

## NEW PYRAN OF AN ENDOPHYTIC FUNGUS *Fusarium* sp. ISOLATED FROM THE LEAVES OF BROTOWALI (*Tinaspora crispa*)

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### ABSTRACT

Endophytes are microorganisms that reside asymptotically in the tissues of higher plants and are relatively unstudied and a promising source of novel organic natural metabolites exhibiting a variety of biological activities. As a part of our systematic search for new bioactive lead structures from endophytic, the endophytic fungus *Fusarium* sp. isolated from the leaves of Brotowali (*Tinaspora crispa*), was cultured for isolation of metabolite. The endophytic fungus was cultivated on 6 L of Potatos Dextose Broth (PDB) medium at room temperature (no shaking) for 8 weeks. The cultures were then extracted with ethyl acetate to afford 9.4 g of residue after removal of the solvent under reduced pressure. The extract was separated into the fractions by column chromatography (CC) on silica gel. The fractions were further separated by silica gel column chromatography to give one compound. The molecular structure was established on the basis of spectroscopic analysis including UV, IR, <sup>1</sup>H-NMR, <sup>13</sup>C-NMR, HMQC, HMBC, COSY, and MS. The compound was determined as a new pyran.

**Keywords:** new pyran; endophytic fungus; *Fusarium* sp.; *Tinaspora crispa*

### ABSTRAK

Jamur endofitik merupakan mikroorganisme yang menempati jaringan tumbuhan tingkat tinggi dan masih terbatas kajiannya, serta merupakan sumber metabolit baru yang menunjukkan berbagai aktivitas biologis. Sebagai bagian dari penelitian sistimatis kami untuk menemukan senyawa bioaktif baru, maka telah diisolasi jamur endofitik *Fusarium* sp. dari daun Brotowali (*Tinaspora crispa*), dan telah dikultur untuk mengisolasi metabolit sekundernya. Jamur endofit dikultur dalam 6 L medium Potatos Dextose Broth (PDB) pada suhu kamar (keadaan statis) selama 8 minggu. Kultur kemudian diekstraksi dengan etil asetat untuk mendapatkan 9,4 g ekstrak pekat setelah penguapan pelarut pada tekanan rendah. Ekstrak difraksinasi dengan kromatografi kolom (CC) pada silika gel. Fraksi selanjutnya dikromatografi kolom silika gel hingga diperoleh satu senyawa. Struktur molekulnya ditentukan berdasarkan analisis spektroskopi UV, IR, <sup>1</sup>H-NMR, <sup>13</sup>C-NMR, HMQC, HMBC, COSY, dan MS. Senyawa hasil isolasi merupakan senyawa baru golongan piran.

**Kata Kunci:** piran baru; jamur endofitik; *Fusarium* sp.; *Tinaspora crispa*

### INTRODUCTION

Endophytes are defined as fungi colonizing healthy tissue plant without overt symptoms or no apparent injury to the host. In the last decade, the exploration of secondary metabolites from endophytic fungi have proven that endophytes are a source of new compounds that have unique structures and a variety of biological activity such as anticancer, antimicrobial and other activities. The new compounds was potentially useful for human medicine, industrial, and agrochemical [1-2].

The selection of the host plant is important factor for the production of endophytic fungi and the isolation of

their bioactive secondary metabolites [2]. The strategi of plant selection to be a promising source of novel endophytes and their compound mainly on the basis of their unique environmental setting, ethnobotanical history (use by indigenous peoples), endemic, unusual longevity, and growing in areas of great biodiversity [3].

*Tinaspora crispa* (locally named "brotowali" in Indonesia) has been used by traditional folklore for antimalarial, antifilarial, and antipyretic [4-5]. Its stem has been used for various therapeutic purposes such as treatment for diabetes, hypertension, stimulation of appetite and protection from mosquito bites [6]. Moreover, *T. crispa* is used in the traditional medicinal

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practice of peoples living in Malaysia, Indonesia, and Thailand to treat ailments like fever, jaundice, hyperglycemia, wounds, intestinal worms, skin infections, tooth and stomach aches, coughs, asthma and pleurisy [7-8].

In our search for secondary metabolites of endophytic fungi, many bioactive and/or new compounds were isolated [4,9-11]. Elfita et al. [4] was reported the isolation of eight endophytic fungi from the stems and leaves of brotowali, but the fungus *Fusarium* sp. was not found in the study. In this paper we report the isolation and structural identification a new pyran from *Fusarium* sp. separated from the leaves of brotowali (*T. crispata*).

Several *Fusarium* species isolated as endophytes have been reported to produce metabolites with antimicrobial, anticancer, and other activities, and also could be potential sources of novel natural products with industrial and agrochemical potential. There is speculation among the researchers about their nature of association in plants, as most *Fusarium* species are commonly encountered in soil as saprophytes, pathogens and are also associated with serious invasive mycoses in immunocompromised patients [2,12].

Bashyal et al. (2010) found triclinonoic acid and tricindiol, two irregular sesquiterpenes from an endophytic strain of *Fusarium tricinctum*, a fungus endophytic in the root tissue of the Sonoran desert plant, *Rumex hymenosepalus* [13]. Wu et al. (2009) reported five compounds from endophytic fungus *Fusarium* sp. isolated from *Trewia nudiflora*. These compounds were including the three *ent*-trachylobane and two atisane-type diterpenoids [14].

## EXPERIMENTAL SECTION

### Materials

The leaves of brotowali were collected on March 2012 from the Indralaya, Ogan Ilir, South Sumatra. Material for isolation and cultivation of endophytic fungi using Oxoid: nutrient agar (NA), chloramphenicol, nutrient broth (NB), potato dextrose broth (PDB), and potato dextrose agar (PDA). Material for sterilization and separation by silica gel column chromatography using Merck: ethanol 70%, NaOCl, Si gel 60 (70-230 mesh.), thin layer chromatography (TLC), and silica gel 60 F<sub>254</sub>. Organic solvents (hexane, ethyl acetate, and methanol) were technical grade and distilled before use.

### Instrumentation

The apparatus in the research were counter colony, autoclave, incubator, water bath, microscope, magnetic hotplate, UV lamp, column chromatography and generally apparatus in organic and microbiology

laboratory, Fisher Johns Melting Point, UV-Cary Varian 100 Conc I-UV Visible Spectrometer, FTIR-Perkin Elmer-Spectrum One, TOF MS ES+4.20e4, NMR spectra were recorded at 500 MHz (<sup>1</sup>H) and 125 MHz (<sup>13</sup>C) on JEOL JNM ECA-500 spectrometer, UV light at 254 nm and 365 nm.

### Procedure

#### Isolation of endophytic fungi

The leaves sample was washed thoroughly in running tap water and air dried before it was processed. The materials were then surface sterilized by immersing them sequentially in 70% ethanol for 3 min and 0.5% NaOCl for 1 min and rinsed thoroughly with sterile distilled water. Then, with a sterile scalpel, outer tissues were removed and the inner tissues of 0.5 cm size were carefully dissected and placed on petri-plates containing potato dextrose agar medium (PDA) (200 g potato, 20 g dextrose, and 15 g agar in 1 L of H<sub>2</sub>O, supplemented with 100 mg/L of chloramphenicol to suppress bacterial growth). The plates were incubated at 25±2 °C until fungal growth appeared. The plant segments were observed once a day for the growth of endophytic fungi. Hyphal tips growing out the plated segments were immediately transferred into new PDA plates and then subcultured until pure cultures were obtained [15].

#### Identification of the endophyte

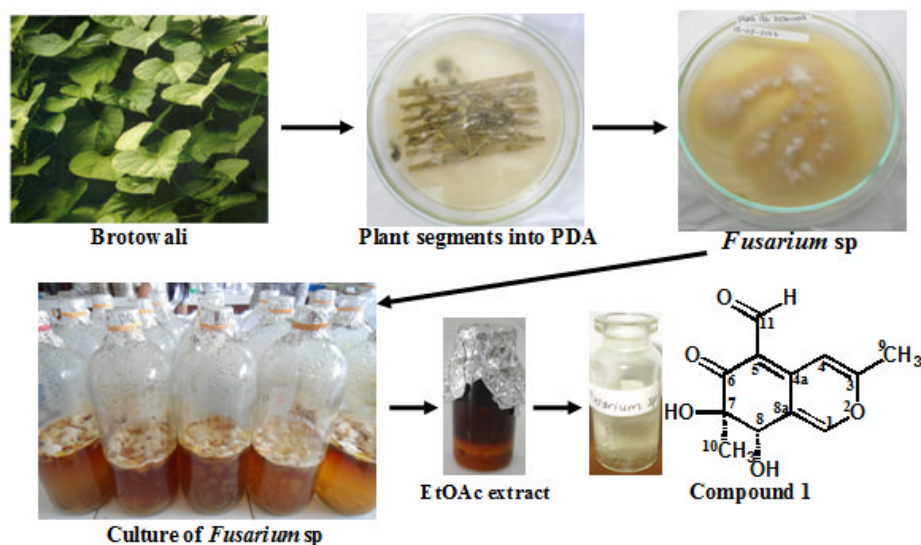
The endophytic fungal strain was identified by the morphological method. The morphological examination was performed by scrutinizing the fungal culture, the mechanism of spore production, and the characteristics of the spores. All experiments and observations were repeated at least twice [16].

#### Cultivation of pure fungal strain

The purified fungi (a small park) were transferred under sterile conditions to the PDB medium. For chemical investigations, the fungal strains were static cultivated into 15 flasks (1 L each) containing 400 mL of PDB medium for 8 weeks at room temperature [9-11].

#### Extraction, exploration, and structure elucidation

Fungi in the 8 weeks cultures were killed by addition of 50 mL of methanol (MeOH). The suspension was vacuum-filtered and the filtrate fractionated thrice by liquid-liquid partition with ethyl acetate (EtOAc) (1:1). Then the solvent phase was evaporated under reduced pressure using rotary vacuum evaporator at 40 °C to produce the ethyl acetate fraction of liquid cultures. The fraction was further separated by silica gel column chromatography



**Fig 1.** Isolation of the compound from ethyl acetate extract of *Fusarium sp.* from the leaves of brotowali

to give a pure compound.

The EtOAc fraction (9.4 g) was preabsorbed on silica gel and then chromatographed over a silica gel column using a gradient elution of hexane:EtOAc (10:0) to MeOH (100%). Based on detection by TLC using the eluent system, collected fractions were combined and rechromatographed over silica gel to give a pure compound. The molecular structure of compound was established on the basis of spectroscopic analysis including UV, IR, MS,  $^1\text{H-NMR}$ ,  $^{13}\text{C-NMR}$ , HMQC, HMBC, and COSY.

## RESULT AND DISCUSSION

The fungal strain was identified as *Fusarium sp.* by the Sekolah Ilmu dan Teknologi Hayati, Institute Teknologi Bandung, Indonesia. High colonization of the genus *Fusarium* were obtained from several plant species such as *Mirabilis jalapa*, *Cephalotaxux mannii*, *Sesbania grandiflora*, family *Araceae*, *Leguminosae* and *Gramineae*. This suggests allopatric existence of endophytic *Fusarium* based on different geographical regions and ecological niches and their ubiquity as endophyte among plant species [15, 17-20].

Then fungal *Fusarium sp.* was cultivated on 6 L of PDB medium for 8 weeks at room temperature. The culture was then extracted with ethyl acetate to afford 9.4 g of residue after removal of the solvent under reduced pressure. The extract (9.4 g) was separated over silica gel column (CC, 230–70 mesh, 30 × 4 cm) using a gradient of hexane:EtOAc (10:0) to MeOH (100%) to provide 6 fractions. Fraction 5 (3.7 g) was rechromatographed over silica gel (CC, 230–80 mesh, 30 × 3 cm) and using a gradient of EtOAc:MeOH (5:5) to MeOH (100%) to yield 3 fractions. Fraction 3 (2.9 g) was

rechromatographed again on silica gel (CC, 230–80 mesh, 30 × 2 cm) using the same gradient elution to yield compound **1** (2.3 g). The isolation of the compound from ethyl acetate extract of *Fusarium sp.* from the leaves of brotowali is described in Fig. 1.

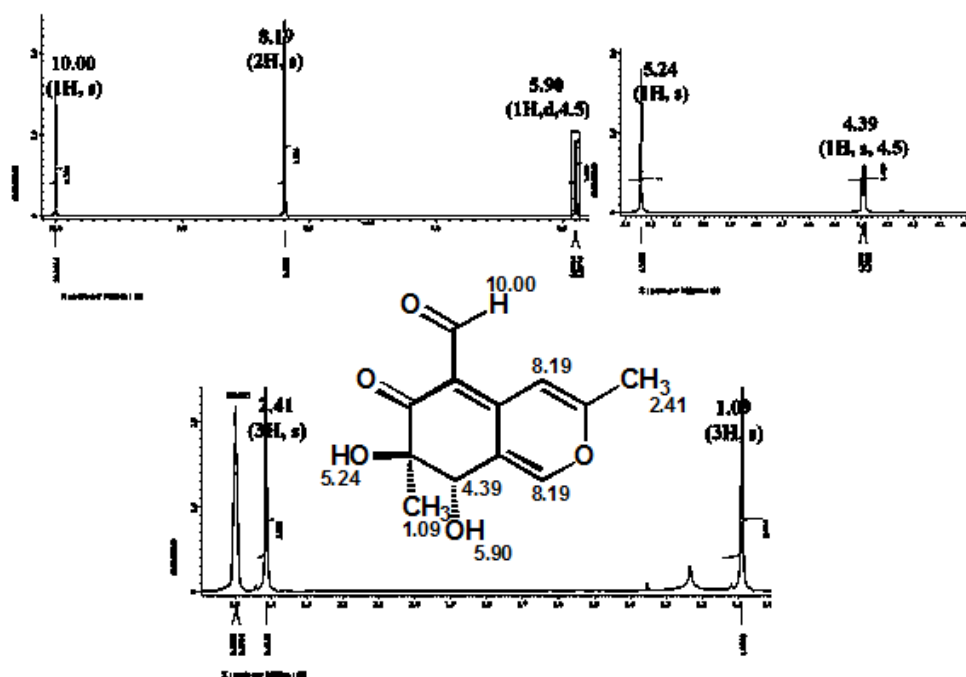
(7R,8S)-7,8-dihydroxy-3,7-dimethyl-6-oxo-7,8-dihydro-6H-isochromene-5-carbaldehyde (**1**), obtained as a white crystal, mp. 230-231 °C; UV (MeOH)  $\lambda_{\text{max}}$  nm: 255 and 381; UV (MeOH + NaOH)  $\lambda_{\text{max}}$  nm: 255 and 381; IR (KBr)  $\nu_{\text{max}}$   $\text{cm}^{-1}$ : 3469.9 and 3375.4 (OH), 3099.6 (CH-vinilic), 2978.1 (CH-aliphatic), 1678.1 (C=O), 1608.3 and 1477.5 (C=C conjugation), 1043.5 (C-O ether); HRESI-MS ion peak at  $m/z$  237.0762  $[\text{M} + \text{H}]^+$ ;  $^1\text{H-NMR}$  (DMSO, 500 MHz)  $\delta_{\text{H}}$  ppm: see Table 1 and Fig. 2;  $^{13}\text{C-NMR}$  (DMSO, 125 MHz)  $\delta_{\text{C}}$  ppm: see Table 1.

The HRESI-MS showed an  $[\text{M} + \text{H}]^+$  ion peak at  $m/z$  237.0762 (calc. 237.0759), in accord with the molecular formula  $\text{C}_{12}\text{H}_{12}\text{O}_5$ , implying seven degrees of unsaturation, and was supported by NMR 1D and 2D data (Table 1). The  $^1\text{H-NMR}$  spectrum, which was recorded in DMSO (Table 1) showed two hydroxy proton signal at  $\delta_{\text{H}}$  5.24 (1H,s) and 5.90 (1H,d,4.5 Hz), two methyl singlets ( $\delta_{\text{H}}$  1.09 and 2.41), two vinilic proton signals which appeared at the same chemical shift at  $\delta_{\text{H}}$  8.19 (1H,s), and one aldehyde proton signal at  $\delta_{\text{H}}$  10.00 (1H,s).

The  $^{13}\text{C-NMR}$  and DEPT 135 spectrum, and was supported by HMQC spectrum (Fig. 3) showed 12 signals consisting of two methyl, four methines, and six quaternary carbons. Two of these signals were assigned to carbon atoms bearing hydroxyl groups, one tertiary at  $\delta_{\text{C}}$  70.6 and one quaternary at  $\delta_{\text{C}}$  74.3. Two of these signals are the carbonyl carbon (aldehyde at 198.2 and ketone at 189.3). Six carbon, resonating

**Table 1.** The NMR data of compound **1**, recorded at  $^1\text{H}$ -500 MHz;  $^{13}\text{C}$ -125 MHz in DMSO

No. C	$\delta_{\text{C}}$ ppm	DEPT 135	$\delta_{\text{H}}$ (ppm), $\Sigma\text{H}$ , multiplicity, J (Hz)	HMBC	COSY
1	151.8	CH	8.19 (1H,s)	70.6; 108.2; 122.2	
3	166.2	C			
4	106.3	CH	8.19 (1H,s)	148.6; 166.2; 19.9; 108.2	
4a	108.2	C			
5	148.6	C			
6	198.2	C			
7	74.3	C			
8	70.6	CH	4.39 (1H,d,4.5)	122.2; 151.8; 18.8; 74.3	5.90
8a	122.2	C			
9	19.9	$\text{CH}_3$	2.41 (3H,s)	166.2; 106.3	
10	18.8	$\text{CH}_3$	1.09 (3H,s)	198.2; 70.6; 74.3	
11	189.3	CH	10.00 (1H,s)	148.6; 108.2	
7-OH			5.24 (1H,s)	198.2; 70.6; 74.3	
8-OH			5.90 (1H,d,4.5)	122.2	4.39

**Fig 2.** The  $^1\text{H}$ -NMR spectrum of compound **1**

at  $\delta_{\text{C}}$  151.8 ( $\delta_{\text{H}}$  8.19, 1H, s), 122.2, 166.2, 106.3 ( $\delta_{\text{H}}$  8.19, 1H, s), 108.2, and 148.6, indicated the presence of three double bonds at  $\Delta^{1,9}$ ,  $\Delta^{3,4}$ , and  $\Delta^{5,10}$ . The coupling constant between H-8 and 8-OH ( $J=4.5$  Hz) indicated H-8 $\beta$ , 8-OH $\alpha$  configurations. The presence of cyclic ethers is shown by the presence of carbon and proton chemical shift at higher field at  $\delta_{\text{C}}$  151.8 ( $\delta_{\text{H}}$  8.19) and quaternary carbon at 166.2.

The HMBC correlations of isolated methyl protons H-10 with the oxygenated quaternary carbon C-7, the oxymethine carbon C-8, and the ketone carbon C-6 indicated that C-7 was connected to C-6, C-8, and C-10. Correlation from H-8 to C-8a, C-1, C-10, and C-7 led to the connection of C-8a to both C-1 and C-8, permitted

completion of the cyclohexenone subunit of **1**. Further HMBC correlation from H-1 to C-3, C-8, C-4a, and C-8, and correlation from H-4 to C-5, C-1, C-9, and C-4a indicated that C-3 and C-4a was connected to C-1 and C-4 that have the same chemical shift. Correlation of the isolated methyl protons H-9 with the oxygenated quaternary carbon C-3 and the methine carbon C-4, thereby completing the pyran subunit of **1**. Then the correlation of the aldehyde proton H-11 with C-5 established the connectivity between C-4 and C-5. The HMBC correlation and  $\delta$ -assignment of compound **1** showed Fig. 4 and 5.

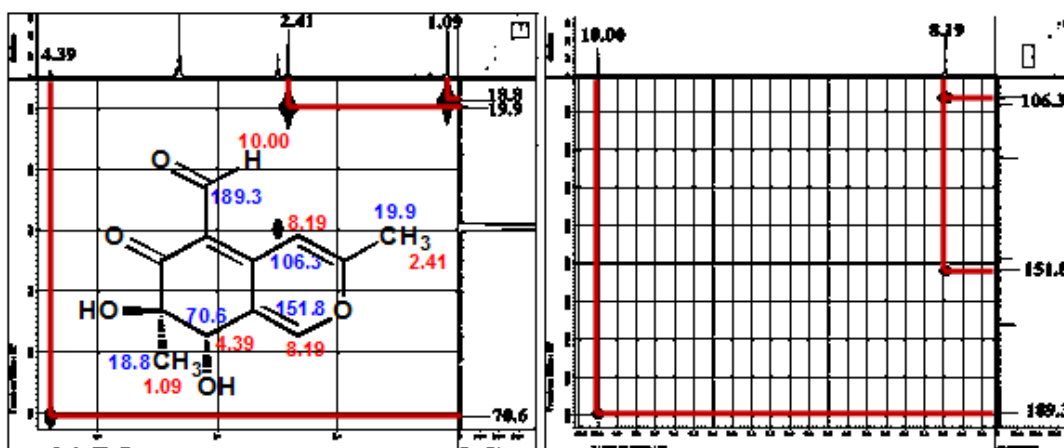


Fig 3. The HMQC correlation and  $\delta$ -assignment of compound 1

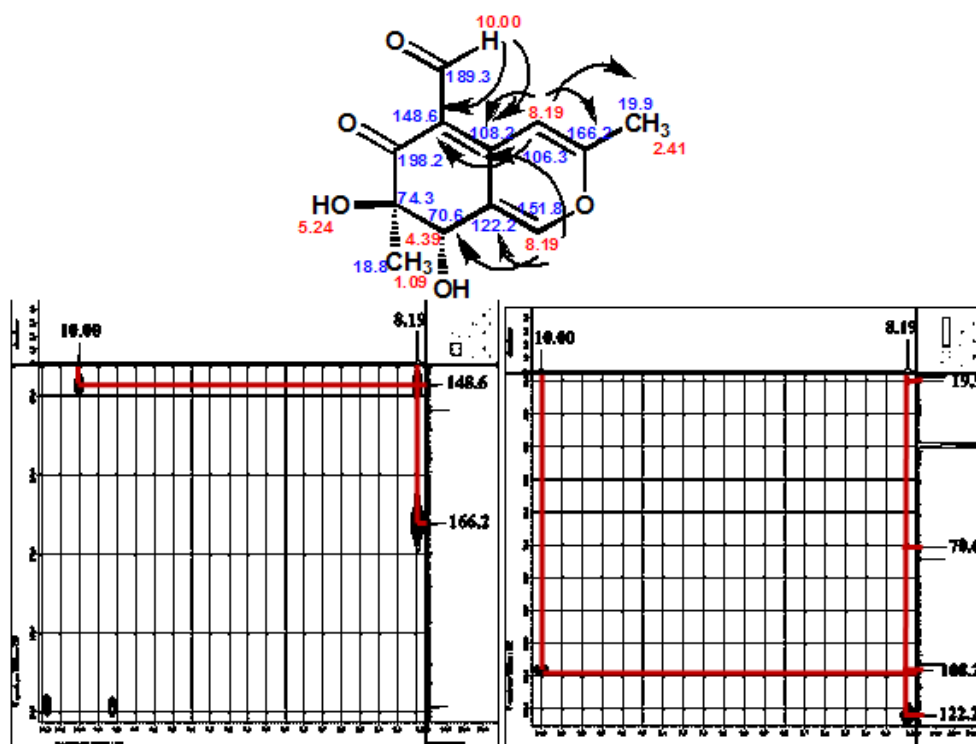


Fig 4. The HMBC correlation of two vinylic and aldehyde proton of 1

Analysis of  $^1\text{H}$ - $^1\text{H}$  COSY spectrum (Fig. 6) led to the identification of two proton spin-system corresponding to the C-8-8-OH subunit of structure 1. All of the data showed that compound 1 has two rings that cyclohexenone and pyran has been called benzopyran.

Benzopyran is a very interesting class of heterocyclic compounds that results from the fusion of a benzene ring to a heterocyclic pyran ring. According to IUPAC nomenclature it is called chromene. Two classes of benzopyrans are 1-benzopyran and 2-benzopyran that vary by the orientation of the fusion of

the two rings compared to the oxygen. The 1-benzopyran has been called chromene and 2-benzopyran called as isochromene.

Base on Dictionary Natural Products data base (2010), compound 1 is new pyran. Previously, the similar pyran compounds have been found which has a ring of 7,8-dihydro-6H-isochromen-6-one that monapurfluor A and its isomers is monapurfluor B from red mold rice fermented by *Monascus purpureus* NTU 568 [21], and with monasfluore B from red mold rice fermented by *Monascus pilosus* BCRC 38 072 [22]. They are both alkylated at C-3 and C-7, and

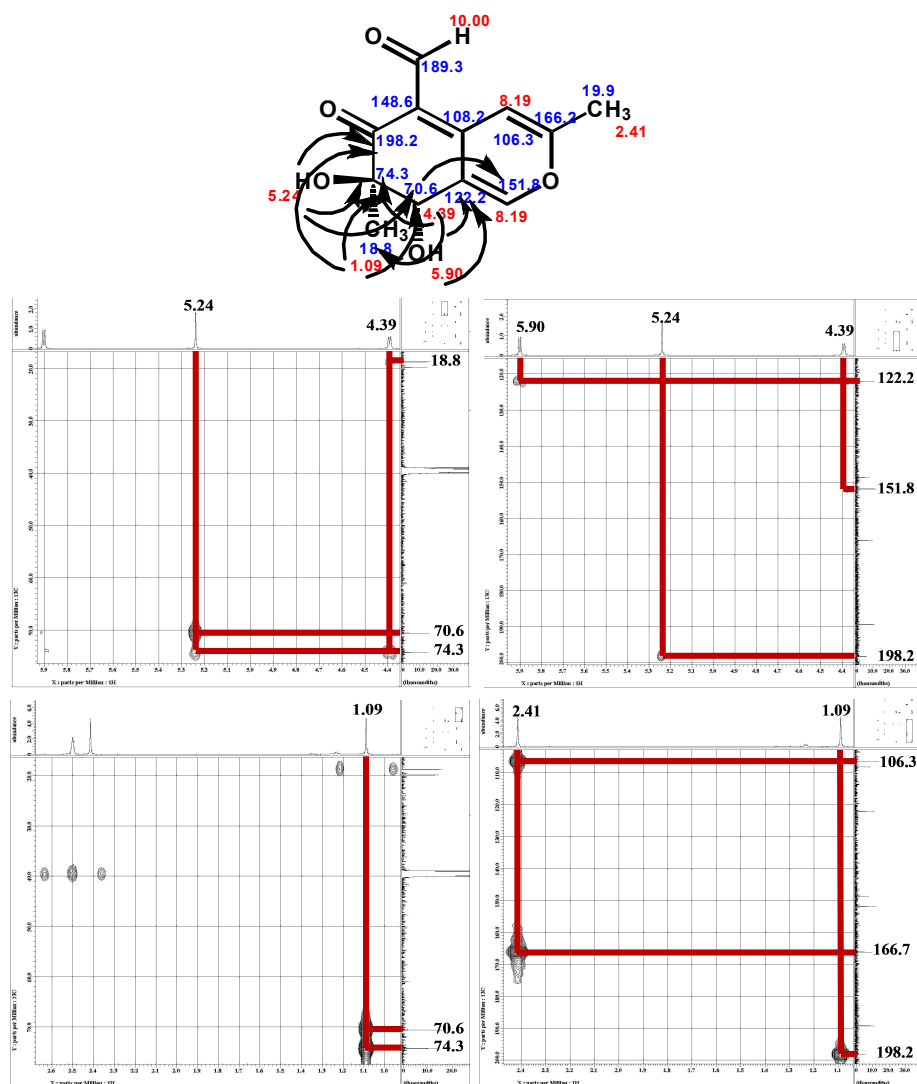


Fig 5. The HMBC correlation of hydroxyl, methyl, and methine proton of 1

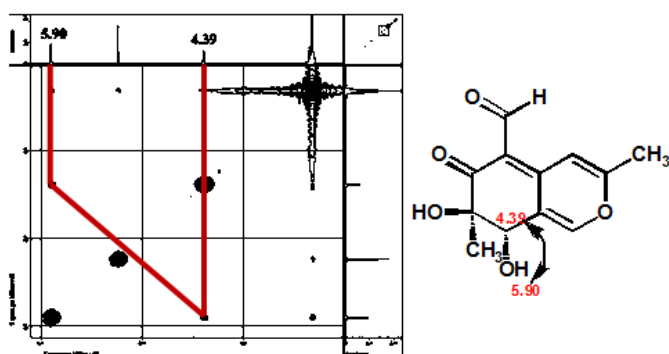


Fig 6. The COSY correlation of 1

oxygenated at C-7. The difference is that compound 1 is substituted carbaldehyde group at C-5 and hydroxyl group at C-8, while others are not. The other three compounds cyclized at C-7 and C-8, respectively with a

furan ring to monapurfluor A and B, and the furanone ring for monasfluor B.

Exploration of the pyran has increased the types of secondary metabolites from endophytic fungi of brotowali. Exploration of secondary metabolites research needs to be done in order to get the profile of organic compounds produced by endophytic fungi of brotowali.

## CONCLUSION

The endophytic fungi *Fusarium* sp. from the leaves of brotowali (*T. crispata*), has produced a new pyran. Based on spectroscopic analysis including UV, IR, MS,  $^1\text{H-NMR}$ ,  $^{13}\text{C-NMR}$ , HMQC, HMBC and COSY, the compound is (7R,8S)-7,8-dihydroxy-3,7-dimethyl-6-oxo-7,8-dihydro-6H-isochromene-5-carbaldehyde.

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