positive effect, but also negative effect such as increasing water vapor transmission rate anddecreasing elongation degree of *edible film respectively*. The addition of eel's extract protein is important to solve this problem. According toArtharn*et al.* (2008),fish protein based *edible film* which consisted or myofibrillar protein or sarcoplasm generally have better mechanical

properties, especially in term of flexibility property. Nakai and Modler 48 (1999) had added that protein types of myofibrillar and sarcoplasmic are 49 50 found in abundant quantity on eels. Amino acid types and pH isoelectric point also have profound effect on the edible film matrix (Prodpranet al., 51 2007).Were et al. (1999)described that amino acids containing sulphur 52 have important role in edible film formation through disulfide bonding. 53 54 PoeloengasihandMarseno (2003) had added that in addition to disulfide bonding, hydrogen bonding and hydrophobic interaction also determine 55 56 edible film properties, especially in term of amino acids having hydophobic 57 charateristic such as alanine, valine, leusin, triptophane and phenylalanine.

58 This research aimedtostudy theeffect of eel's 59 proteinextractoncharacteristics of modified Canna's starch ediblefilm and 60 eel' protein extract which in turn can improve elongation percentage, 61 compressive strength, and water vapour transmission rate of edible film.

62

63 Material and method

64 Materials and equipments

Material used in this study were canna's starch (white color) from Pagaralam City, eel from Perumnas Market of Palembang. Eel is processed into surimi to obtain protein extract. Chemicals used for preliminary study were alcohol, distilate water, ether, HCl, NaOH, Fehling solution, ethanol, methylene blue, iod solution, acetate acid, glycerol and trisodium citrate having technical quality. Chemicals for production of modified starch and edible film were natrium sulphate, POCl₃, glycerol,

Comment [W2]: acetic

Comment [W3]: technical grade

CMC, *beeswax*, pp indicator, HCI and ammonium molybdate.Equipments
used in this study were *hot plate*, *magnetic stirrer*, *vortex*, oven, desicator,
texture analyzer, testing Machine MPY (Type: PA-104-30, Ltd Tokyo,
Japan), and water vapor transmission rate tester Bergerlahr.

76

77 Method

78 Modified Starch Processing from Canna's Starch with POCI₃by Using Cross

79 Linking Method (Modified method of Wattanachantet al., 2003)

80 Procedure for modified starch processing with cross-linking method by using multi functional reagent of POCI3 is as follows:natrium sulphate 81 (Na₂SO₄) with magnitude of 30g (15% dry weight of starch) is prepared and 82 83 added with 300mL distilate water and it is stirred by using magnetic stirrerat 3 scale; 200g starch is added while stirred; 5% NaOHis added while stirred 84 85 by using magnetic stirrer at 8 scale to prevent starch gelatinization and pH level of solution is set at 10.5 and stirred for 30 minutes at room 86 temperature; The solution is incubated using shaker incubator at 87 temperature of 40+2°C (200rpm, 24 hours); POCI₃is added 0.08%(w/w) 88 89 while stirred by using magnetic stirrer at 8 scale for 30 minutes and then 90 incubated at temperature of 40+2°C (200rpm, 2 hours); pH of solution is set 91 to 5.5 using 10% HCl solution to stop the reaction; Starch is sieved by using Whatman paper no. 4 and washed with distilate water for 5 minutes. 92 Starch drying is done at temperature of 45°Cfor 6 hours to get starch with 93 water content of 10-12%. 94

95

96 Protein extract preparation

97 Procedure for protein extract from eel and its preparation for 98 addition into edible film is as follows (modified method ofHeruwati and Jav, 1995): Eel is cleaned by discharging its head part and stomach content 99 followed by cleaning with clean water; Cutting operation is done to separate 100 flesh portion from bone and skin (fillet)followed by smashing of eel 101 102 flesh;Smash flesh is cleaned with cold water at temperature of about 1 to 103 5°Cusing water having volume 5 times of smash flesh volume for 10 104 minutes; Smash flesh is stirred within cold water until homogenous and is stopped to precipitate smah flesh, whereas impurities and lipid are floating 105 106 in water surface, followed by discharging the impurities; Water is separated 107 from cleaned smash flesh by using pressing equipment; Smash flesh is recleaned within cold water and added with 0.3% salt (w/v) at the third 108 109 cleaning and followed by recompressing to discharge water as much as possible; 2% of sorbitol (w/v)is added and stirred until homogenous; Smash 110 flesh is freezed within *freezer*at temperature of about -15°C for 1 week; 111 Frozen smash flesh is thawed for 30 minutes followed by weighing with 112 magnitude of 2% (w/v), 4% (w/v) and 6% (w/v) from total aguadest volume; 113 100ml aquadest and NaOH 1 M are added to achieve pH 11 and stirred, 114 115 heated at temperature of 55°C for 30 minutes; Heating and screening of suspension are done to produce protein extract; Reheated is done at 116 temperature 60°C; and solution (suspension) is ready to be used as edible 117 film material. 118

120 Edible film preparation.

Procedure for processing of canna starch edible film without and 121 122 with the addition of protein extract of eel is as follows: Preparation of canna starch native (P₁) and (P₂) modified using POCl₃ 0,08% with magnitude of 123 4% (w/v); The addition of aquadest followed by stirring and sieving; Starch 124 suspension heating at gelatinization temperature of 65°Cusing hot 125 126 platefollowed by stirring with magnetic stirrer; The slowly addition of 127 glycerol 3% (v/v) to starch suspension that had fully gelatinized and then 128 followed by heating for about 10 minutes; Addition of eel extract protein according to treatments with concentration of S_1 (2%), $S_2(4\%)$, and $S_3(6\%)$ 129 v/v; Addition of CMC surfactan according to treatments with concentration 130 of 2% (b/v); Suspension is stirred until homogenous and addition of bees 131 wax with concentration of 1% (b/v); Removing dissolve gas (degassing)by 132 133 using vacuum pump for 1 hour; Pouring the 40mL of suspension to petri dish 15cm in diameter followed by moulding and heating at 70°C for 12 134 hours using oven; and cooling at room temperature followed by edible film 135 removal from moulder and wrapped with plastics, put it into desicator for 136 137 24 hours.

138

139 Statistical analysis

Statistical analysis on completely randomized experimental was done by using SAS computer Program of Window 6.12 version. One-way analyses of variance (ANOVA) were carried out and mean comparisons were processed by Duncan test. Significance was defined as p<0.05.

144

145 **Results and discussion**

Addition of protein extract of eel has an objective to improve elongation degree of canna starch edible film. Protein molecules of eel within edible film matrix is bound with hydrophilic components such as starch, glycerol and CMC. Astiana (2012) had explained that eels contain 15.90% essential and non-essential hydrophobic proteinswhich consisted of lysine and glutamic acid with magnitudes of 7.13g / 100g and 12.89g/100g, respectively.

Analysis of variance results showed that extract protein of eel 153 treatments had significant effect, whereas their interaction had no 154 significant effect on elongation percentage value of edible film (α =0.05). 155 Results of Duncan test (Table 1) showed that elongation percentage value 156 157 of modified canna starch edible film was significantly different than that of native canna starch edible film. Treatment 2% protein extract of eel was 158 significantly different than that of 4% and 6%, but treatment 4% protein 159 extract of eel was not significantly different than that of 6% in term of edible 160 film elongation percentage. 161

Elongation degree value of modified canna starch edible film (57.33%) was higher than that of native canna starch edible film (48.00%). This was due to the fact that modified canna startch had more open starch molecules structure and low retrogradation characteristics. This starch structure cause protein molecules of eel is easily enter to edible film matrix and boundwith hydrophilic components such as starch, glycerol and CMC.

Low retrogradation characteristics of modified canna starch cause proteinmolecules of eel trapped and stable within edible film matrix.

170 According toNakai and Modler (1999), protein extract of eel contain the same protein such as found in fish, i.e. myofibrillar and sarcoplasmic 171 with magnitude of 65 to 75% and 20 to 30%, respectively. Weng et al. 172 (2007)had stated that myofibrillar protein has fibrous form and elastics 173 174 whereas sarcoplasmic protein is globular. Myofibrillar protein affect edible 175 film elasticity increment and sarcoplasmic protein decrease polymers 176 interaction and cohesive power of edible film matrix as well as produce more flexible edible film. Fish protein based edible film consisting of 177 myofibrillaror sarcoplasmicprotein generally have better mechanical 178 179 properties, especially flexibility property, but has low barrier to water vapour transmission rate (Hamaguchiet al., 2007) 180

Table 1 showed that higher protein extract of eel results in higher elongation degree of *edible film*. Higher protein extract of eel results in higher quantity of myofibrillar and sarcoplasmic protein molecules which are trapped within edible film matrix.

Average value of elongation percentage of canna starch edible film with protein extract of eel was higher than that of edible film without protein extract(Figure 1). However, this edible fim elongation percentage had not fulfilled the stated standard of JIS 1975, i.e minimum of 70% for all addition levels of protein extract.

Analysis of variance results showed that protein extract of eel had significant effect on edible film compressive strength value, whereas their **Comment [W4]:** Please make it clear. The figure 1 can not confirm this statement. The treatmen C2 (in figure 1) needs to be clearly defined in edible film preparation whether it is a control (treatment without addition of protein extract).

interaction was not significant (α =0.05). Compressive strength value of modified canna starch edible film was significantly different than to native starch edible film. The use of protein extract of eel with concentrations of 2%, 4% and 6%gave significantly different values of edible film compressive strength (Table 1).

197 Compressive strength value of modified canna starch edible film 198 was higher than native starch film. This was caused by stronger molecular 199 structure of canna starch modified than native. POCl₃ cross linked canna 200 starch had some substituted OH groups by phosphate. This subtitution 201 increase the structural strength of starch molecules that resulted the 202 stronger starch molecules. Degree of edible film matrix strength is 203 increased by increasing starch strength.

Table 1 showed that the higher the protein extract concentration of 204 205 eel, the higher the compressive strength of edible film. This was due to the fact that protein extract of eel contains myofibrillar protein. This protein 206 moleculair chains have fibrous form and length. Higher myofibrillar protein 207 content results in more compact edible film and higher resistance power to 208 209 pressure. This results was in accordance to study that reported bySobralet 210 al.(2005) which showed that the use of 2g of myofibrillar protein in 100g film 211 solution had higher pressure strength than that of 1g myofibrillar protein in 100g film solution.Artharn et al. (2008) had stated that edible film 212 formulation with ratio myofibrillar protein and sarcoplasmic protein of 10:0 213 had produced the highest value of compressive strength. 214

Compressive strength value of canna starch edible film with protein extract of eel was lower than that without protein extract addition (Figure 2). 216 217 Sarcoplasmic protein is globulair form protein. This protein is dispersed amongst edible film matrix which lower interaction with film matrix polymers 218 and lower compactness which subsequently decrease the edible film 219 resistance power to pressure.Artharn et al. (2008)had reported that 220 221 compressive strength of protein-based edible filmwill decrease when 222 concentration of sarcoplasmic protein is increase.

215

223 Analysis of variance results showed that treatments of protein extract of eel had significant effect on water vapour transmission rate of 224 225 edible film (α =0,05). Results of Duncan test (Table 1) showed that 226 treatments of 2%, 4% and 6% protein extract of eel had significant effect on 227 water vapour transmission rate of edible film.

228 Table 1 showed that higher protein concentration had produced lower water vapour transmission rate. This was due to protein extract of eel 229 which had myofibrillar and sarcoplasmic proteins. Molecular structure of 230 myofibrillarprotein is consisted of fibrous molecular chains. The higher the 231 232 concentration of protein extract of eel, the higher the quantity of myofibrillar 233 protein which in turn produce more solid and compact of film matrix 234 structure. This condition impede water vapour to penetrate edible film matrix. Shiku et al. (2003) had added that water vapour transmission rate of 235 edible film produced from myofibrillar protein was relatively lower than that 236 of edible films produced from other proteins.Sarcoplasmic proteins are 237 globular proteins containing most of the hydrophobic and SH groups hidden 238

Comment [W5]: Please make it clear. The figure 2 can not confirm this statement. The treatmen C2 (in figure 2) needs to be clearly difined in edible film preparation whether it is a control (treatment without addition of protein extract).

in the interior of molecules. Formation of sarcoplasmic protein films
prepared from blue marlin (*Makairamazara*) has mainly involved thermal
treatment of film-forming solutions at temperature ranging between 55°C
and 90°C (lwata *et al.*, 2000).

Figure 3 showed that water vapour transmission rate of cannal 243 starch edible film with protein extract of eel was higher than that without 244 245 protein extract of eel. This was due to protein molecules as structural 246 components of edible film matrix which has hydrophilic characteristics. 247 Protein addition results in increase of hydrophilic components within edible film matrix. Therefore, the higher the hydrophilc components, the easier the 248 249 water vapour to penetrate edible film. Yoshida et al. (2002)described that 250 natural hydrophilic property of protein in edible film formulation facilitate the interaction with water which lower edible film resistance power to water 251 252 vapour. Nayak et al.(2008) described that protein-based edible film was very effective as barrier to oxygen gas and aroma, but this edible film 253 254 showed relativey high value of water vapour transmission rate.

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256 Conclusion

Addition of eel's protein extract had increased elongation percentage and decreased water vapour transmission rate of edible film. Compressive strengthof edible film had decreased with eel's protein extract addition, but addition of higher concentration of eel protein extracts had increased its compressive strength.

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Comment [W6]: See w5

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Table 1. Results of Duncan test for the effect of eel's protein extract on

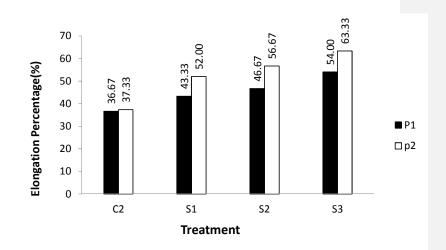
elongation percentage, compressive strength and water vapour transmission

336 rate of canna's starch edible film.

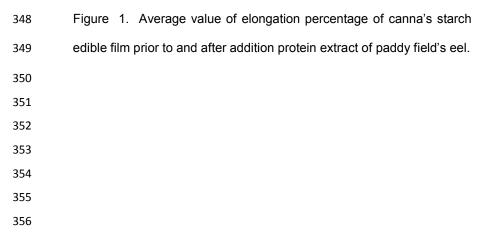
Treatment	Elongation percentage (%)	Compressive Strength (Newton)	Water vapour transmission rate (g.m ⁻² .d ⁻¹)
S ₁	47.60 ^a	51.69 ^a	22.60 ^a
S_2	51.67 ^b	58.17 ^b	21.68 ^b
S ₃	58.84 ^b	63.87 ^c	18.85 [°]

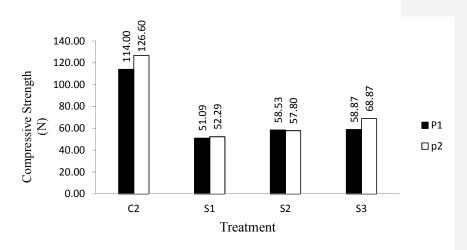
338	Remarks:	Numbers	followed	by	the	same	letter	in	the	same	column	are	not
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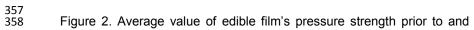
339 significantly different at 5% level of Duncan test



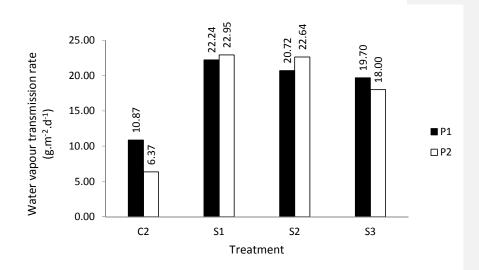








359 after protein extract addition of paddy field's eel



368

369 Figure 3. Average value of water vapour transmission rate of canna's starch

edible film prior to and after protein extractaddition of paddy field's eel