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[Antibacterial Potential Screening of Halimeda sp on Some Types of Pathogenic Bacteria](#)

Hendri M., Darmanto J. S., Prayitno B., Radjasa O.K.

Research Report

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Antibacterial Potential Screening of *Halimeda* sp on Some Types of Pathogenic BacteriaHendri M.^{1,2}, Darmanto J. S.³, Prayitno B.³, Radjasa O.K.⁴

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Abstract The study was conducted as a test to determine the effectiveness of *Halimeda* sp seaweed extract on the growth of some types of pathogenic bacteria. Seaweeds extracted consist of four (4) types which include: *Halimeda macrophysa*, *Halimeda gracillis*, *Halimeda Opuntia* and *Halimeda renschi*. While the types of pathogenic bacteria used were (*Salmonella typhi*, *Staphylococcus aureus*, *Escherichia coli* and *Bacillus subtilis*). This study uses methanol in the ratio of 1: 1 (v/v) and were observed for 48 hours. The test results showed that the extract of *Halimeda* sp is effective as antibacterial pathogen. Phytochemical test showed the presence of steroid and saponin compounds.

Keywords Antibacterial; Bacterial Pathogens; *Halimeda* sp; Seaweed

1 Introduction

Seaweed is one of important marine commodities that have high economic value for export. Currently seaweed has been developed by means of cultivation. This activity is carried out by various parties such as companies, governments, and fishermen community. The benefits of this plant are commonly known as product of food, beauty, medicines and others (Aslan, 1998; Anggadireja et al., 2006).

Some marine organisms, especially from the class of marine algae, have the ability to produce chemical compounds that are not found or rarely found in organisms that live on land (Nybakken, 1993). Some types of marine biota synthesize and store toxic compounds called marinetoxin on parts of his body and released into the environment (Djapiala et al., 2013; Anggadireja et al., 2006). These compounds are secondary metabolites which are used as a defense and to preserve life, to avoid interference from predators. These compounds have pharmacological activity, so it is possible to be developed (Paul and Fenical, 1983; Paul and Puglisi, 2004; Paul and Fenical, 1984; Paul and Van Alstyne, 1988).

Halimeda is a marine plant that has green leaves and is one type of green algae group. *Halimeda* has the ability

to produce biotif substances for antifouling. The active substance produced for biofouling is known as *halimeditrial* and *halimeditetraasetat*. *Halimeditrial* is *diterpenoid* that yet trialdehyde, known as the major secondary metabolite in six species of algae containing calcium *Halimeda* (Paul and Fenical, 1983; Paul and Fenical, 1984; Paul and Puglisi, 2004; Kumar et al., 2010; Bachtiar et al., 2012; Paul, 1987).

Seaweed, primarily from the group *Halimeda* sp has the ability to issue a secondary metabolite in the process of metabolism to defend themselves against predators and pests. The active ingredients released by *Halimeda* are very effective to prevent attacks of predators and bacteria (antifouling). *Halimeditrial* and *halimeditetraasetate* a bioactive compounds contained in seaweed (*Halimeda* sp) (Paul and Fenical, 1983; Paul and Fenical, 1984; Paul and Fenical, 1986; Paul, 1987; Atmadja, 1992; Paul and Van Alstyne, 1992).

The ability of algae to produce halogenated secondary metabolites that act as bioactive compounds might happen, because the environmental conditions such as high salinity or will be used to defend themselves from the threat of predators. In the last decade, a variety of structures of bioactive compounds that very unique from red algae have been isolated. However,

utilization of bioactive ingredients from algae has not been done. Based on the biosynthesis process, marine algae are rich in compounds derived from the oxidation of fatty acids called oxylipin. Through these compounds various types of secondary metabolites are produced (Paul and Fenical, 1983; Paul and Fenical, 1986; Hay and Fenical, 1988; Paul and Puglisi, 2004; Hay, 1996; Karthikaidevi et al., 2009; Kolsi et al., 2015).

Halimeda chemically able to produce diterpenoid metabolites halimedatriol and Halimeda tetra acetate at various concentrations. This metabolite has been observed to play a role in chemical defense against herbivores, based on their chemical structure and biological activity. Halimedatriol more effective than halimedatetraacetate in marine algae defense system to repel natural enemies (Paul and Fenical, 1983; Paul and Fenical, 1984; Paul and Fenical, 1986; Paul and Van Alstyne, 1988; Paul and Puglisi, 2004)

2 Material and Methods

Materials

Seaweed *Halimeda* sp, collected from the waters of the Gulf of Lampung. Sampling was carried out in June - July 2014 and analyzed at the Marine Biological Laboratory Faculty of University of Sriwijaya, Basic Chemistry and Biotechnology Laboratory LIPI Cibinong.

Samples seaweed washed with running water and rinsed with sterile water and cut into small pieces. Subsequently dried and crushed made flour. Halimeda sp extracted with methanol, evaporated with a rotary evaporator. Extracts of secondary metabolites identified by thin-layer chromatography (TLC) or thin-layer chromatography (TLC). Dry extract sample dissolved in methanol is used as the test solution, then spotted by 5 mL of test solution and standard solution on the plates of silica gel GF 254 as the stationary phase. Put the plates into the chromatography vessel that has been saturated with mobile phase consisting of a mixture of Chloroform-methanol (10: 1) v/v. Elution until the upper limit of the stationary phase plate. Identification chromatography with UV light of 254 nm, and then sprayed with cerium sulfate reagent. Then dried and viewed with UV 254 nm.

Other materials used are pathogenic bacterial culture types *S.typhi*, *S. aureus*, *E. coli* and *B.subtilis* obtained from laboratory Basic Chemistry and Biotechnology

LIPI Cibinong Bogor. Media for the pace of the bacteria used are nutrient agar (NA) and liquid nutrient broth (NB).

The tools used include blenders, autoclaves, incubators, distillator, pH meter, ose needle, micropipette, magnetic stirrer, micrometers, shaker, hot plates and oven.

Antimicrobial Materials Selection

At this stage, the analysis of water content materials is conducted (Apriyantono et al., 1989) and the selection of materials using solvent extraction of water and testing activities by the agar diffusion method.

Extraction of materials

The extraction step includes the destruction of material, the addition of water at a ratio of materials and water 1:1, 1:2, 1:3 (w/v), then filtering treatment. The filtrate obtained is sterilized.

b. Testing antimicrobial activity by agar diffusion method (Wolf and Gibbons, 1996). Nutrient Agar (NA) which has been sterilized cooled to a temperature of 50o C in a water bath. Each bacterial culture was aged 24 hours at a concentration of 107-108 cells per mlk inserted into the NA of 40 uL for every 20 ml of NA. Subsequently made to the cup with a thickness of 4-5 mm.

Then put the paper disc that has been dipped in each extract Halimeda. Subsequently incubated at 37o C for 48 hours. Then observed the presence of inhibitory and in measuring the diameter of inhibition (in mm) using a micrometer measuring tool. This stage is carried out with two replications.

3 Result and Discussion

Result

Antibacterial Test Results

Halimeda sp crude extract was tested by using four (4) types of pathogen bacteria (*S.typhi*, *S. aureus*, *E. coli*, *B. subtilis*) with treatment four (4) types of *Halimeda* sp extracts which include: *H.macrophysa*, *H. incrassata*, *H.opuntia* and *H.renschi*. This test is done observation for 48 hours. In general, the effect of this extract is significant to the growth of these bacteria (see Table 1).

Phytochemicals Test Results

The phytochemical test results showed extracts *H.renschi* and *H. gracillis* containing steroids and saponins compounds, while alkaloids, terpenoids, tannins and flavonoids are not contained in the extract (Table 2).

Table 1 Methanol Extract Antibacterial Test Result from *Halimeda* sp (48 hours)

No.	Material	<i>S.Typhi</i> (-)	<i>S.aureus</i> (+)	<i>E.coli</i> (-)	<i>B.subtilis</i> (+)
1	<i>Halimeda opuntia</i>	12 mm	4 mm	15 mm	7 mm
2	<i>Halimeda gracillis</i>	10 mm	12 mm	-	11 mm
3	<i>Halimeda renschi</i>	11 mm	4 mm	-	13 mm
4	<i>Halimeda macrophysa</i>	3 mm	8 mm	13	4 mm

Table 2 Result Test of Extract Phytochemicals of *H.renschi* and *H. gracillis*

No	Phytochemicals Test	Phytochemicals Test Analysis Result		Method
		<i>H. gracillis</i>	<i>H. renschi</i>	
1.	Alkaloid	Negative	Negative	Qualitative Analysis
2.	Steroid	Positive	Positive	
3.	Terpenoid	Negative	Negative	
4.	Tanin	Negative	Negative	
5.	Saponin	Positive	Positive	
6.	Falavonoid	Negative	Negative	

KLT Test Result

Furthermore, each extract was analyzed with TLC plate, TLC Results showed suspected stain patterns potentially contain secondary metabolites with the invisibility of the dominant pattern fluorescent stain under UV light but the compound is not pure and there are still impurities. Isolation and purification are still needed to determine the type of the active compound (Figure 1).

The test results on the four (4) types of *Halimeda* sp are extracted on the growth of *E. coli* bacteria showed that extracts of *H.opuntia* and *H. macrophysa* which has a significant influence with a diameter of between 13-15 mm. This shows that the extract has a *Halimeda* extract inhibition against the bacteria *E. coli*. While *H.gracillis* and *H.renschi* no effect. The pattern of growth can be seen in Figure 2.

To test the growth of *S. aureus* on the four *Halimeda* sp seaweed extract shows have influence with a diameter of 4-12 mm. The largest to the smallest diameter is the extract of *H. gracillis*, *H. macrophysa*, *H. renschi*, and *H. Opuntia*. This means that all sample extracts have the ability/inhibition of the growth of bacteria. The highest inhibition owned by extracts of seaweed species *H. gracillis* with 12 mm. While the lowest inhibitory owned by *H. renschi* and *H. opuntia* with inhibition of 4 mm. The pattern of growth can be seen in Figure 3.

Halimeda sp extract test results against bacterial growth related *B. subtilis* shows that extracts of *Halimeda*

have influence with diameter about 4-13 mm. The highest inhibition by 13 mm at *H.renschi* extracts, whereas inhibition of the lowest in the extract of *H. macrophysa* with a diameter of 4 mm (see Figure 4).

The fourth extract *Halimeda* also tested for bacterial growth *S.typhi*. The test results showed that the extract had an influence with a diameter of 3-12 mm. The most high-power inhibitor is owned by *H. opuntia*

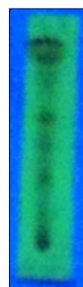


Figure 1 KLT test result

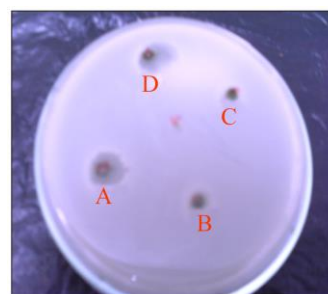


Figure 2 Results of Antibacterial Test Methanol Extracts of the *E. coli* bacteria on a 48 hours observation (A. *H.opuntia*, B *H.gracillis*, C. *H renschi* and D. *H macrophysa*)

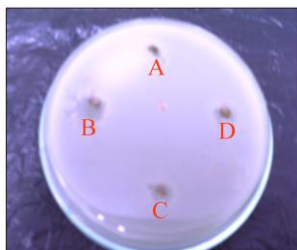


Figure 3 Antibacterial Test Results methanol extract against *S. aureus* bacteria at 48 hours observation (A. *H. opuntia*, B. *H. gracillis*, C. *H. renschi* and D. *H. macrophysa*)

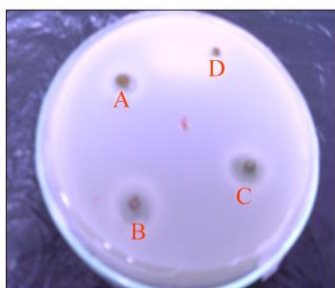


Figure 4 Results of Antibacterial Test Methanol Extracts of the bacteria *B. subtilis* at 48 hours observation (A. *H. opuntia*, B. *H. gracillis*, C. *H. renschi* and D. *H. macrophysa*)

with diameter of 12 mm, then *H. renschi* with 11 mm and *H. gracillis* with a diameter of 10 mm. While the lowest seaweed extract is *H. macrophysa* with a diameter of only 3 mm. The pattern of growth can be seen in Figure 5.

These results indicate that the aforementioned extraction have a fairly good inhibitory to the growth of pathogenic bacteria such as *S. typhi*, *S. aureus*, *B. subtilis*, *E. coli* bacteria. Anti-bacterial activity demonstrated in this study is active. The types of extracts based on test results of the highest *H. opuntia* active are in *E. coli* bacteria, the highest *H. gracillis* extract in *S. aureus*, the highest *H. macrophysa* extract in *E. coli* bacteria. While the highest extract of this type of *H. renschi* is in *B. subtilis* bacteria (Figure 6).

In addition each extract has anti-bacterial capabilities that varies depending on the type of extracts and bacterial strains. This means that the zone of inhibition showed antimicrobial activity against pathogens bacteria is varied. The ability of *Halimeda* sp extract to inhibits the growth of bacteria is also influenced by the test bacterial cell wall. (Fardiaz, 1983) states that the positive and gram-negative bacteria have different cell wall sensitivity against

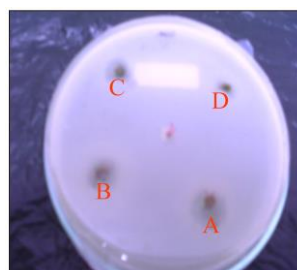


Figure 5 Results of Antibacterial Test Methanol Extracts against bacteria *S. Typhi* in observation 48 hours (A. *H. opuntia*, B. *H. gracillis*, C. *H. renschi* and D. *H. macrophysa*)

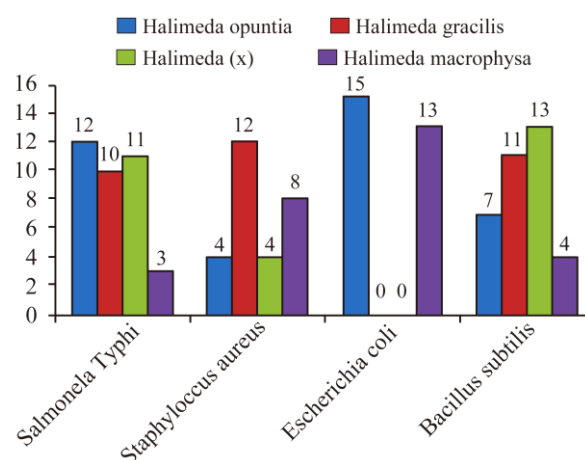


Figure 6 *Halimeda* sp extract inhibiting activity against pathogenic bacteria growth activity

physical treatment, enzymes and antibodies (Atmadja, 1992; Izzati, 2007; Shanab, 2007).

Discussion

The bacteria used in this study is a gram-negative and gram-positive, was able to be inhibited by the extract of *Halimeda* sp. Gram negative bacteria have a better resistance to anti-microbial compounds compared with gram-positive. (Branen and Davidson, 1983) states that gram-negative bacteria have a selection system against foreign substances at the lipopolysaccharide layer (Davidson et al., 2005). While (Pelczar and Chan, 1986) states positive gram bacterial cell wall structure is relatively simpler making it easier for antimicrobial compounds to enter the cell and find a target to work. The structure of the cell wall of gram-negative bacteria are relatively more complex, triple layers namely the outer layer in the form of lipoproteins, the middle layer in the form of lipopolysaccharide and peptidoglycan layer.

The phytochemical test results showed *H.renschi* and *H. gracillis* extracts contain steroid and saponins compounds, while alkaloids, terpenoids, tannins and flavonoids are not contained in the extract. In accordance with the phytochemical test results that containing steroids compound, then it is consistent with the NMR test results which found the active compounds in the form of β -sitosterol in the n-hexane solvent. β -sitosterol included in one type of steroid. In addition to the discovery of β -sitosterol, the extract was also found that the oleat acid compound is part of primary metabolite (Hendri, 2015; Anam, 1999; Shanab, 2007).

The phytochemical test results on other research state that extracts of *Caulerpa* sp, *Euchema* sp, *Gracilaria* sp and *Sargassum* sp contain alkaloids, flavonoids, steroids, triterpenoids and tannins (Siregar et al., 2012). Other phytochemical test results state that bioactive steroid compounds always found in a variety of phytochemical test (Siregar et al., 2012; Alamsyah et al., 2014), whereas the saponin compound in (Siregar et al., 2012) study is not found at all of the four seaweed extract tested, whereas in the study of (Firdaus., 2008; Alamsyah et al., 2014), saponins can be found.

(Kolanjinathan et al., 2009) reported the discovery of several compounds that are bioactive metabolites derived from several types of seaweed that is; brominated, aromatic, nitrogen-heterocyclic, sterols, protein, and polysaccharide sulfate. Results of another study states that *Sargassum* sp has potential as an antioxidant. Specifically, this plant contains phlorotannin, a polyphenol that is not found in other plants or seaweed. These compounds have proven capable of inhibiting lipid peroxidation and free radical activity. *S. duplicatum* contain alkaloids, saponins, tannins, steroids and glycosides with phlorotannin levels from 9.2822 to 37.3693 mg/g. Retention time fraction extract: 0.97; 0.75, and 0.46, and efficiency of anti-radical is 11264.54 (Firdaus., 2008).

Conclusions

Results of the study of the effectiveness of extracts of *Halimeda* sp against pathogens is have antibacterial activity against bacteria of *S.typhi*, *S. aureus*, *E. coli* and *B. subtilis* and has effectiveness to decrease the amount of pathogenic bacteria. The phytochemical

test result showed steroid and saponins compounds. While the TLC test results indicate the potential of the compound, although not pure. *Halimeda* sp extracts have antimicrobial. However, further research is needed to determine the compounds that exist and chemical structure. Environmental and geographical factors need to be done to see the influence on the type and content of the active compound.

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